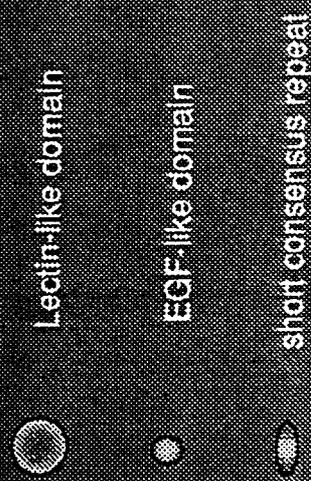
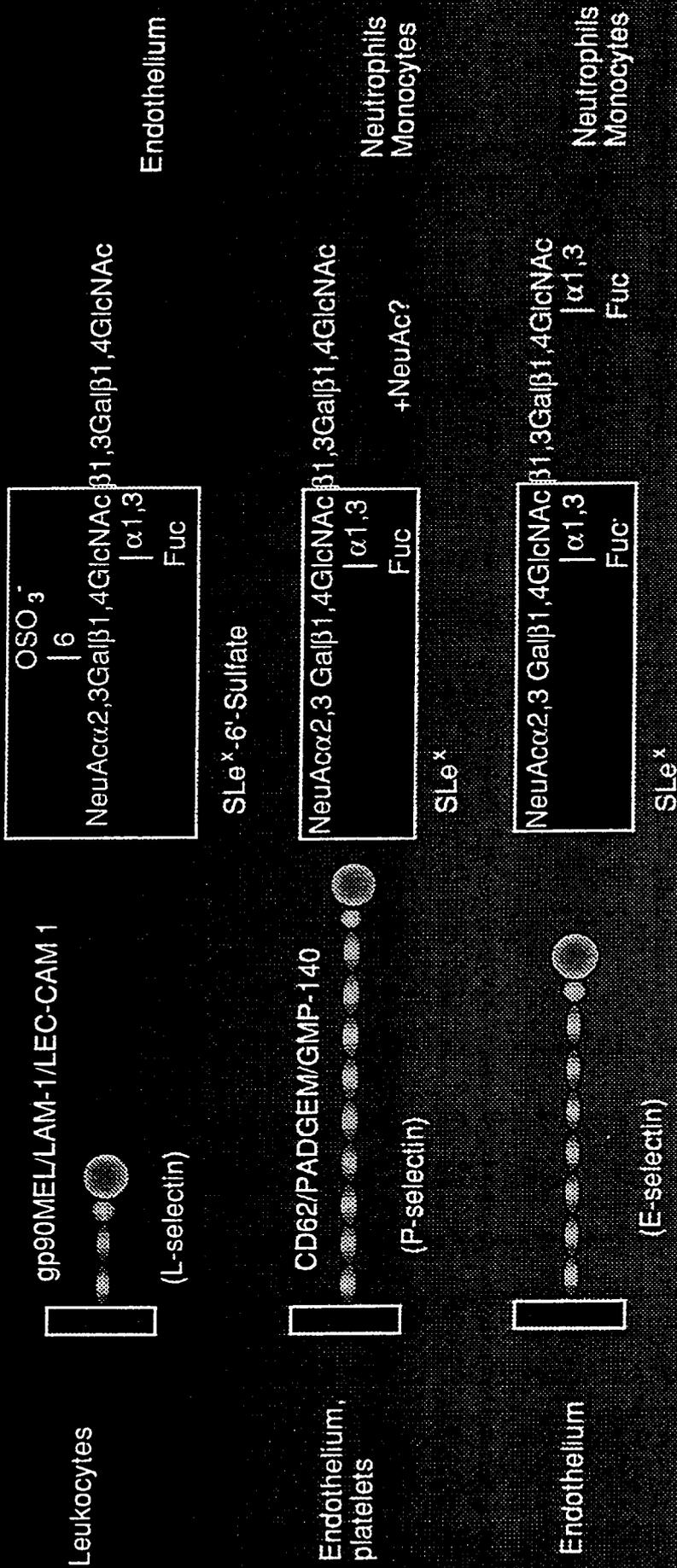
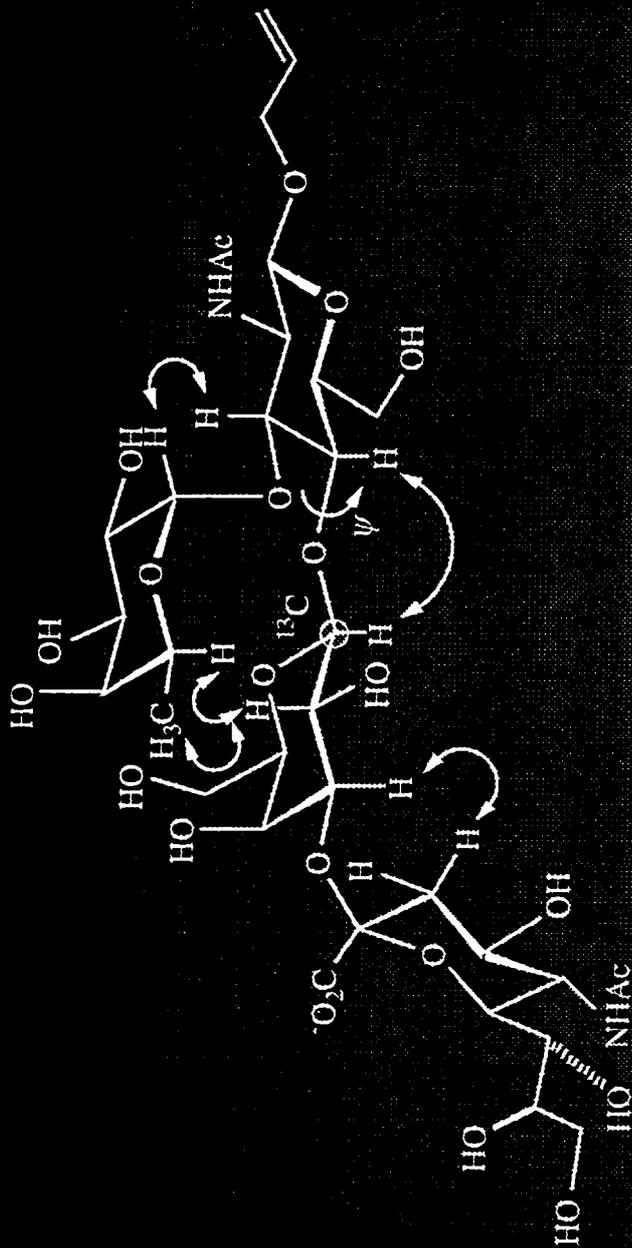


Carbohydrate-Binding Proteins (Selectins) and their Ligands



Springer, T. A.; Lasky, L. A. *Nature* 1991, 349, 196.
 Hemmerich, S.; Rosen, S. D. *Biochemistry* 1994, 33, 4830



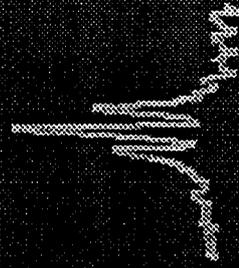
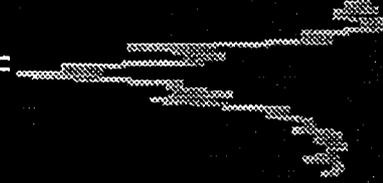
Calculated $\psi = 15^\circ$

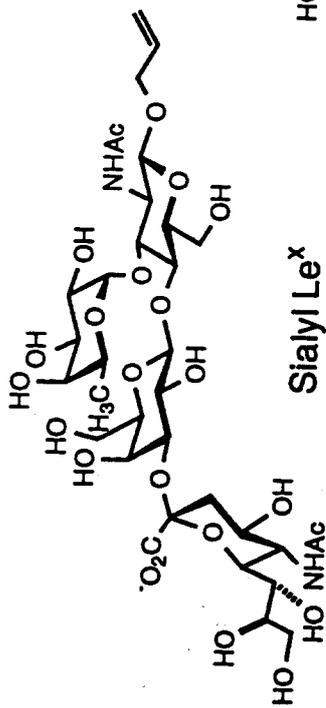
$${}^3J_{C-H} = 0.5 - 0.6\cos\psi + 5.7\cos^2\psi$$

$${}^3J_{C-H} = 5 \text{ Hz} \longrightarrow \psi = 18^\circ \text{ for Gal-GlcNAc}$$

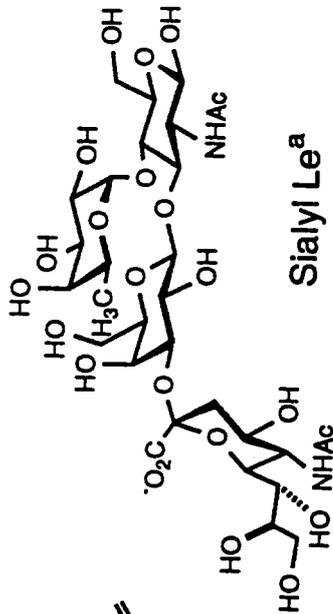
${}^3J_{C-H}$

←

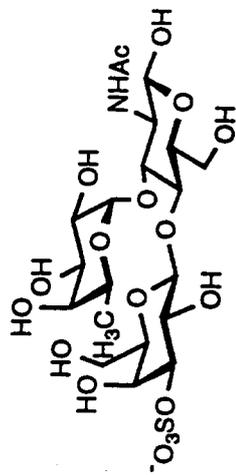




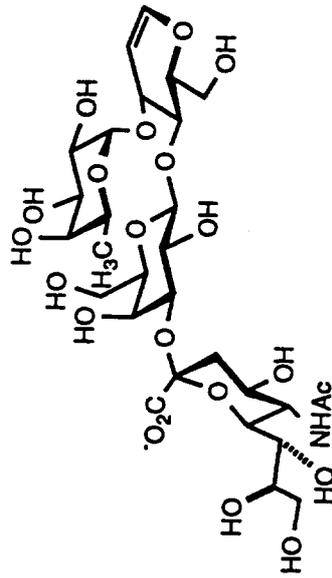
IC₅₀ = 2.1 mM
1.0 μM (*in vivo*)
JACS 1992, 114, 9283.



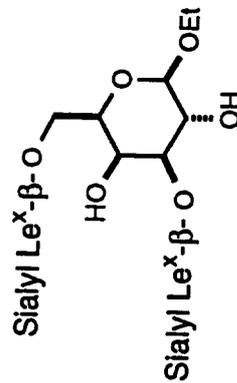
IC₅₀ = 2.1 mM
JACS 1992, 114, 9283.
PNAS 1991, 88, 10372.



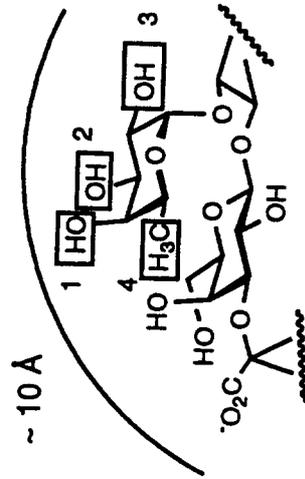
IC₅₀ = 2.1 mM
Biochemistry 1992, 31, 9126



IC₅₀ = 2.1 mM
JACS 1992, 114, 9283.

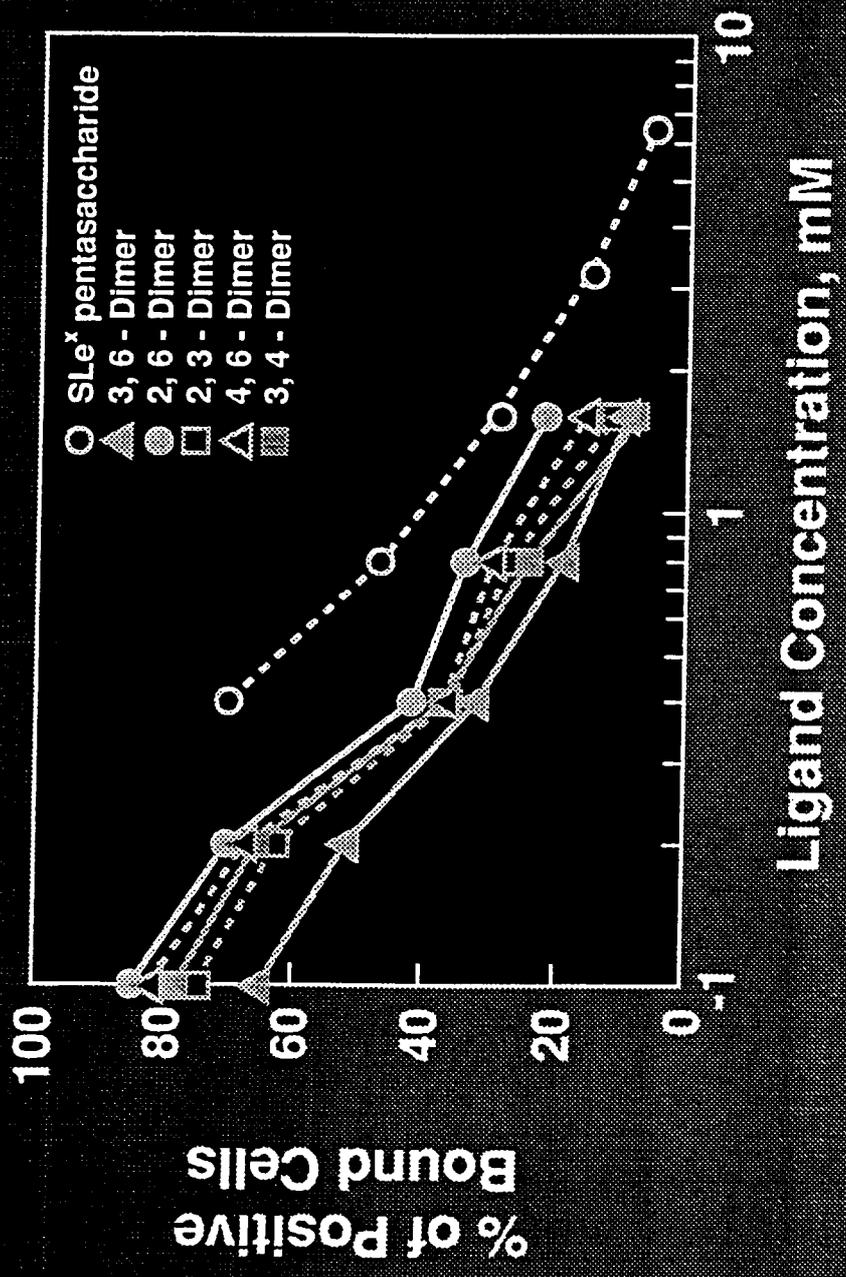


IC₅₀ = 0.4 mM
JACS 1993, 115, 7549.

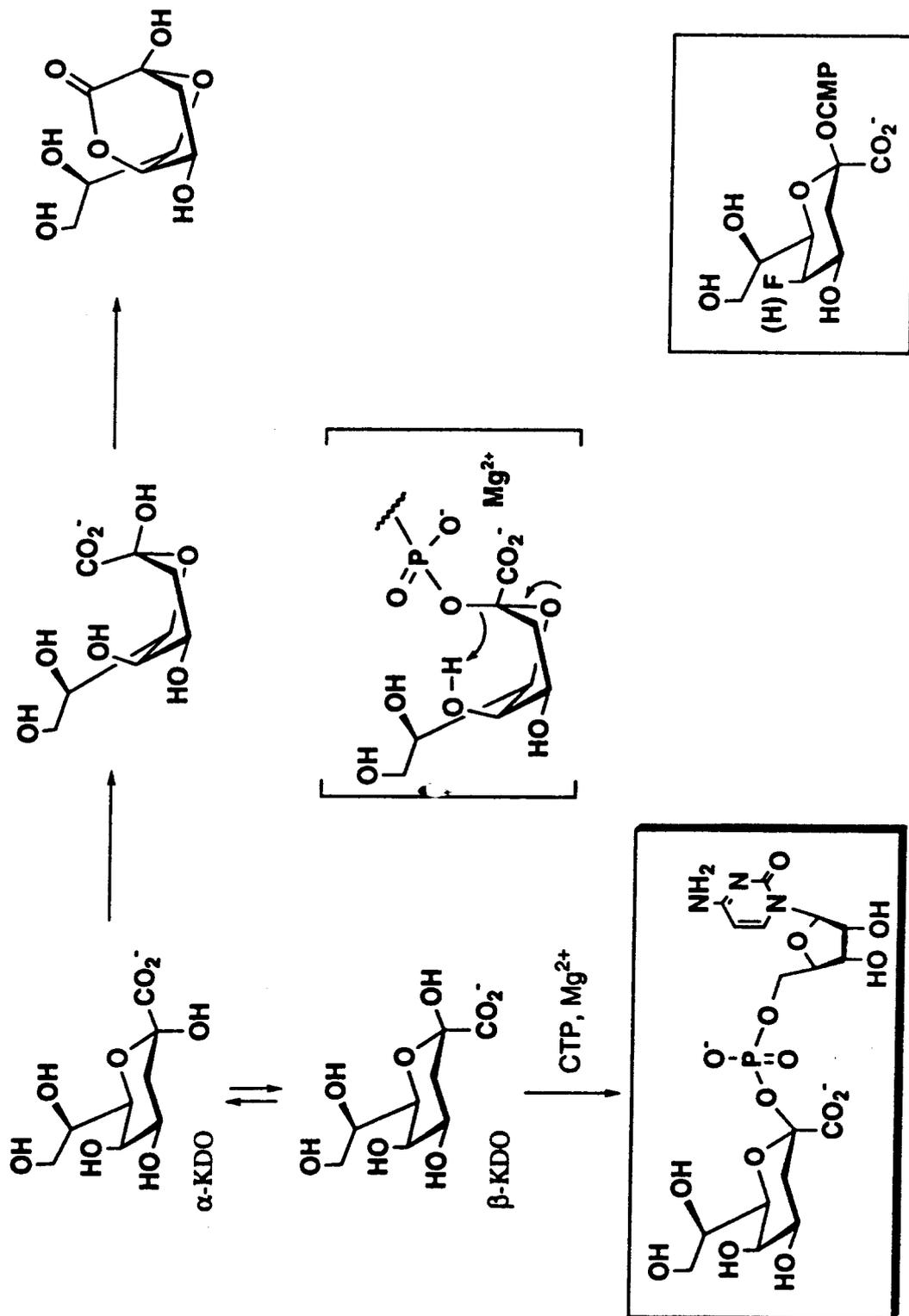


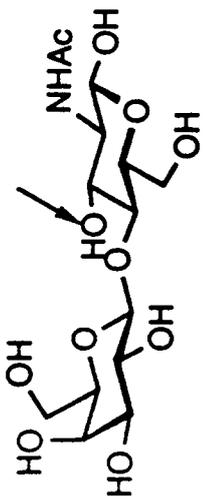
JACS 1992, 114, 9283.

Inhibition of HL-60 Adhesion to E-Selectin Coated Plates



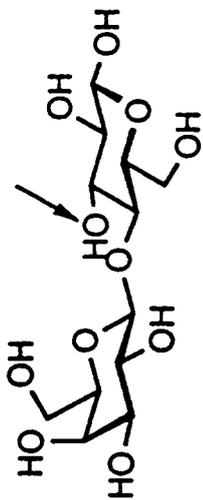
Stabilities of KDO and CMP-KDO





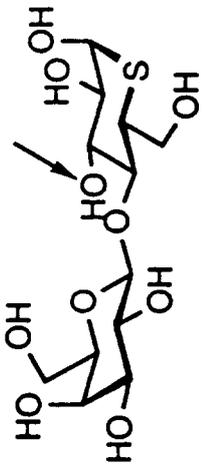
Galβ1,4GlcNAc

$K_m = 35 \text{ mM}$, $V = 100$



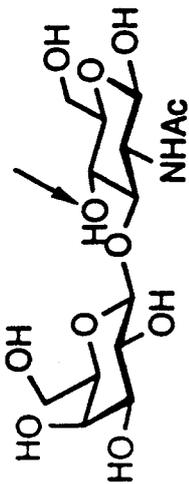
Galβ1,4Glc

$K_m = 500 \text{ mM}$, $V = 160$



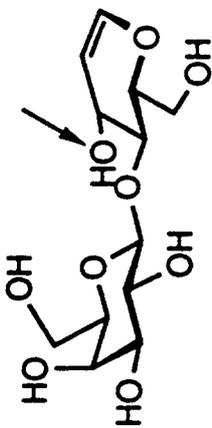
Galβ1,4(5-S-Glc)

$K_m = 12 \text{ mM}$, $V = 51$



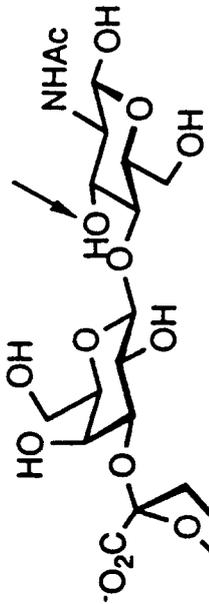
Galβ1,3GlcNAc

$K_m = 600 \text{ mM}$, $V = 130$



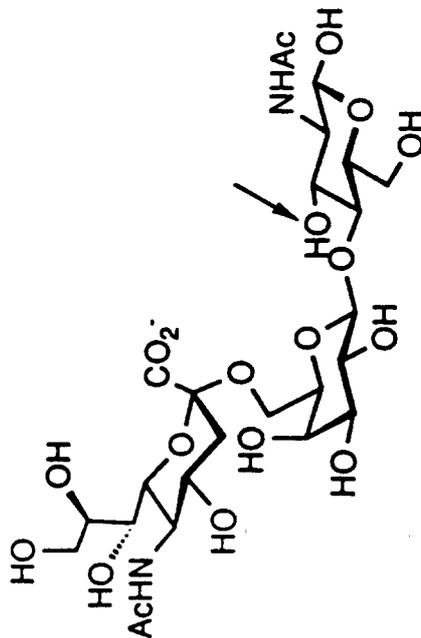
Galβ1,4Glucal

$K_m = 34 \text{ mM}$, $V = 10$



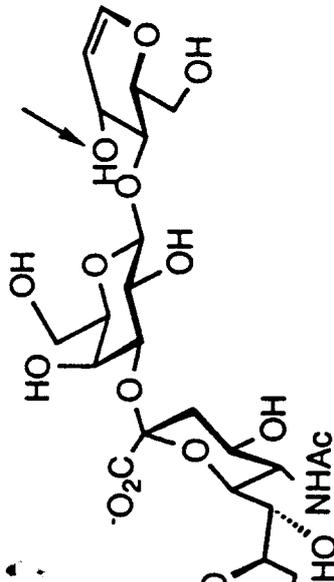
NeuAcα2,3Galβ1,4GlcNAc

$K_m = 100 \text{ mM}$, $V = 620$



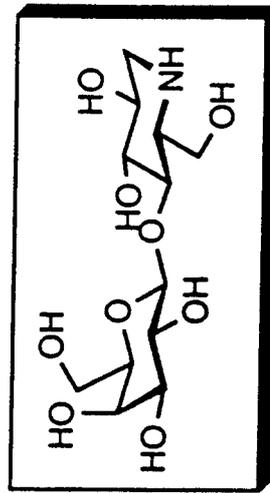
NeuAcα2,6Galβ1,4Glc

$K_m = 70 \text{ mM}$, $V = 13$



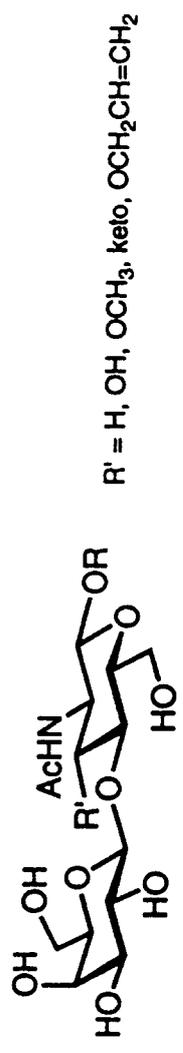
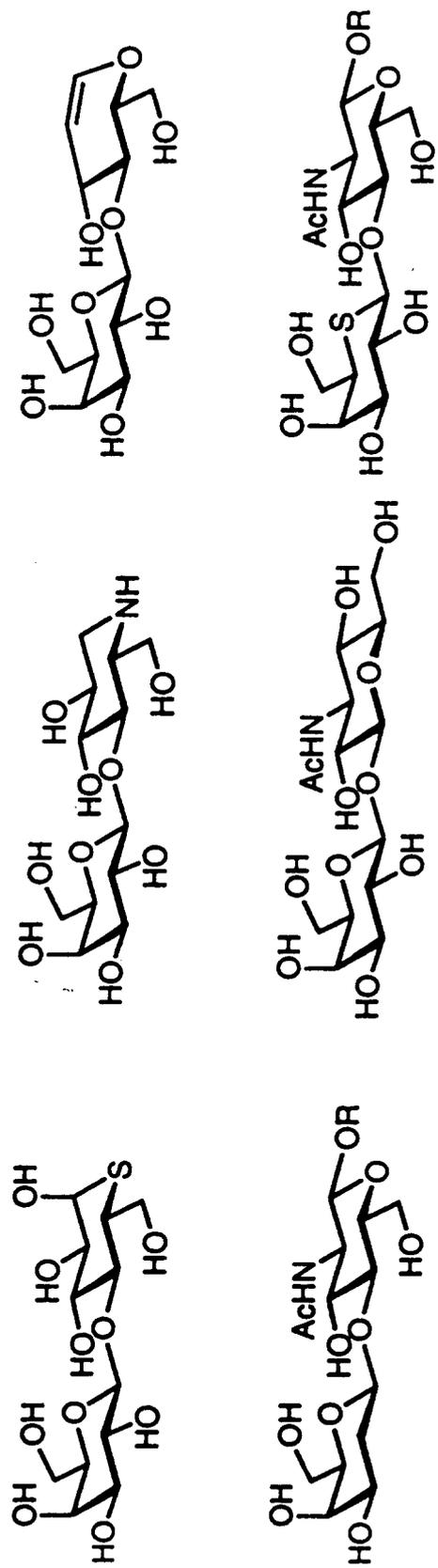
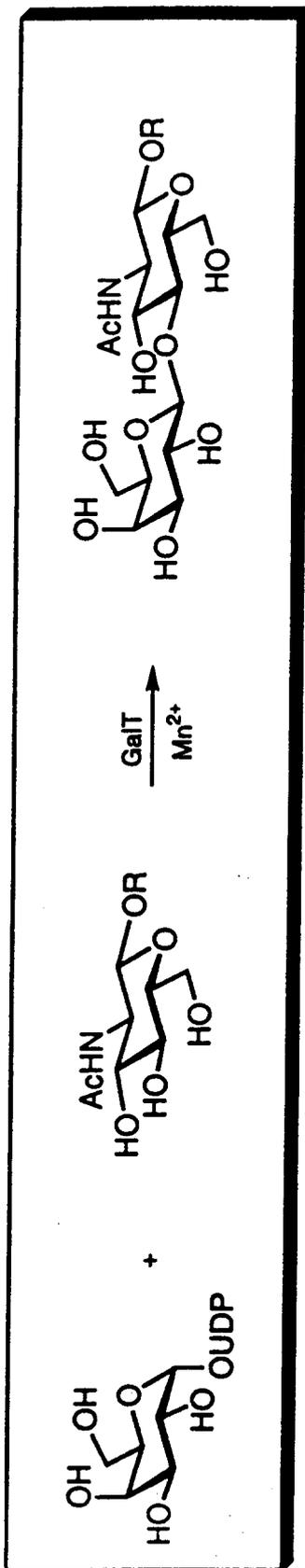
NeuAcα2,3Galβ1,4Glucal

$K_m = 64 \text{ mM}$, $V = 330$

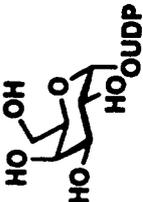
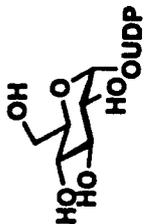
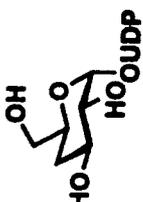
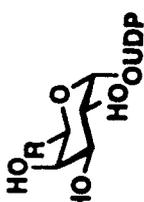


Galβ1,4deoxynojirimycin

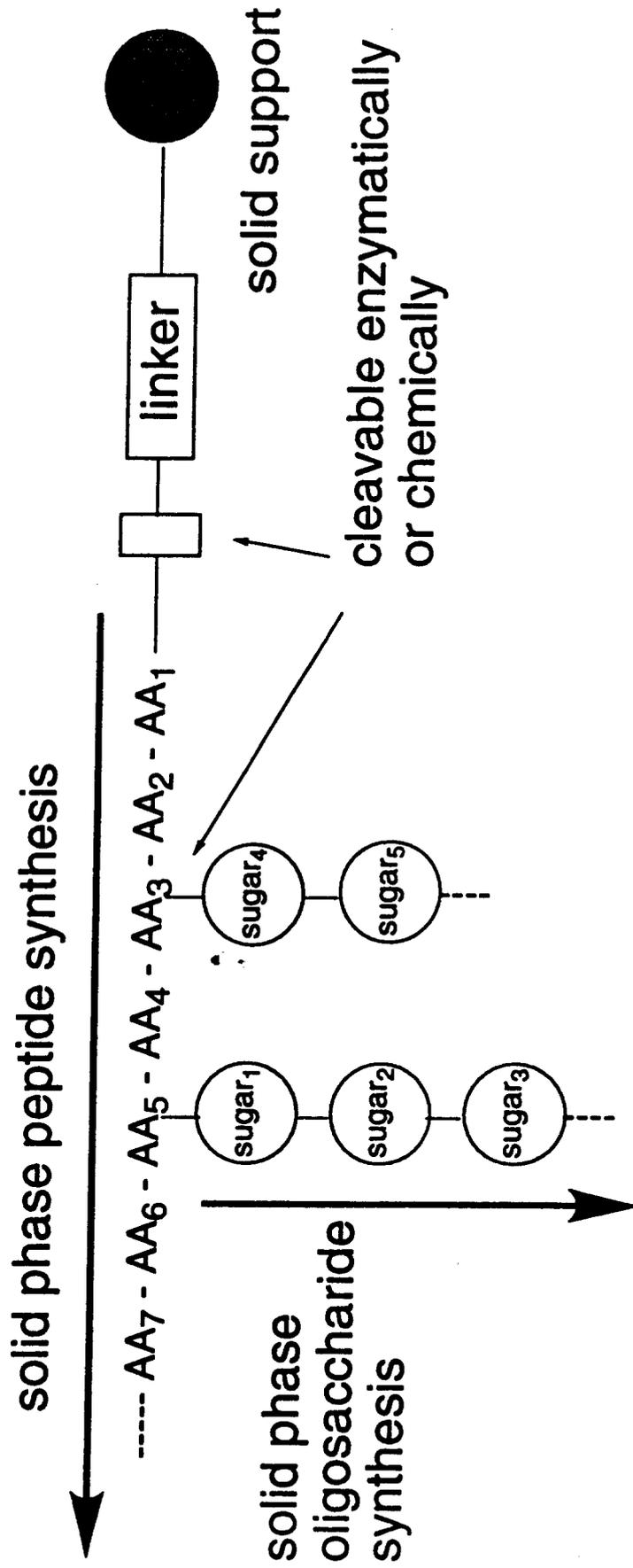
$IC_{50} = 8 \text{ mM}$



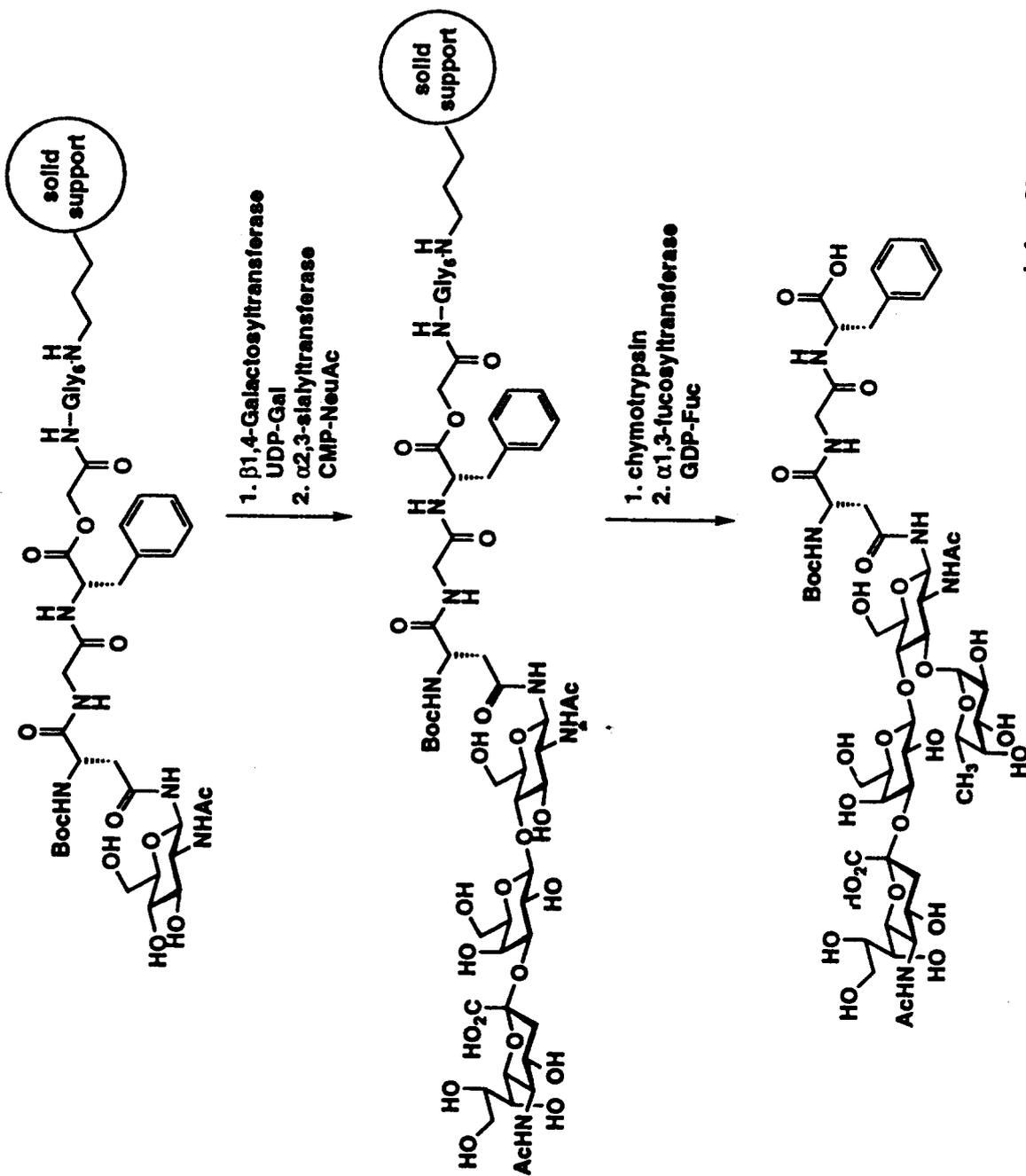
Donor Specificity of β -1,4-Galactosyltransferase

Rel. rate	Ref.	Chemical Structure	Rel. rate	Ref.
100	Berliner & Robinson 1982		0.00	Paicic & Hindsgaul 1991
0.3	Paicic & Hindsgaul 1991		0.09	Paicic & Hindsgaul 1991
5.5	Berliner & Robinson 1982		5.0	Yuasa 1992
4.0 1.3 0.2	Berliner & Robinson 1982		90	Srivastava & Hindsgaul 1993
4.0	Paicic & Hindsgaul 1991			

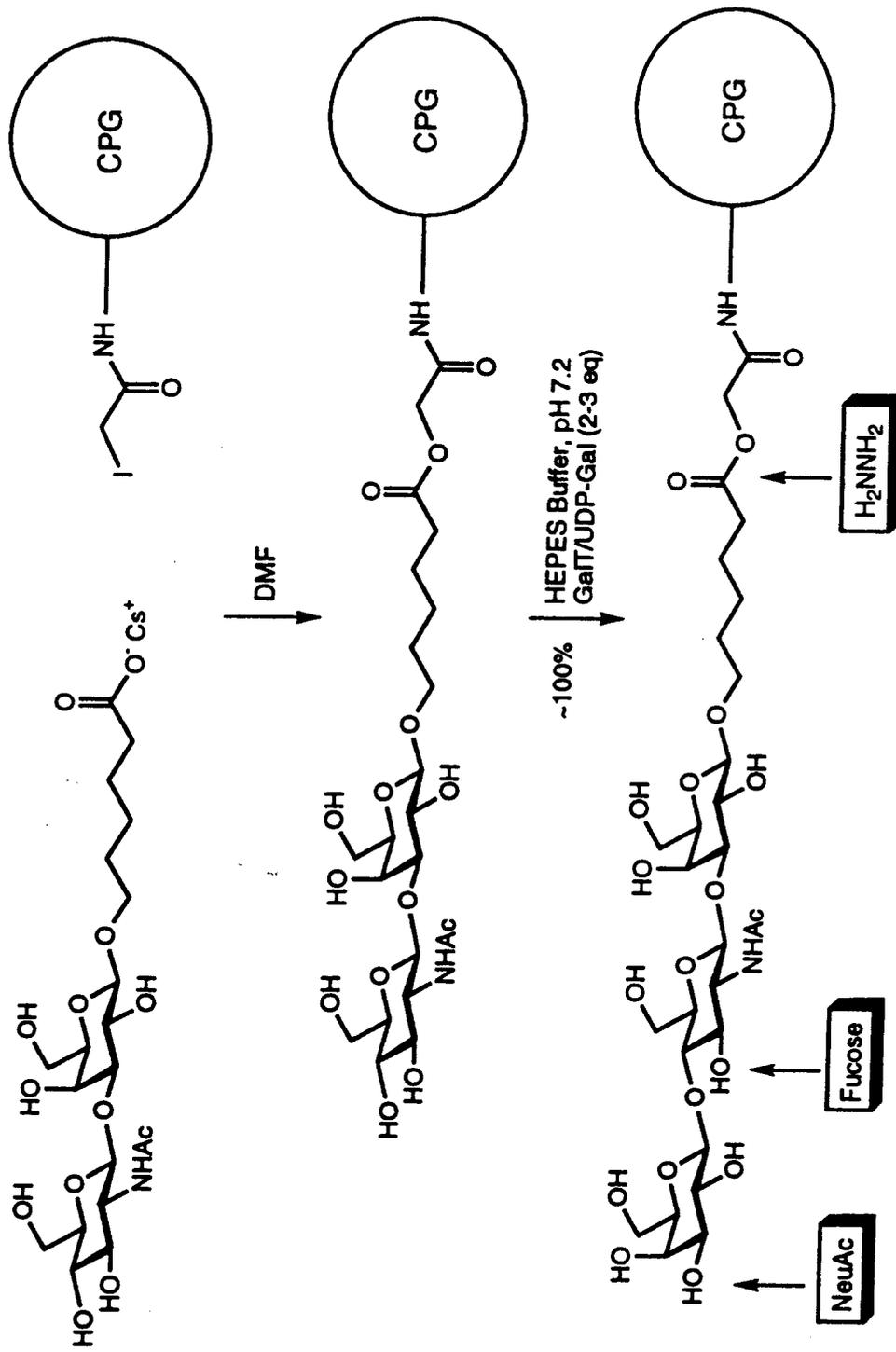
Solid phase synthesis of glycopeptides and oligosaccharides

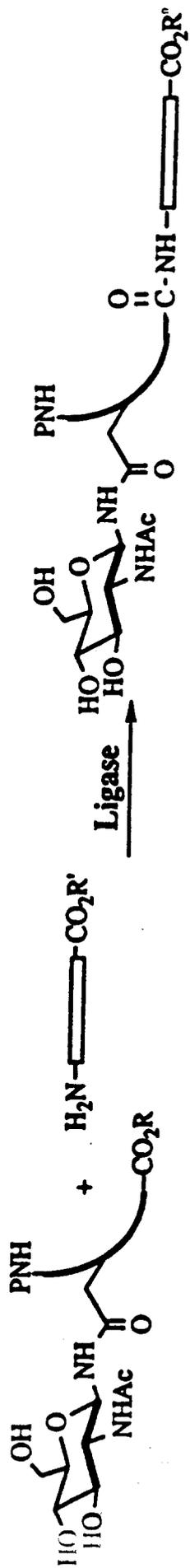


Solid-Phase Synthesis of Glycopeptides



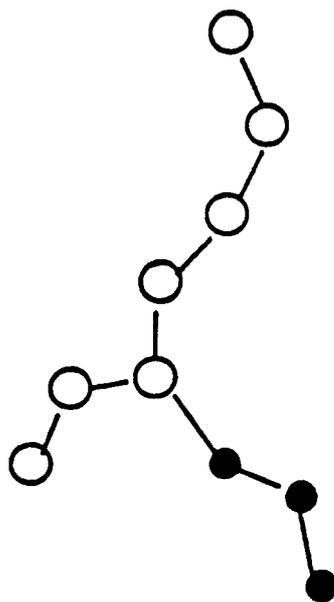
Solid-Phase Synthesis of Oligosaccharide





Glycosyltransferase

Ligase: N²-Me-His⁵⁷-Chymotrypsin
(Ser195 → Cys) Subtilisin

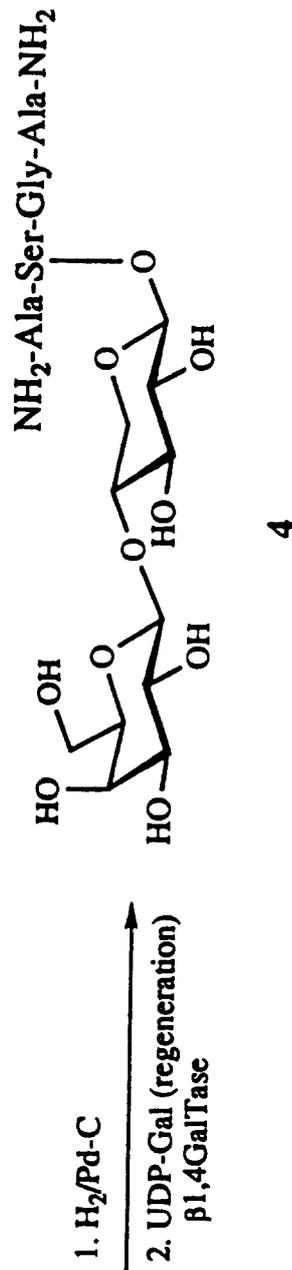
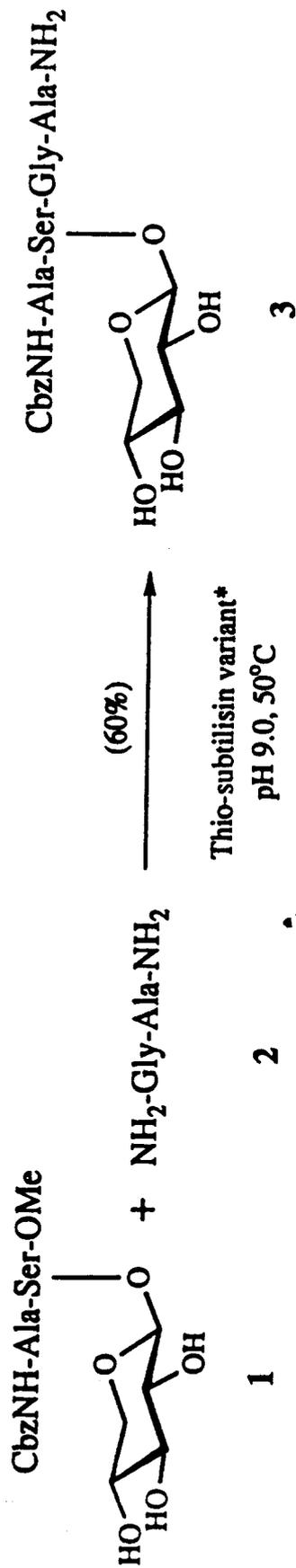


●: sugar

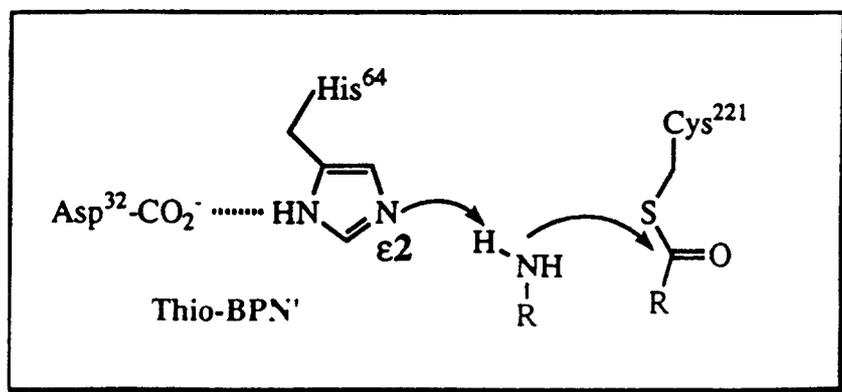
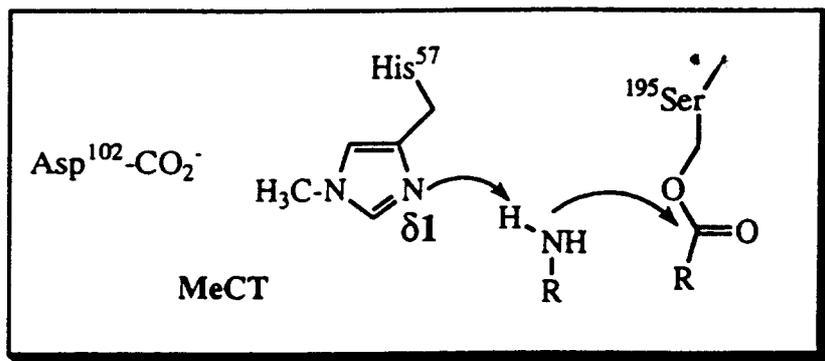
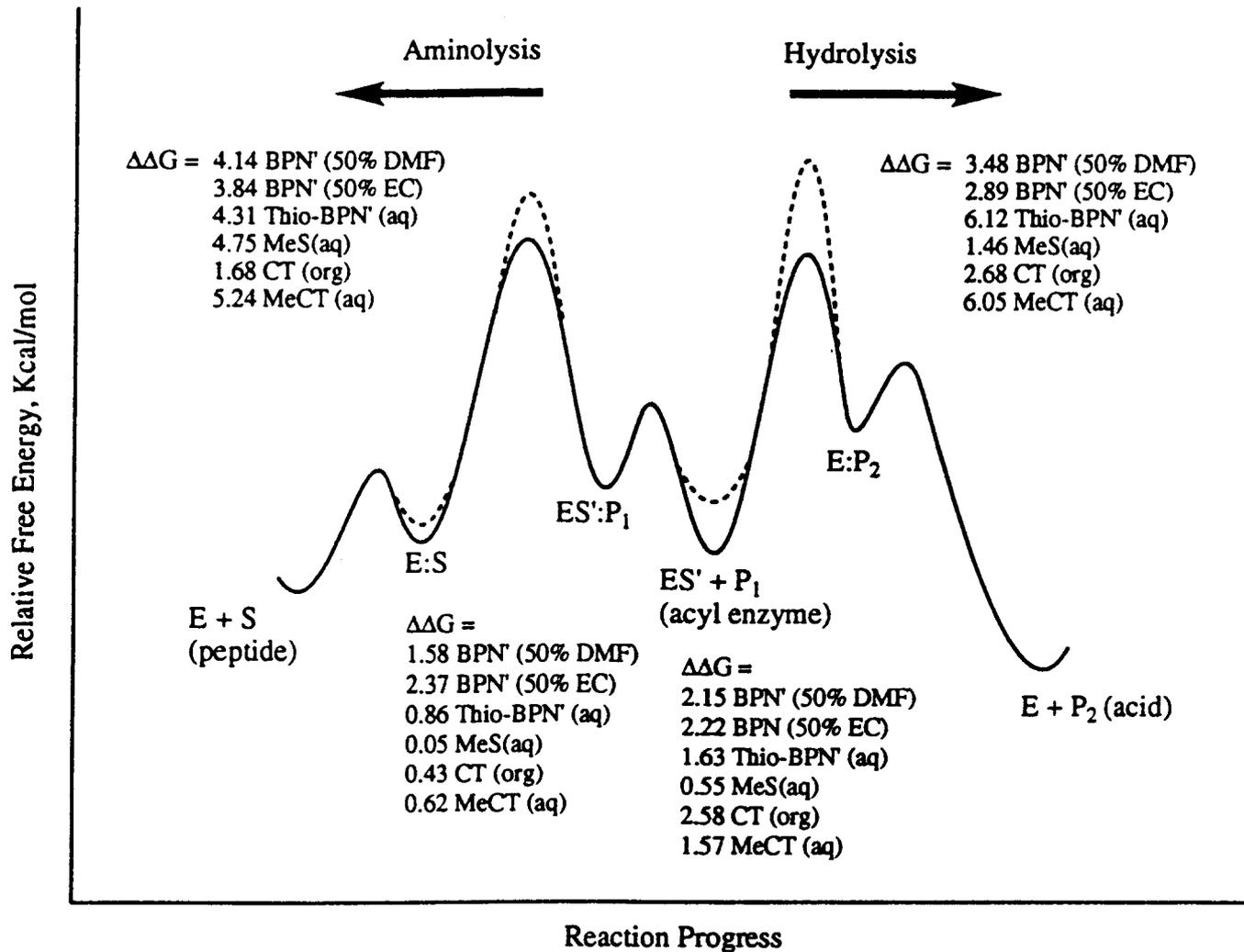
○: amino acid

Glycopeptides

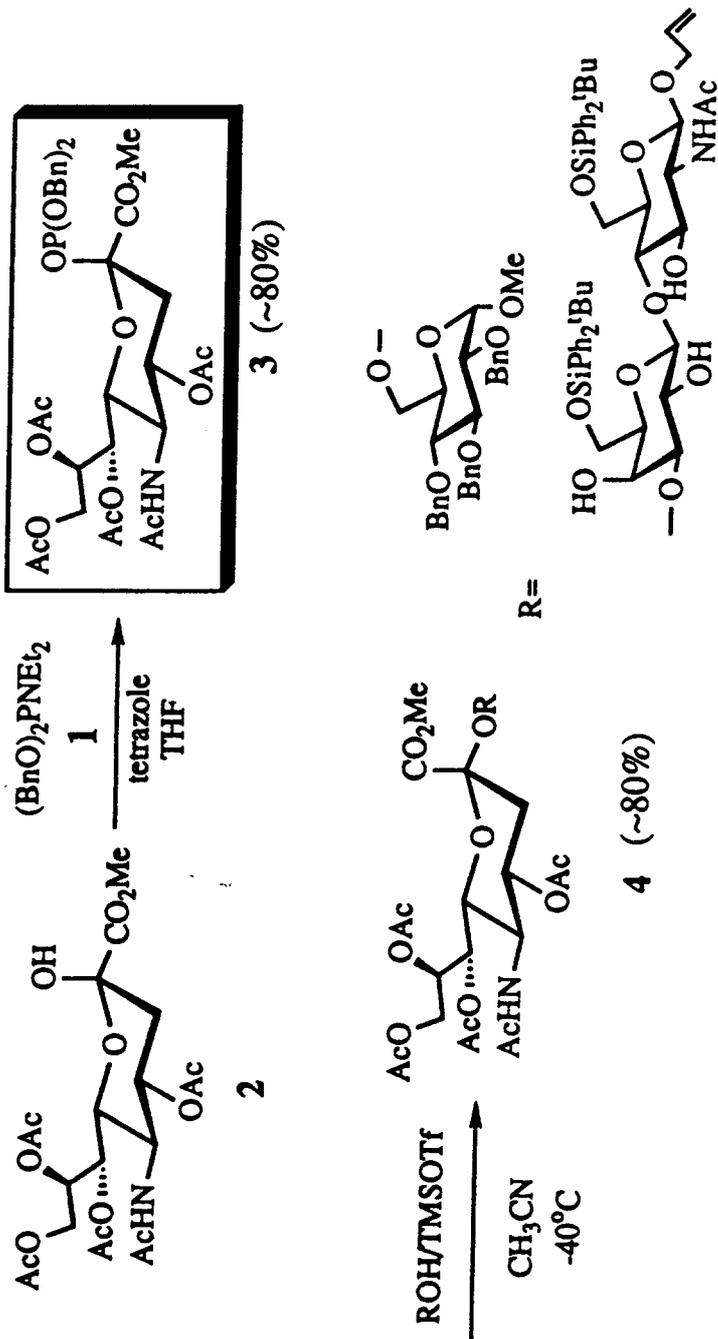
Enzymatic Synthesis of a Glycopeptide.



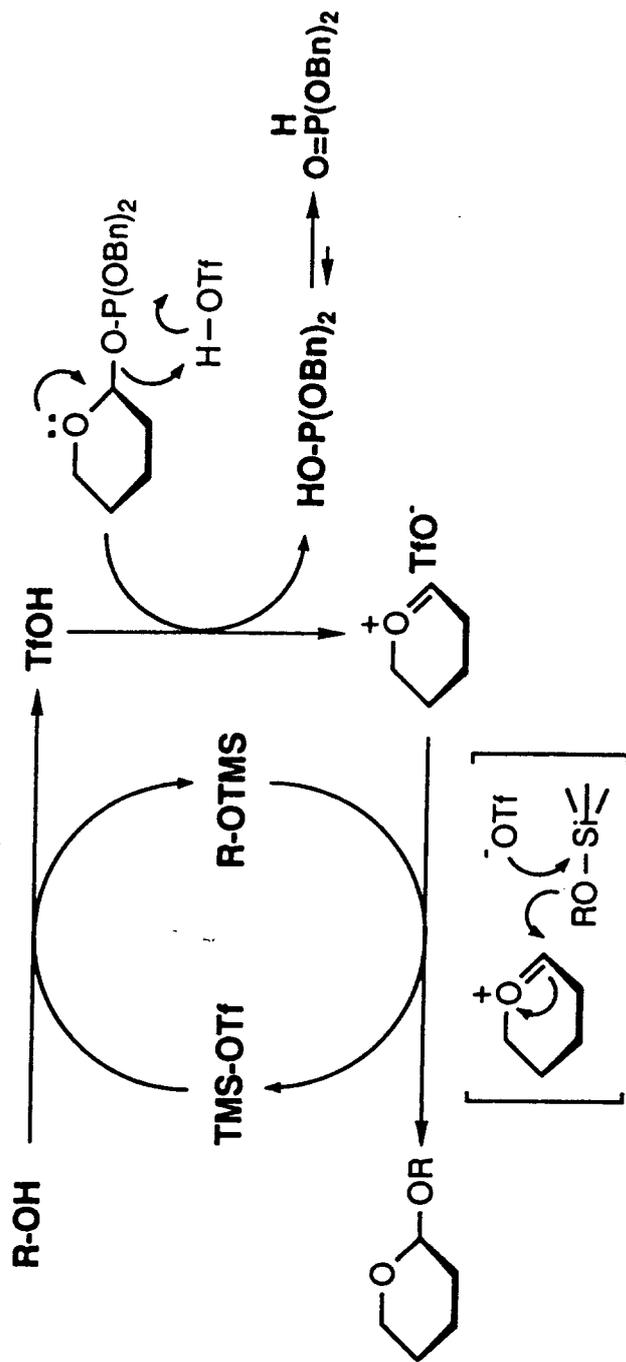
*Thio-subtilisin variant contains the following changes:
 Met50Phe, Asn76Asp, Gly169Ala, Asn218Ser, and Ser221Cys.



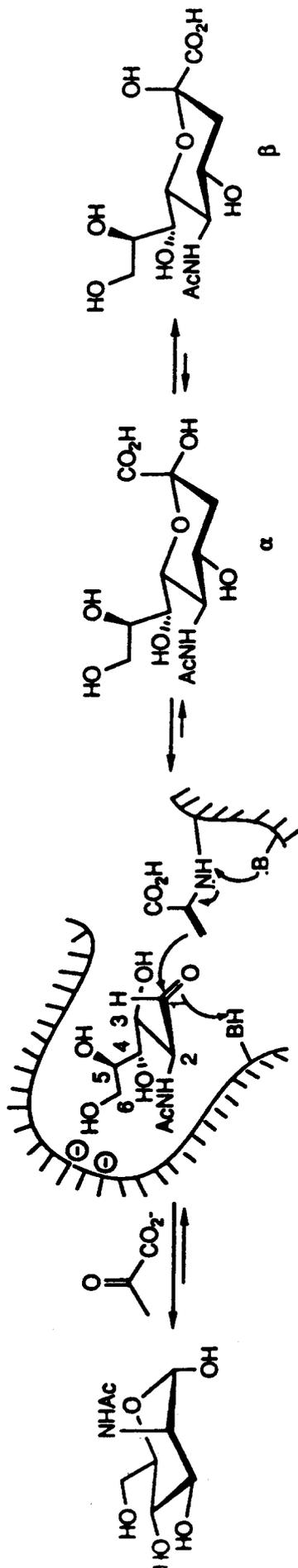
Chemical Sialylation Employing Sialyl Phosphate.



Proposed Mechanism of Glycophosphite-Mediated Glycosylation
 TMSOTf Reacts with Acceptor First



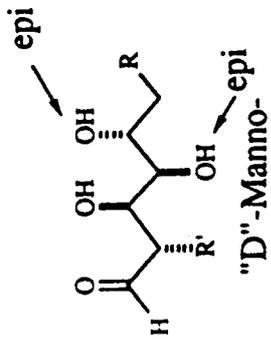
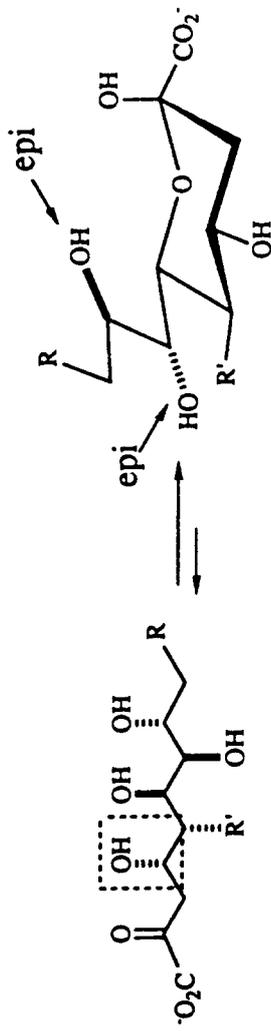
Sialic Acid Aldolase : Substrate Specificity



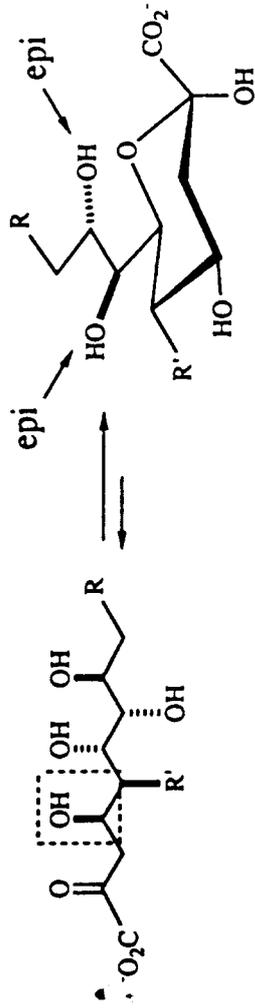
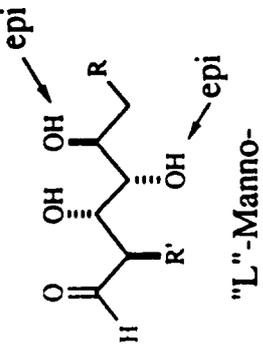
Acceptor Position	2	3	4	5	6
Very good	Ph		CH ₃ O	AcO	F
	HO		epi-F (6-F)	AcO	AcO
	H		AcO		CH ₃ CH(OH)CO ₂
	HOCH ₂ CONH		epi-OH (2-OH)		CH ₃ O
	N ₃		H (2-OH)		N ₃
Weak	epi-F	F (2 epi-OH)	F (2 epi-OH)		(CH ₃) ₂ P(O)O-
No	epi-OH	epi-OH	epi-OH (2 epi-OH)	CH ₃ O	deleted to C-5
	epi-NHAc				OPO ₃ ⁻

S David, *Tetrahedron*, 46, 201 (1990)

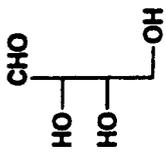
Kim and Wong, *JACS*, 110, 6481 (1988)
Liu, Ichikawa, Wong, *JACS*, 1992



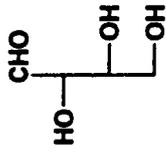
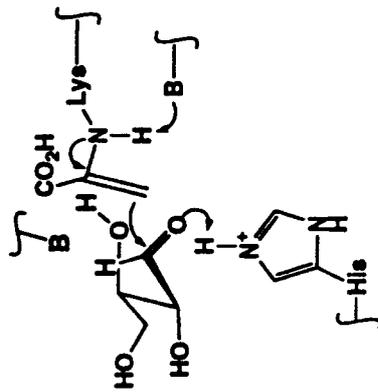
NeuAc aldolase



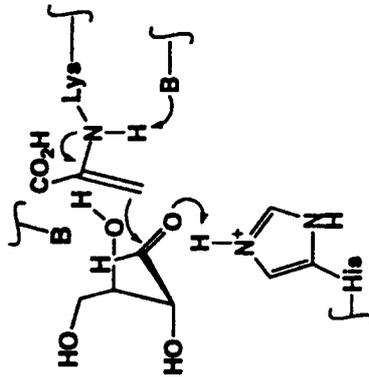
R = OH, H R' = OH, AcNH, N₃, H



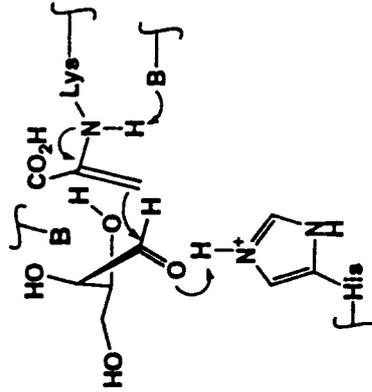
si attack



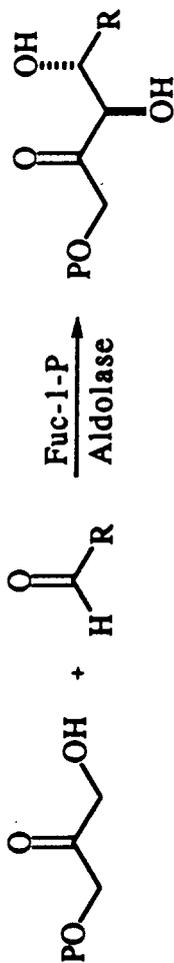
si attack



re attack



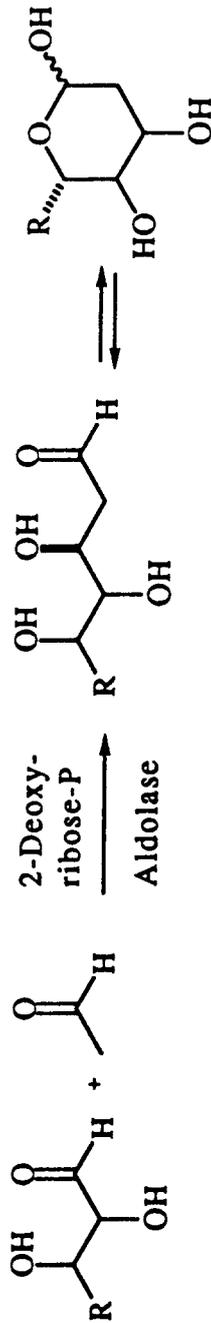
Synthesis of Fucose Analogs

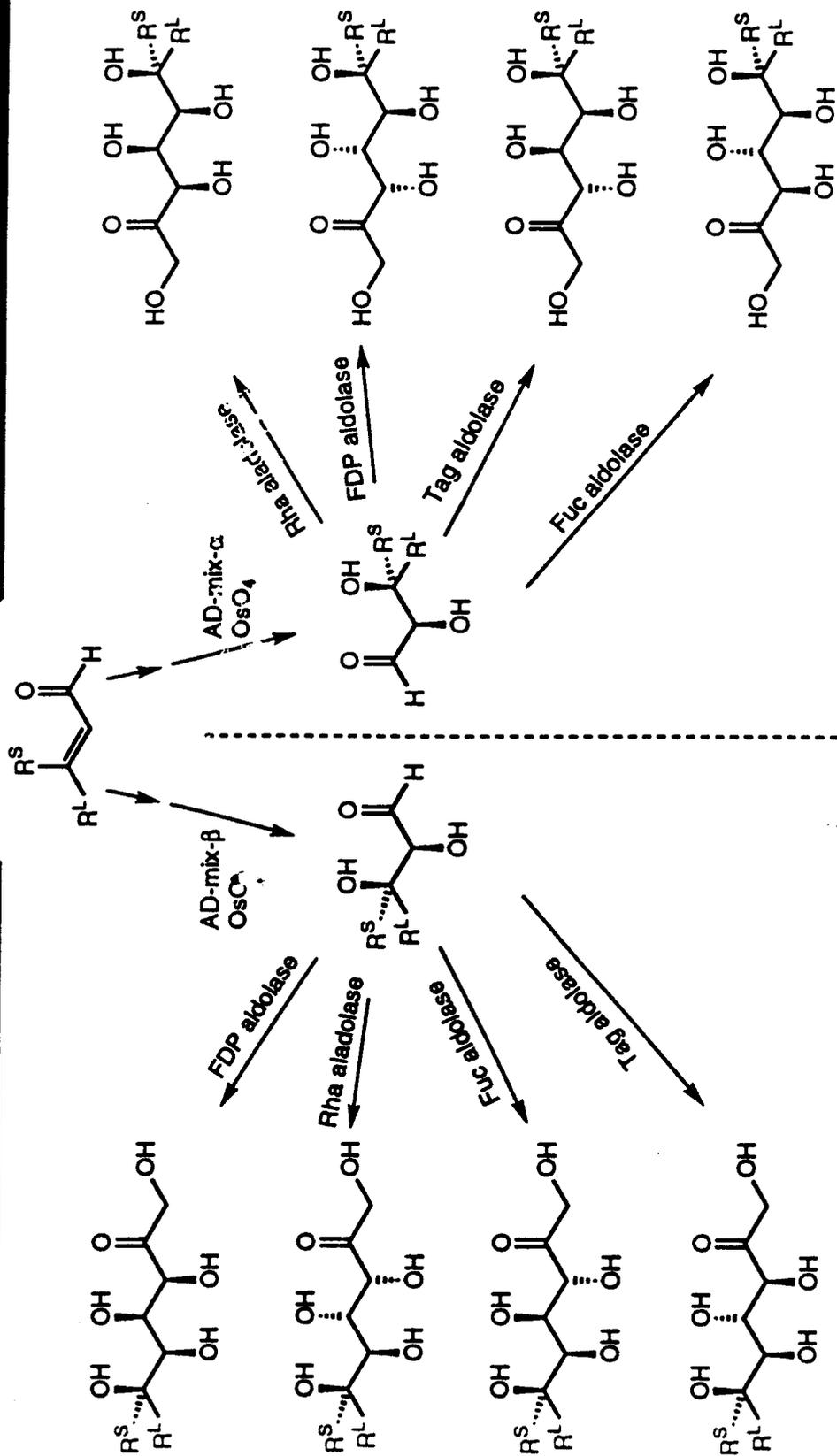
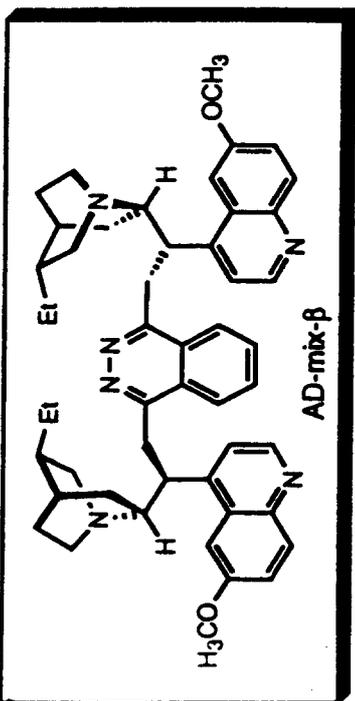
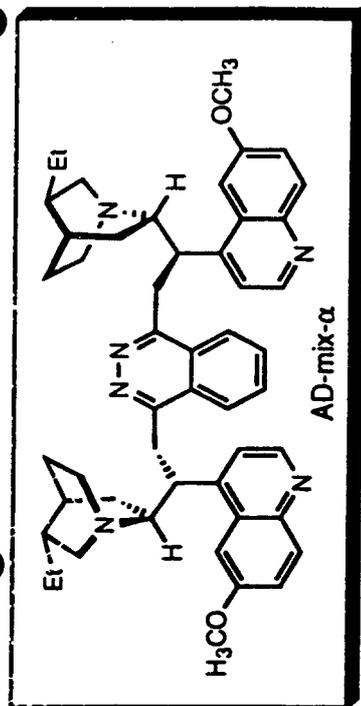


1) Acid phosphatase
2) Fucose isomerase

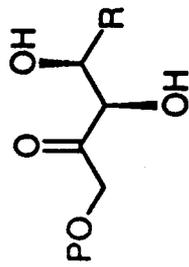


R = CH₃, CH₂CH₃, CH₂F, CH₂Ph, Ph

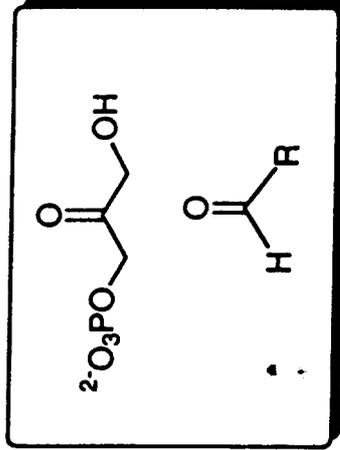




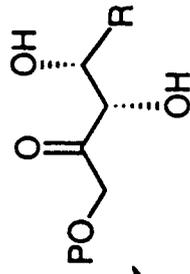
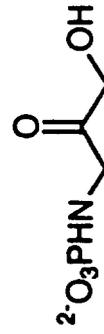
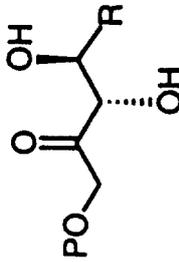
Dihydroxyacetone phosphate-dependent Aldolases



Rham-1-P
aldolase

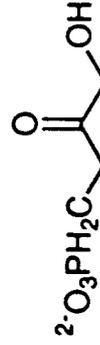
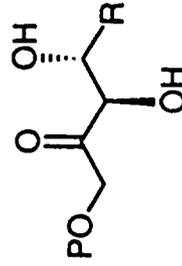


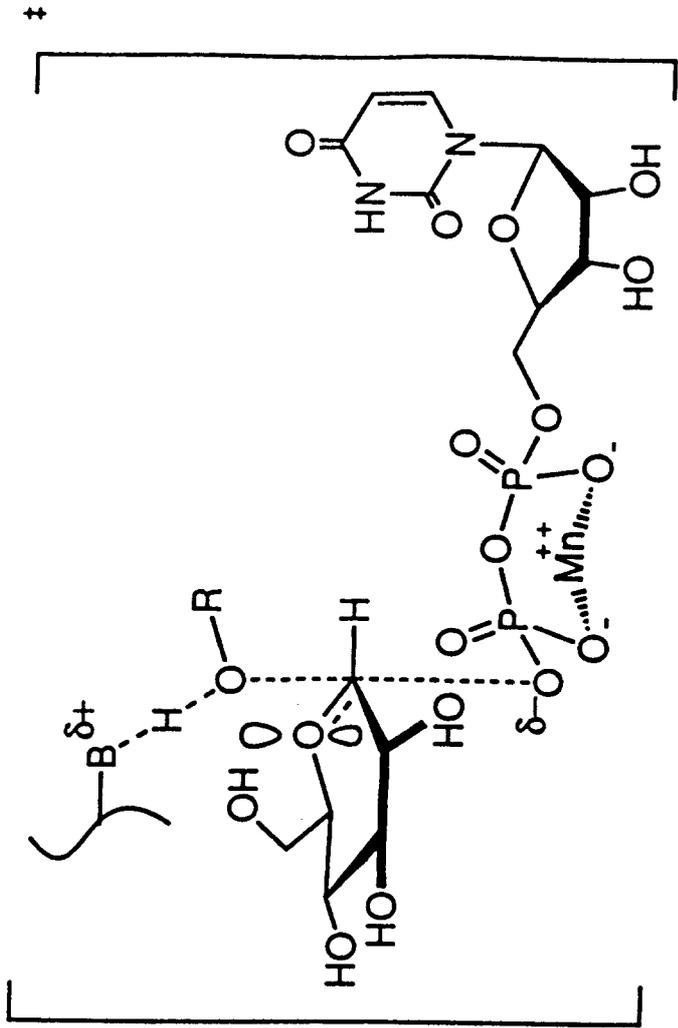
TDP
aldolase



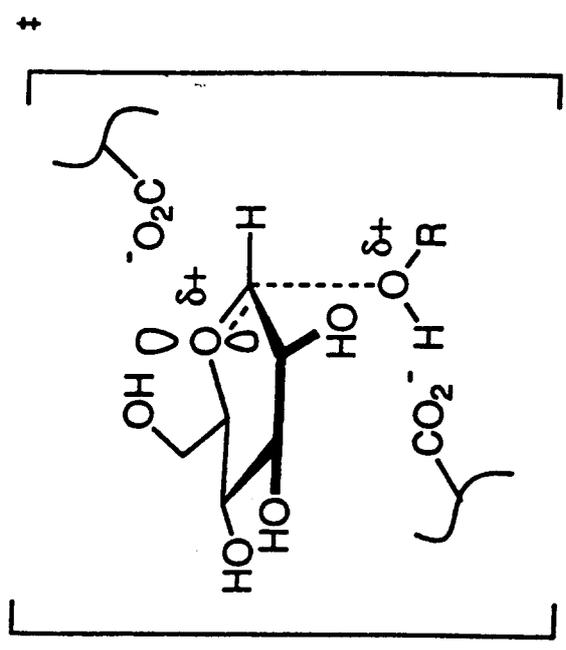
FDP
aldolase

Fuc-1-P
aldolase

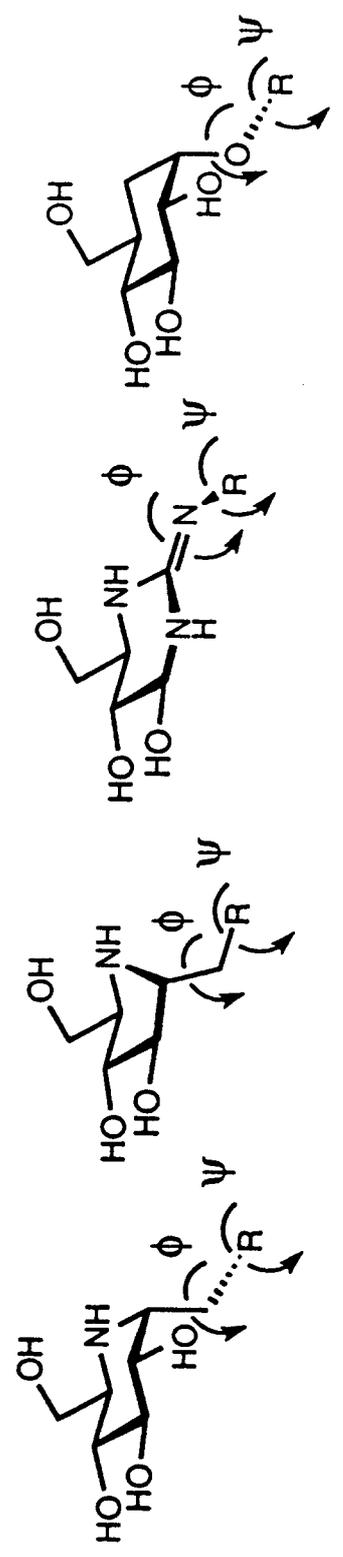
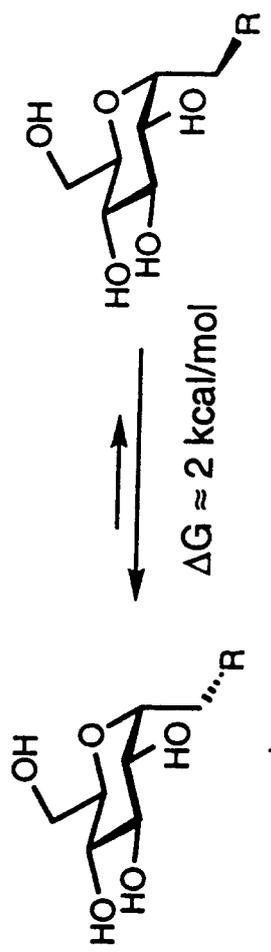
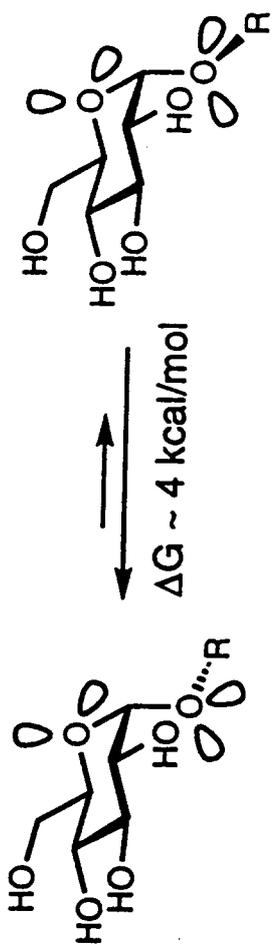




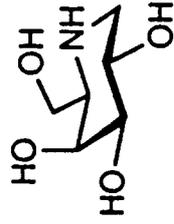
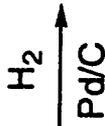
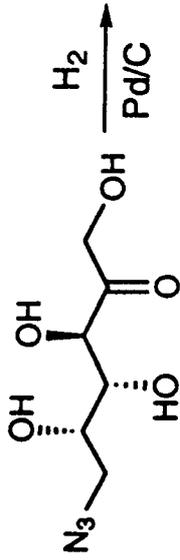
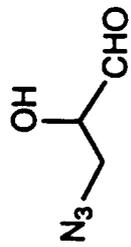
Glycosyltransferase Reaction



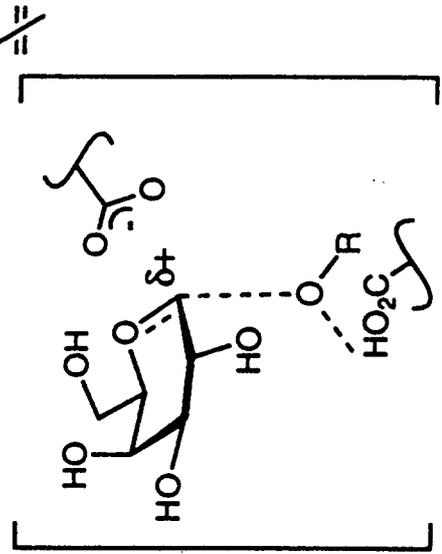
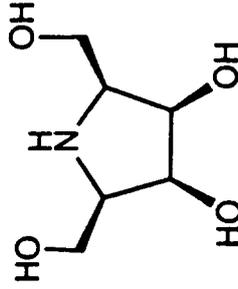
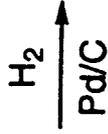
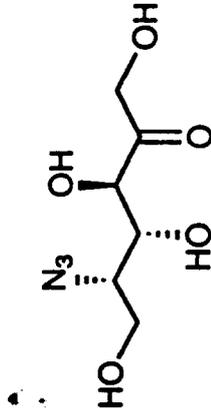
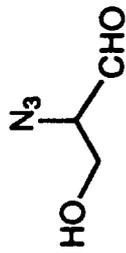
Glycosidase Reaction



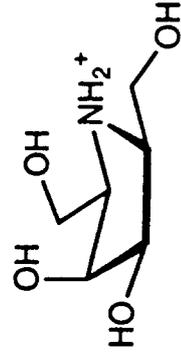
Inhibitors of α -Galactosidase



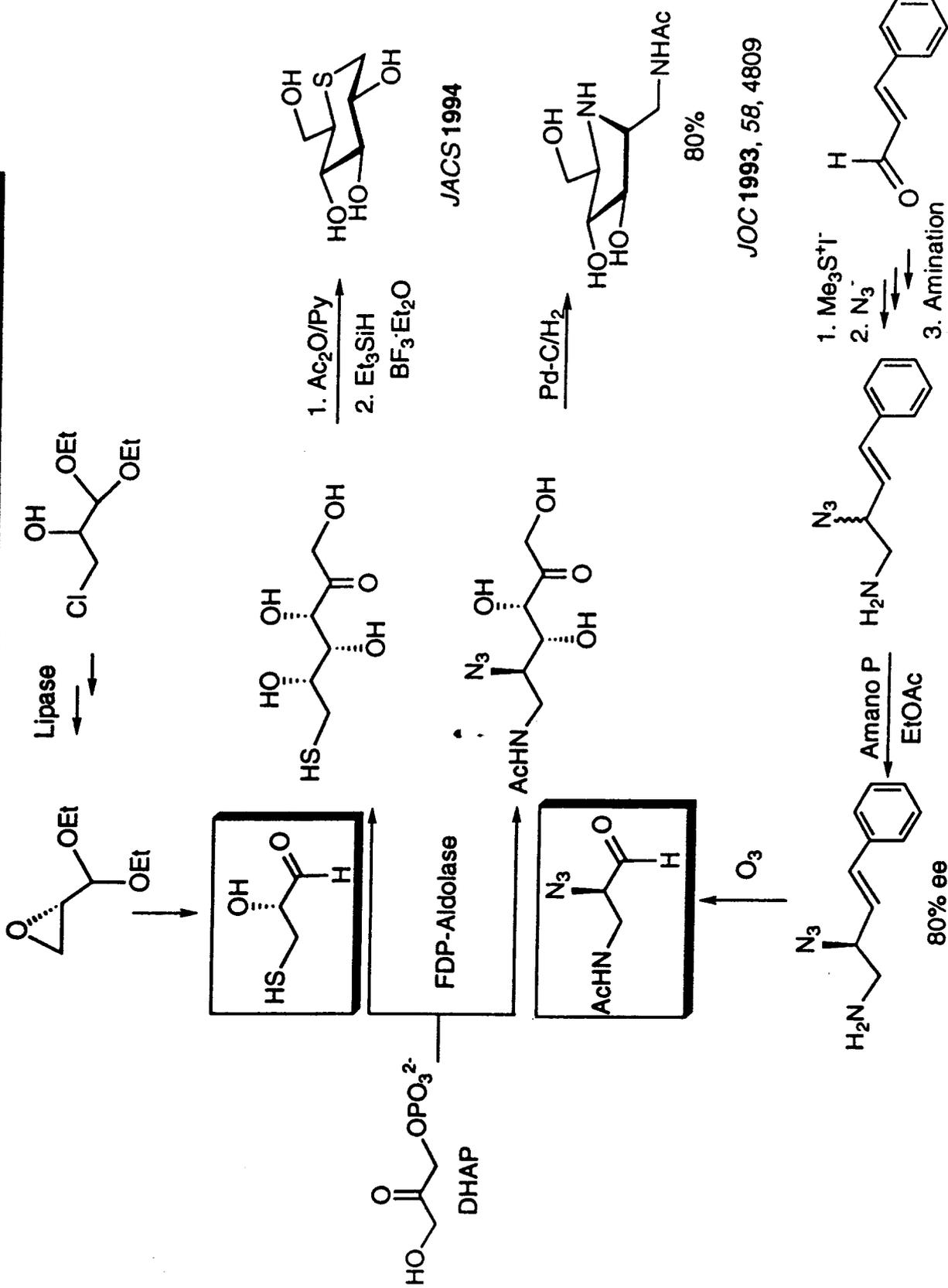
K_i = 1.6 nM



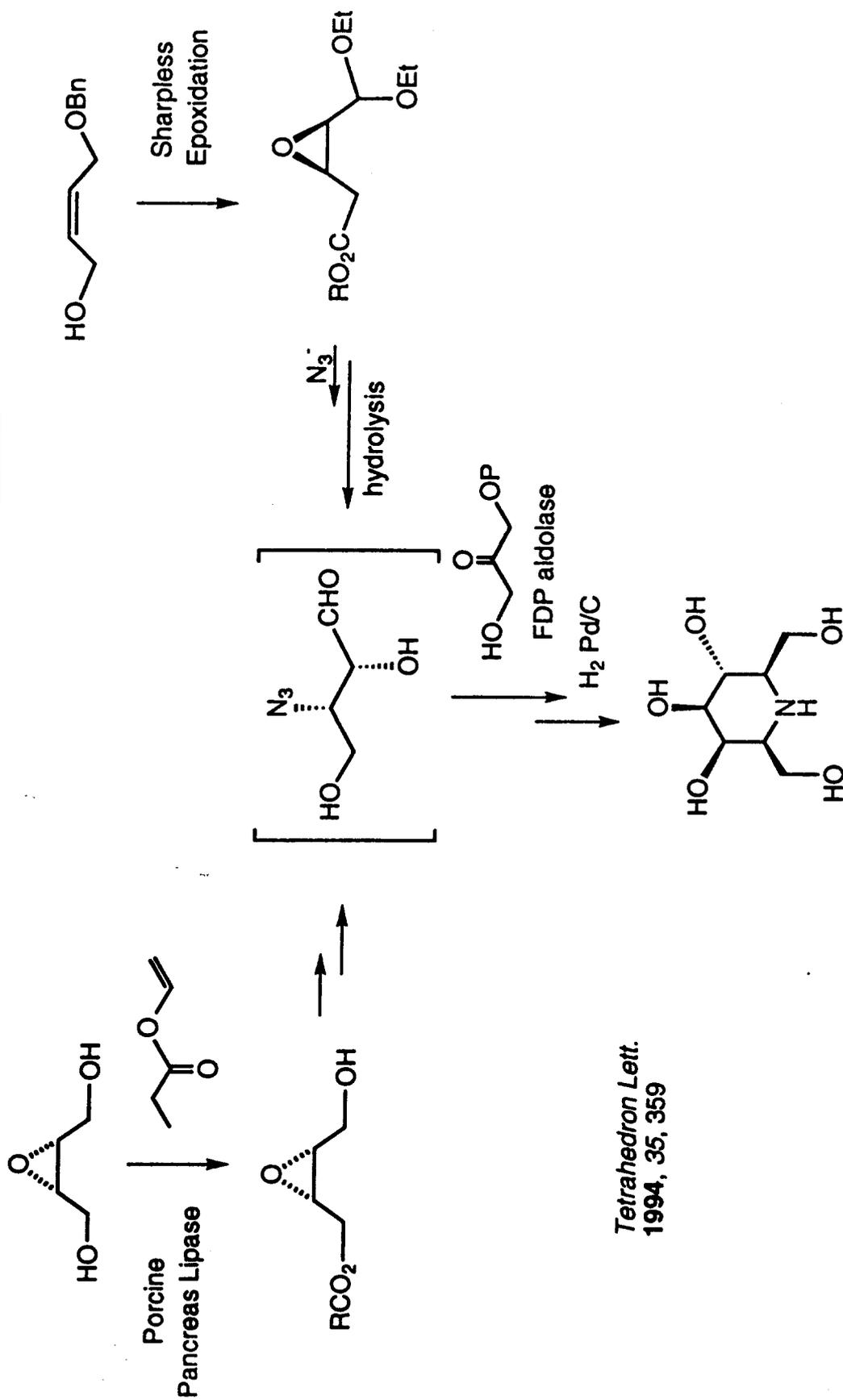
K_i = 50 nM



Aminoazasugar and Thiosugar from FDP-Aldolase

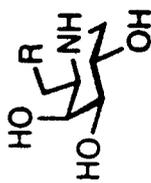


Synthesis of Homoazasugar



Tetrahedron Lett.
1994, 35, 359

From Fructose-1,6-diphosphate aldolase



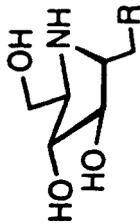
R=OH,H



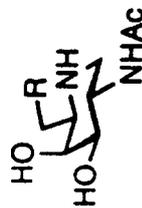
R=OH,H



R=F,OEt



R=OH,NHAc



R=OH,H



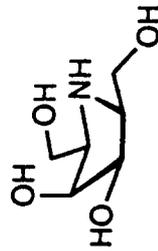
R=OH,H



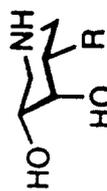
R=OH,H



R=H,CH₃



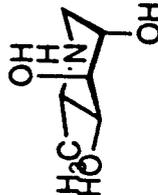
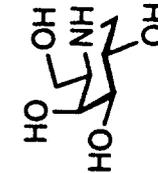
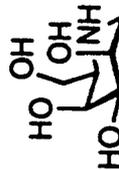
From 2-Deoxyribose-5-phosphate aldolase



R=H,CH₃



R=CH₃,CH₂F

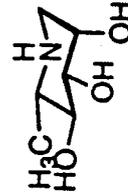


From Fucose-1-phosphate aldolase

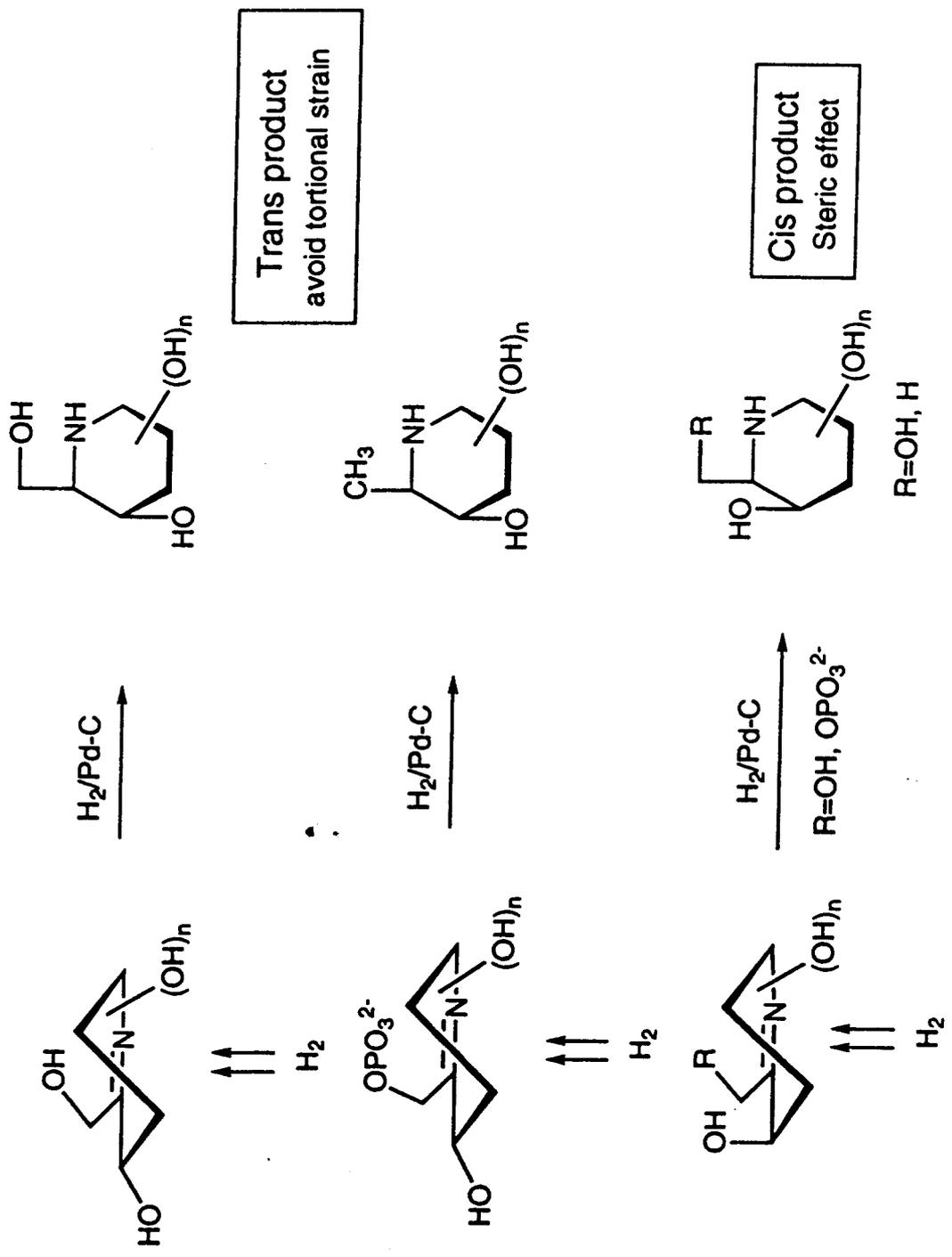
From Sialic acid aldolase



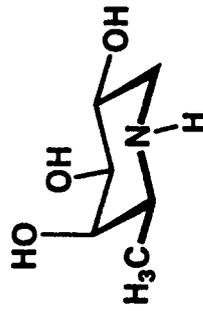
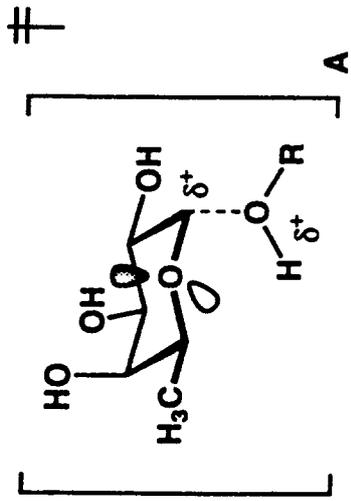
From Rhamnose-1-phosphate aldolase



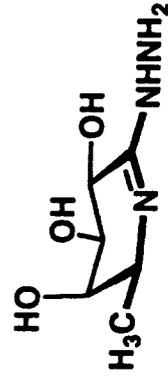
Stereochemistry of Pd-Mediated Reductive Amination



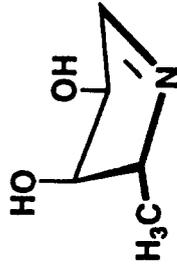
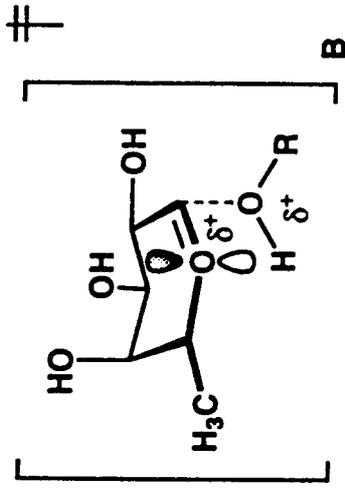
α-Fucosidase-Catalyzed Reaction



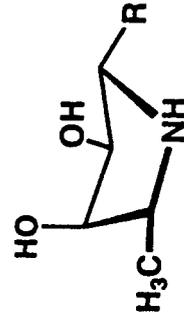
$K_I = 10 \text{ nM}$



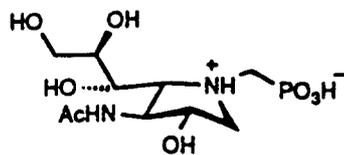
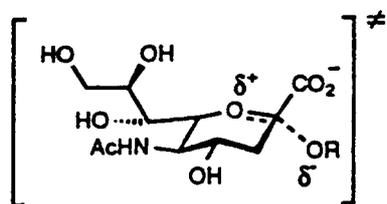
$K_I = 820 \text{ nM}$



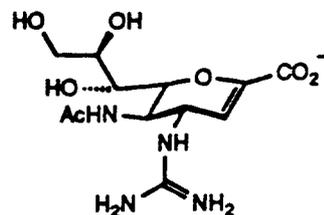
$K_I = 160 \text{ nM}$



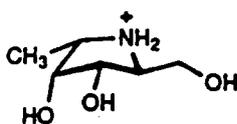
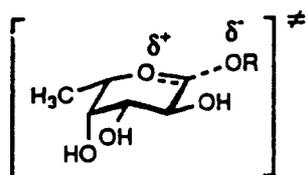
$\frac{R}{H}$	$K_I (\mu\text{M})$
CH ₂ OH	2.0
CH ₂ NH ₂	1.2
	1.9



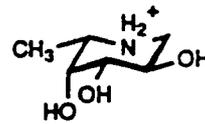
$K_i = 5.3 \times 10^{-5} \text{ M}$
sialidase (*V. cholerae*)
(ref. 325b)



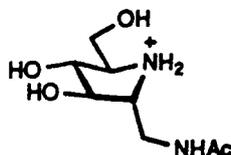
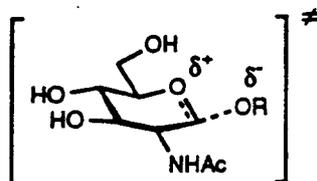
$K_i = 2 \times 10^{-10} \text{ M}$
influenza virus sialidase
(ref. 325c)



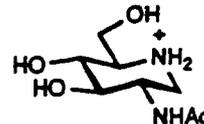
$K_i = 1.4 \times 10^{-6} \text{ M}$
 α -fucosidase (bovine kidney)
(ref. 54)



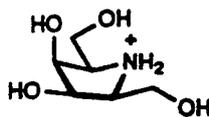
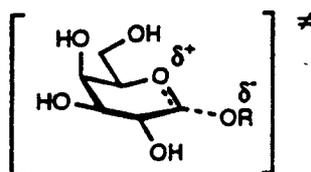
$K_i = 1 \times 10^{-8} \text{ M}$
 α -fucosidase
(human liver) (ref. 325d)



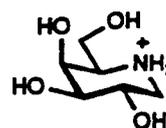
$K_i = 1.9 \times 10^{-6} \text{ M}$
 β -N-acetylglucosaminidase
(Jack beans) (ref. 58)



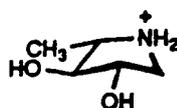
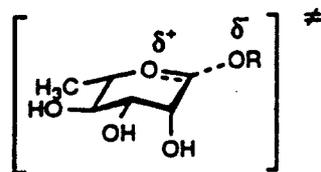
$K_i = 3.8 \times 10^{-7} \text{ M}$
 β -N-acetylglucosaminidase
(bovine kidney) (ref. 51)



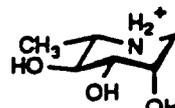
$K_i = 5.0 \times 10^{-8} \text{ M}$
 α -galactosidase
(green coffee beans) (ref. 325e)



$K_i = 1.6 \times 10^{-9} \text{ M}$
 α -galactosidase
(green coffee bean) (ref. 325f)



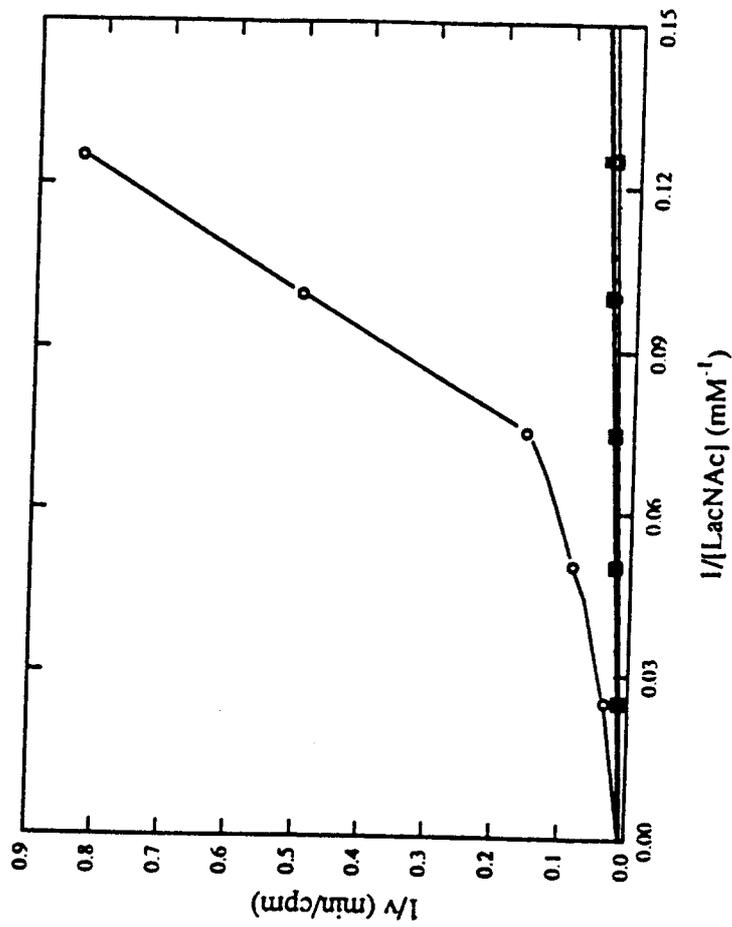
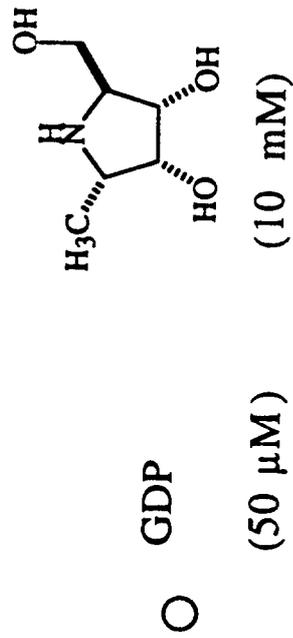
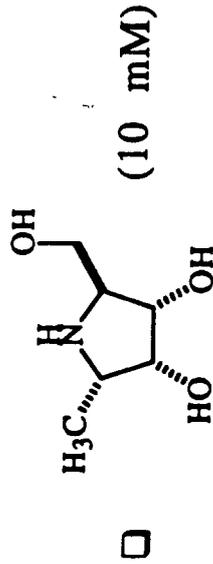
$K_i = 5 \times 10^{-6} \text{ M}$
 α -rhamnosidase
(ref. 525g)



$K_i = 5 \times 10^{-5} \text{ M}$
 α -rhamnosidase
(ref. 325g)

Synergistic Inhibition of α 1,3-Fucosyltransferase

▲ GDP (50 μ M)



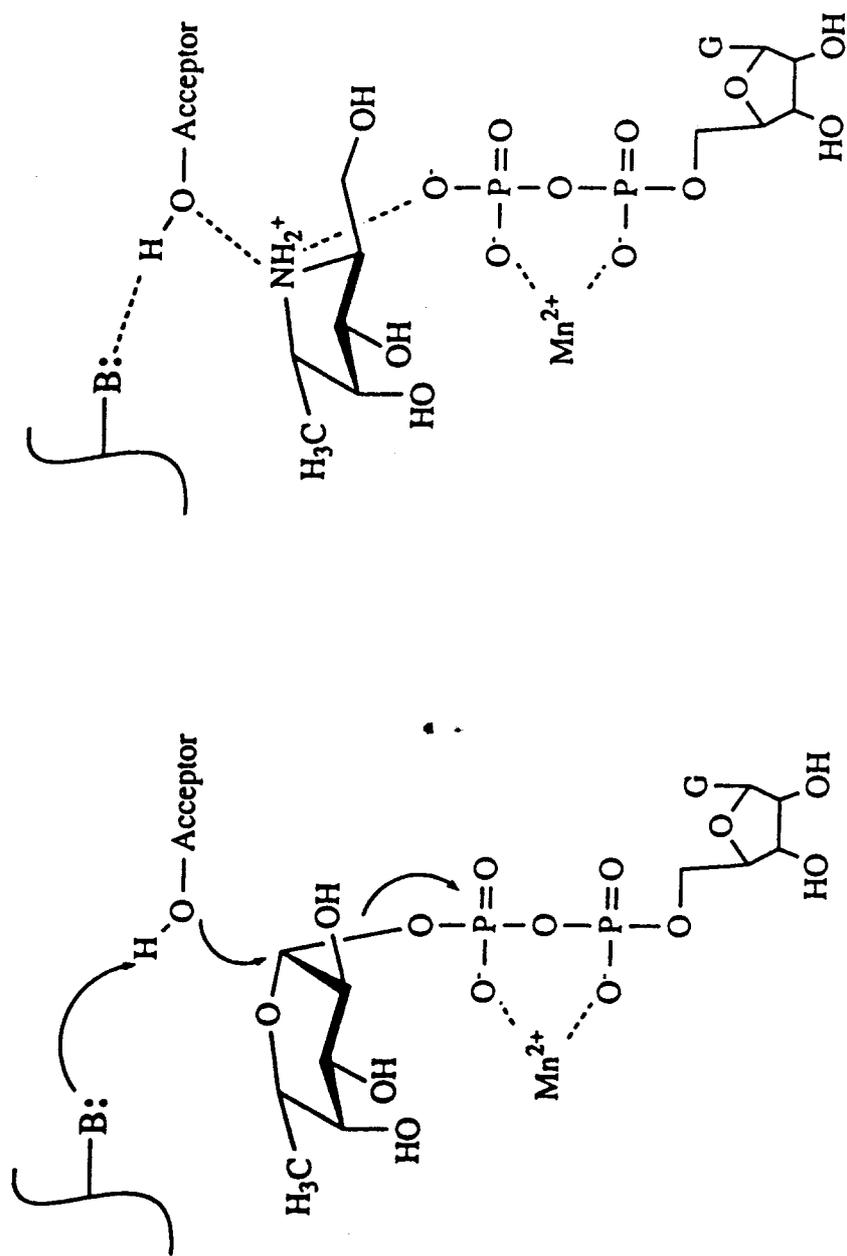
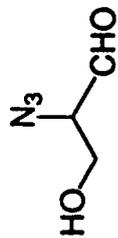
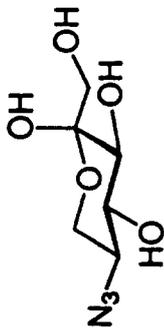


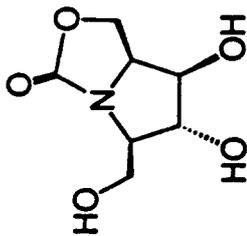
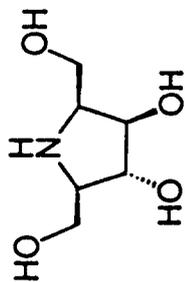
Figure 7. A proposed complex of GDP and a five-membered aza sugar which mimics the transition state of the fucosyltransferase reaction.



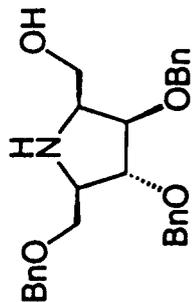
DHAP
FDP Aldolase



H₂/Pd-C

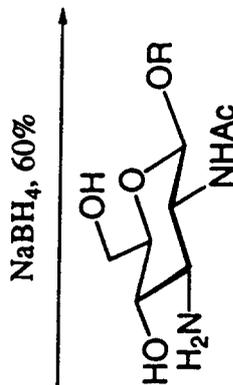
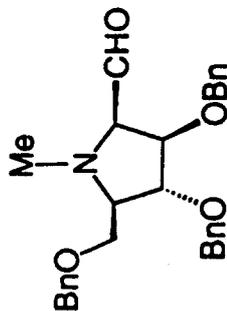


Triphosgene
91%

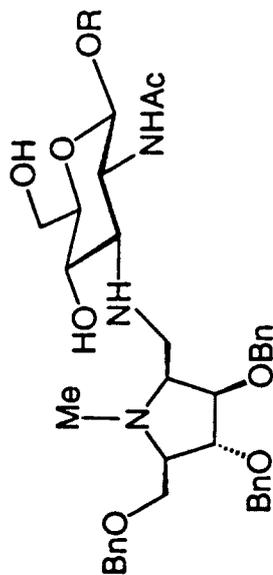


1) BnBr/NaH
88%
2) KOH

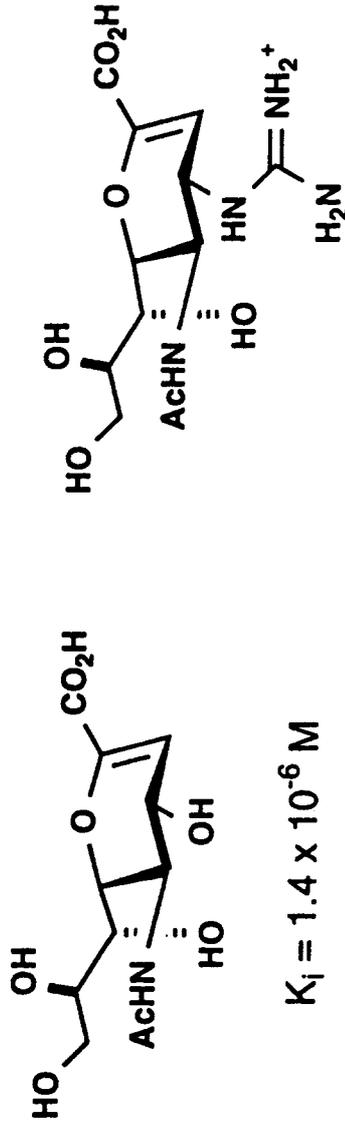
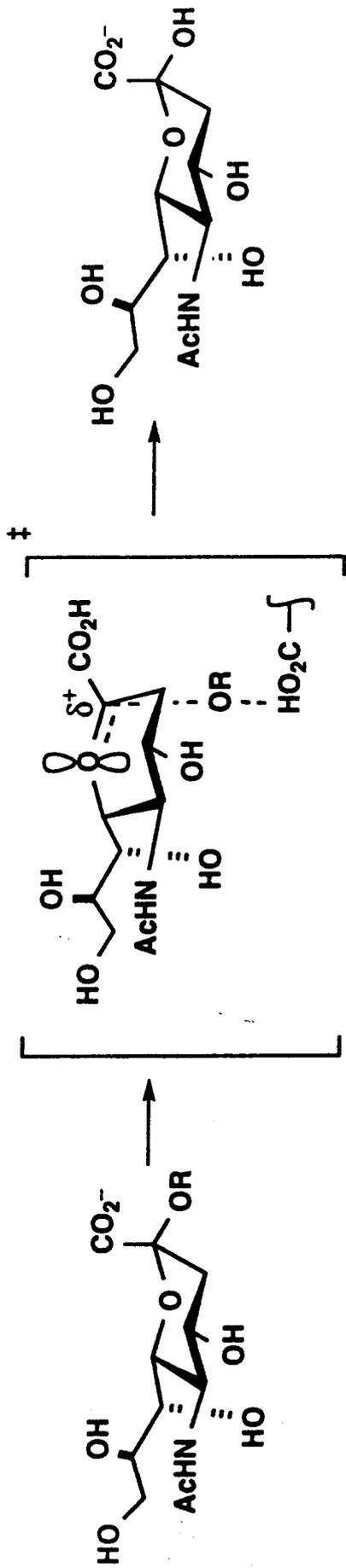
1) (CH₂O)_r/NaCNBH₃
81%
2) DMSO/(CO)₂Cl₂



NaBH₄, 60%



Sialidase (Neuraminidase) Reaction



$$K_i = 1.4 \times 10^{-6} \text{ M}$$

Miller et al.

BBRC 1978, 83, 1479

$$K_i = 2 \times 10^{-10} \text{ M}$$

von Itzstein et al.

Nature 1993, 363, 418

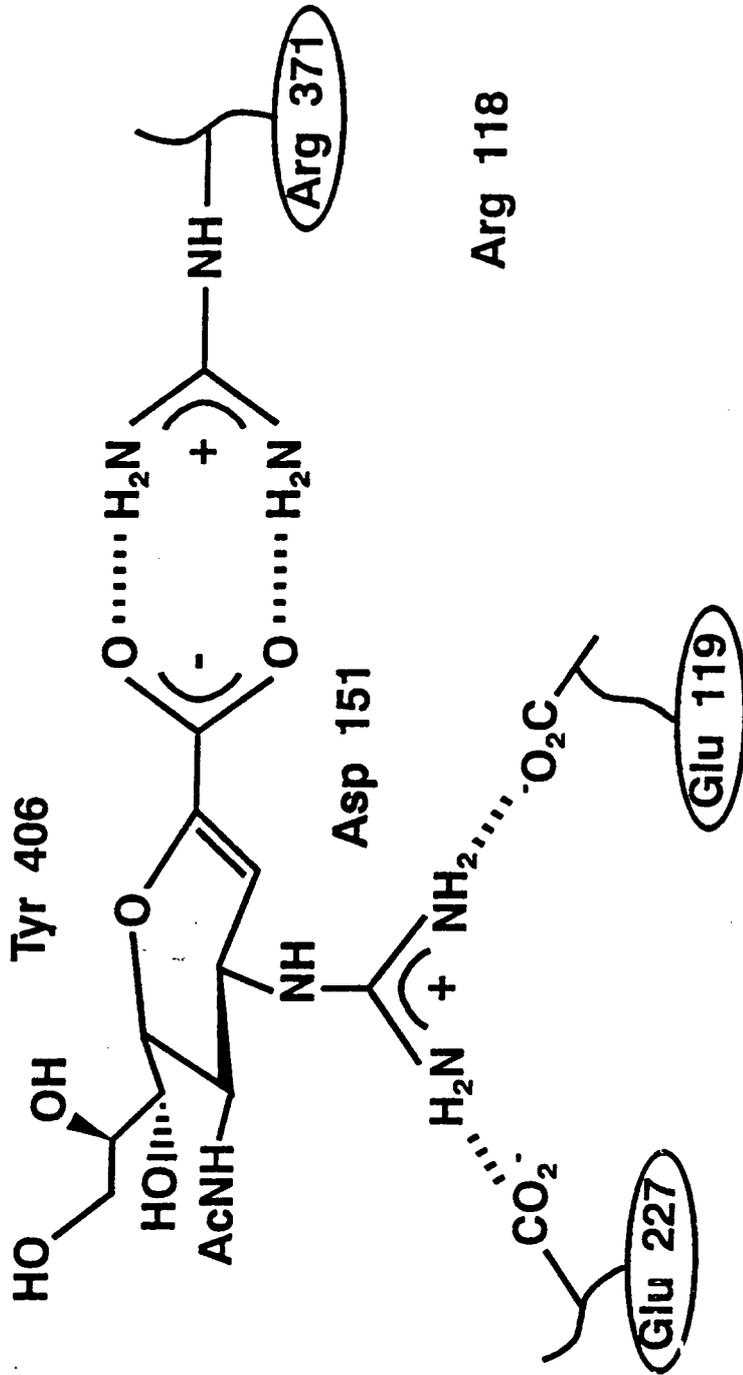
Influenza Neuraminidase Inhibitor: 2-Deoxy-2,3-Dideoxy-D-N-Acetylneuraminic Acid

Glu 276

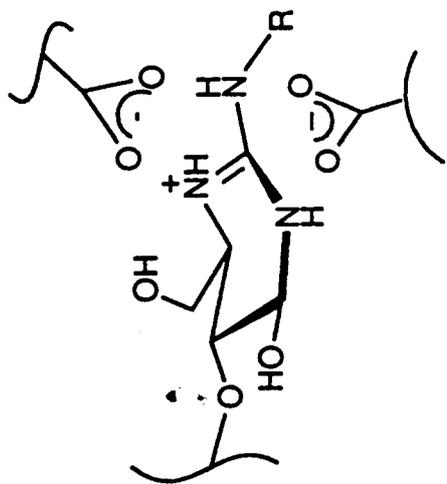
Arg 292

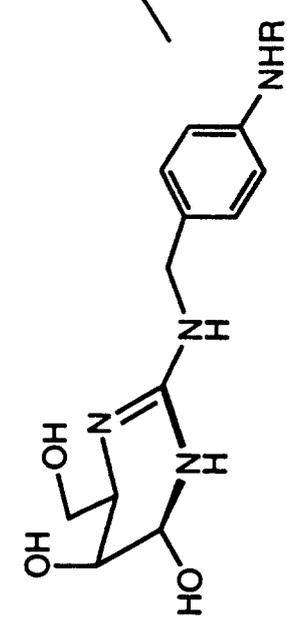
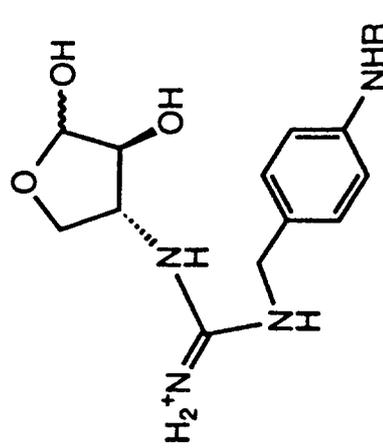
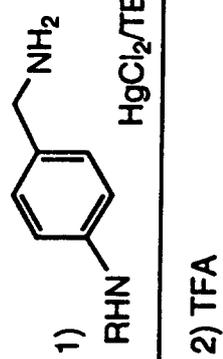
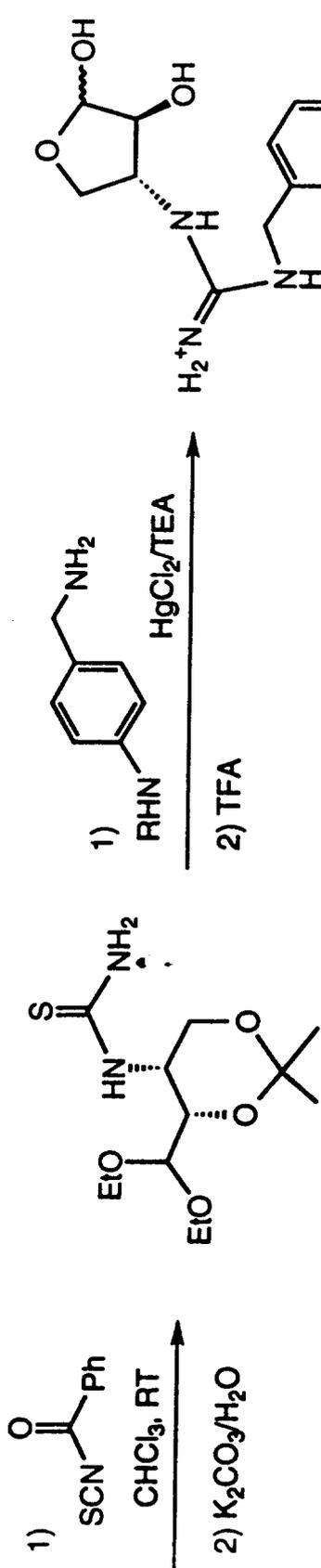
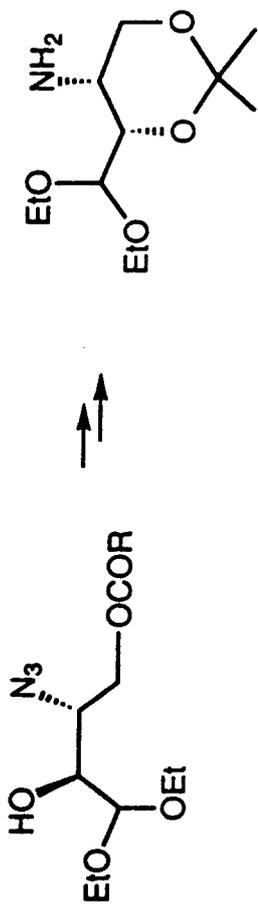
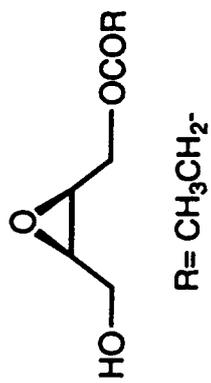
Glu 277

Tyr 406



$K_i = 2 \times 10^{-10} \text{ M}$ *Nature* 1993, 363, 418





51%

Guanidino-galactoside

Enzymatic Synthesis of Phospholipid-Inhibitor Conjugates



Assembly

