

Peptides as Switchable Molecular Scaffolds and Asymmetric Catalysts

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Catalytic Activity

Selective Cleavage

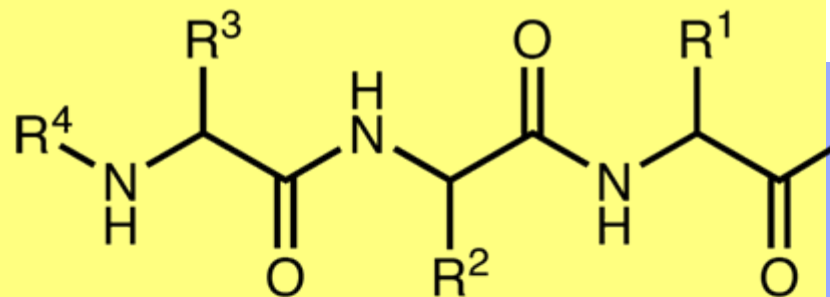
Peptides

Molecular Recognition

Stereoelectronic Effects

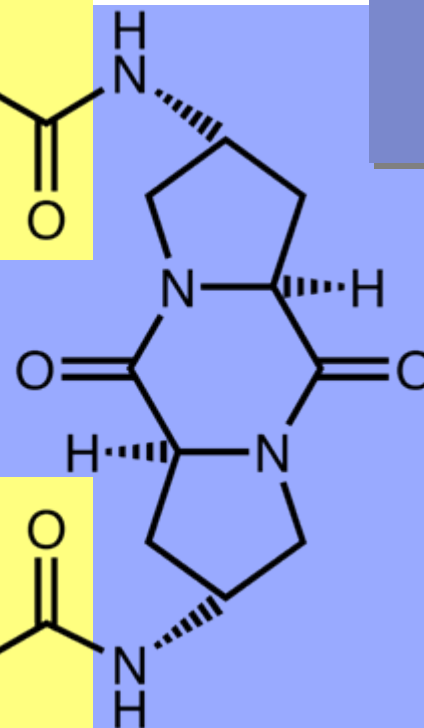
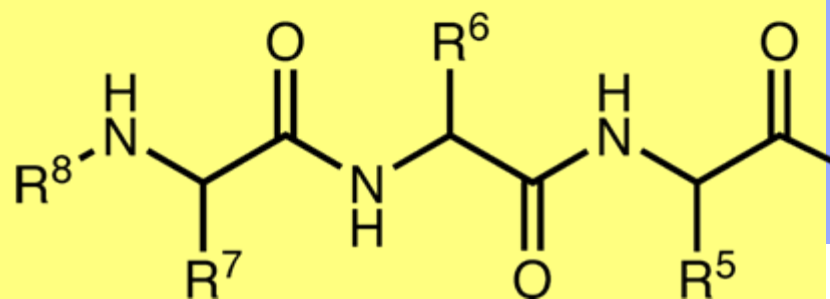
Polyproline

Diketopiperazine Receptors - Sequence Selective Binding of Short Peptides



crucial for
binding

determine the
binding selectivity



Applications of Diketopiperazine Receptors

Sensors

Prof. J. Bargon, University Bonn

Material Sciences

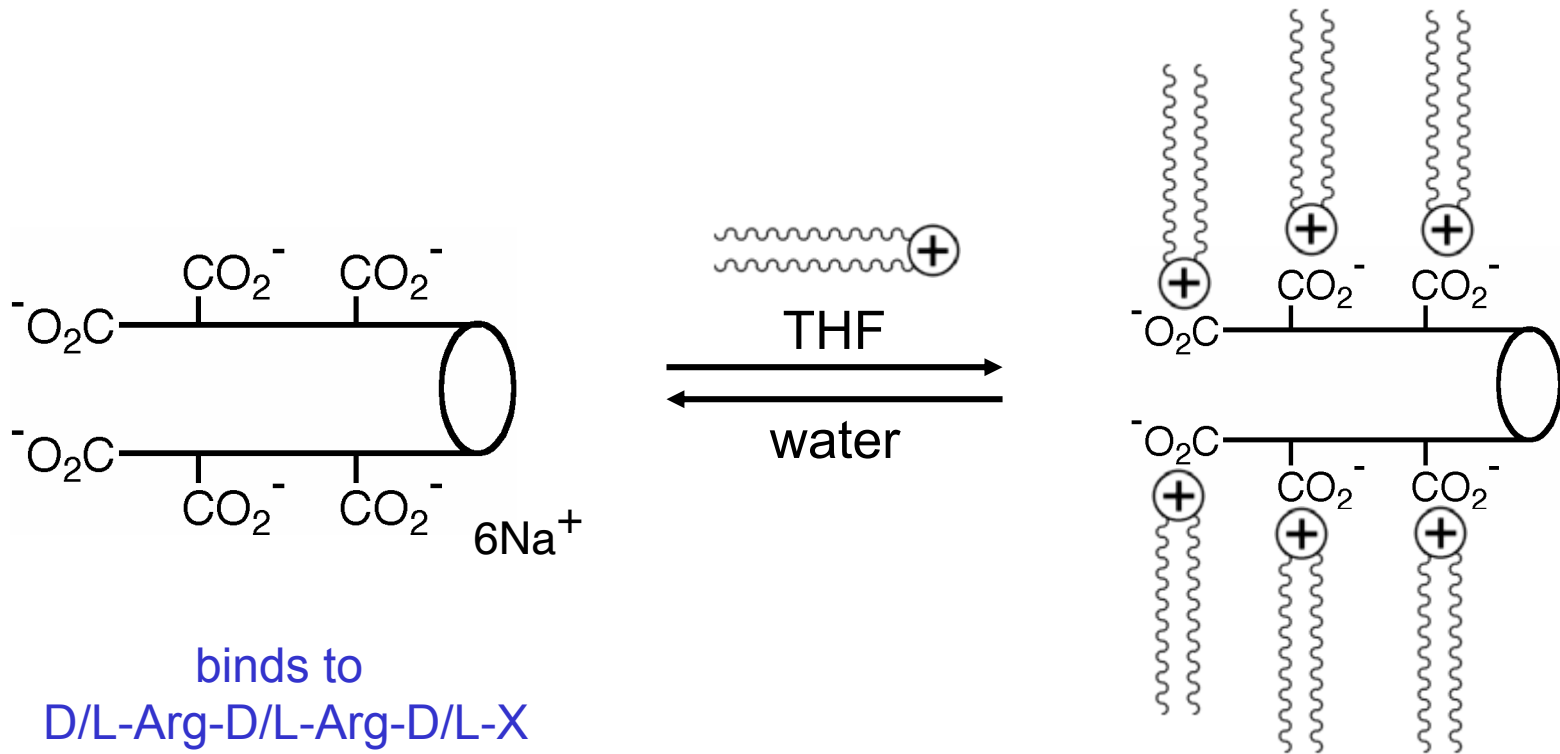
Prof. C. Faul, Bristol University

Diagnostics / Synthetic Antibodies

Prof. M. Przybylski, University of Konstanz

Prof. U. Jenal, Biocentre Basel

Diketopiperazine Receptor-Surfactant Complexes

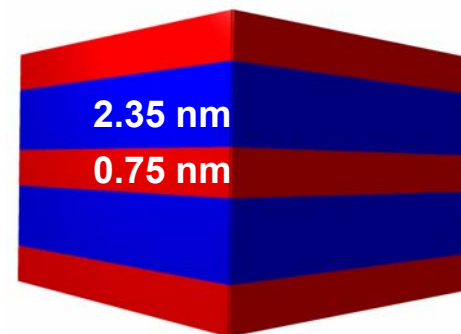
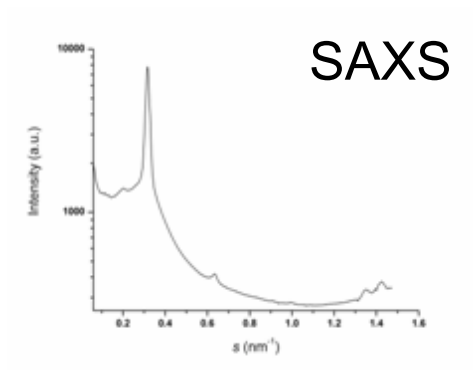
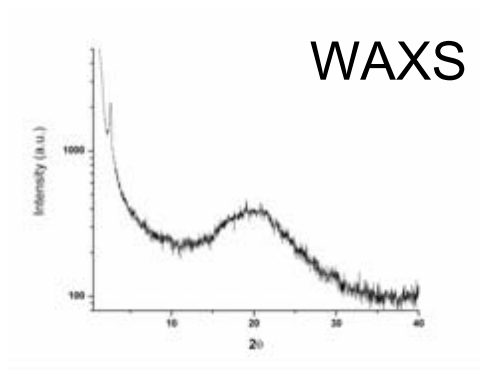
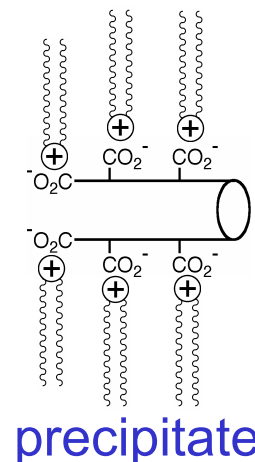
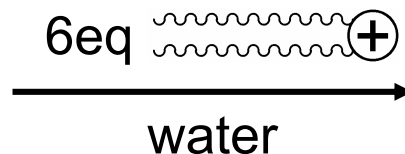
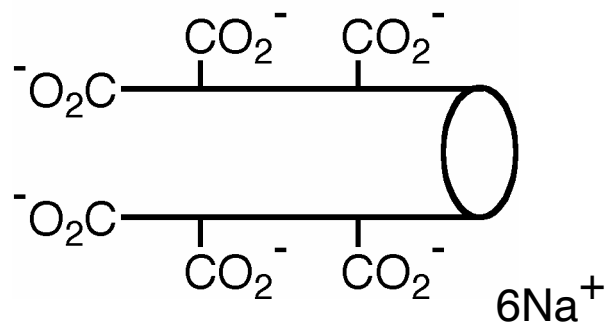


binds to
 D/L-Arg-D/L-Arg-D/L-X
 (X = Arg, Ser, Thr, or Cys)
 in aqueous solution at pH 7 or 10
 $\Delta G = 5-6 \text{ kcalmol}^{-1}$

no binding
 to any of 29791 peptides



Material Properties of Diketopiperazine Receptor-Surfactant Complexes



→ liquid crystalline properties

Catalytic Activity

Selective Cleavage

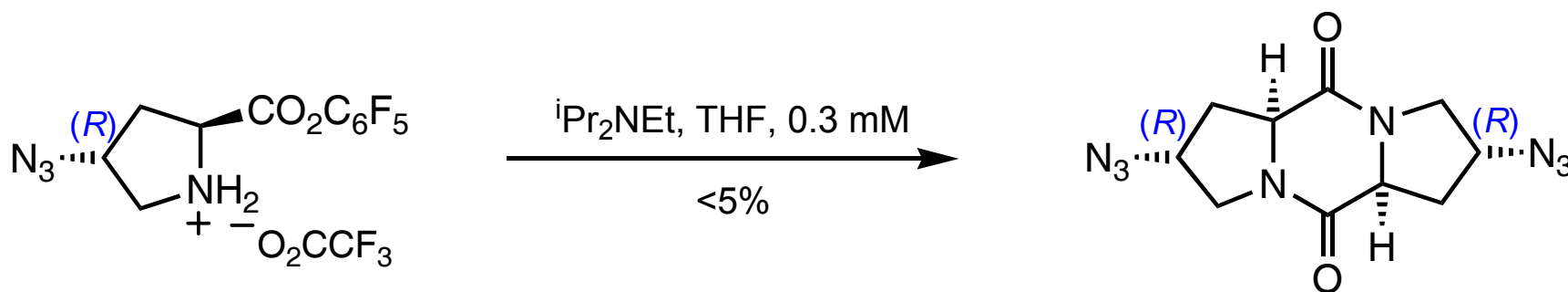
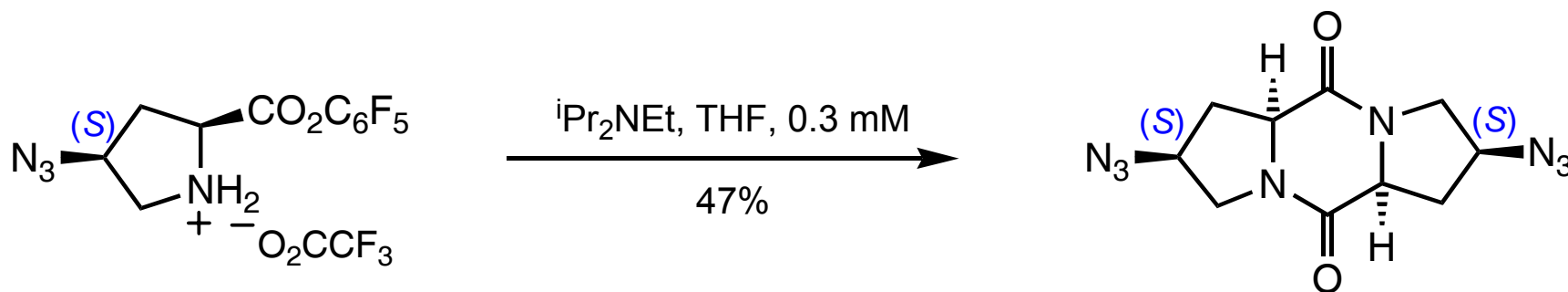
Peptides

Molecular Recognition

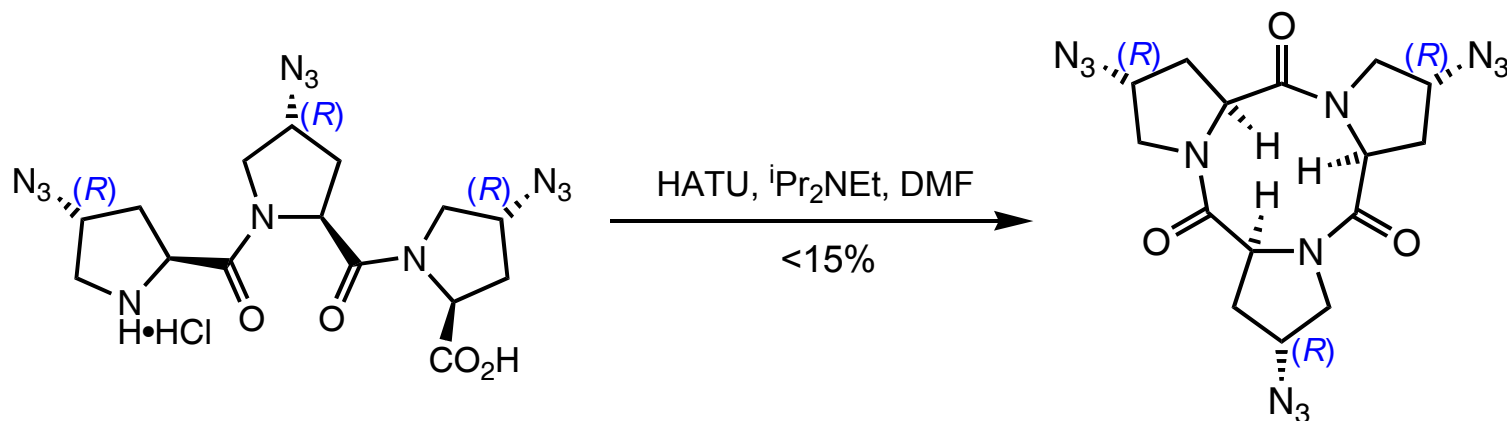
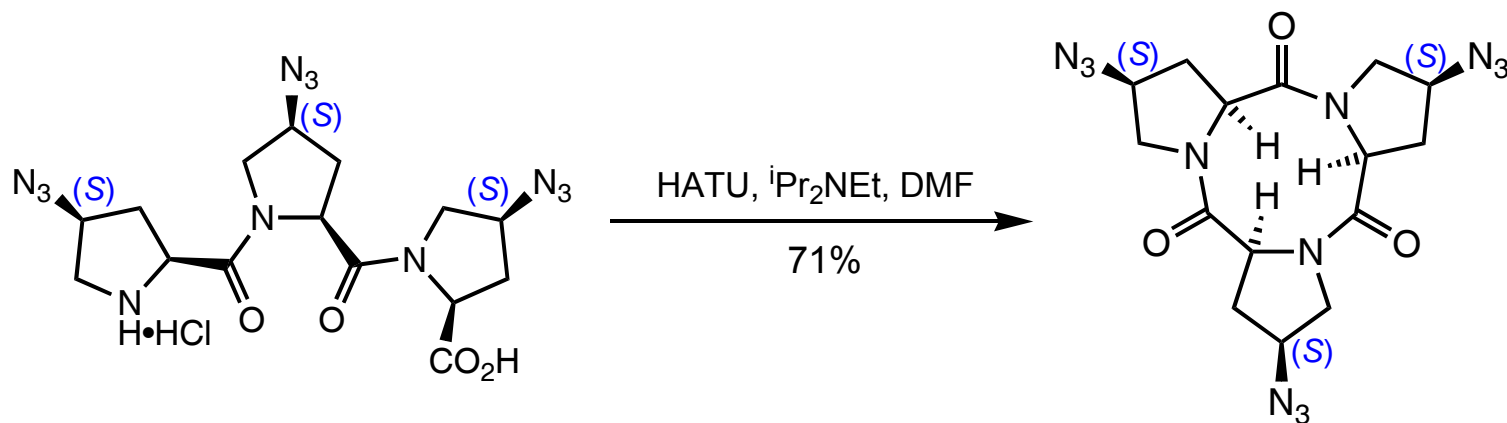
Stereoelectronic Effects

Polyproline

Syntheses of (4*S*,4'*S*)- and (4*R*,4'*R*)-Azp-Diketopiperazines

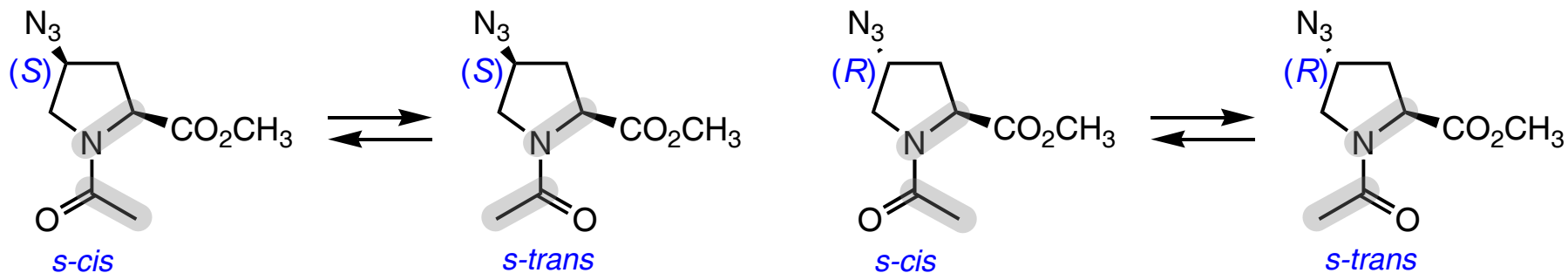


Syntheses of Cyclotri[(4*S*)- and (4*R*)-azidoproline]



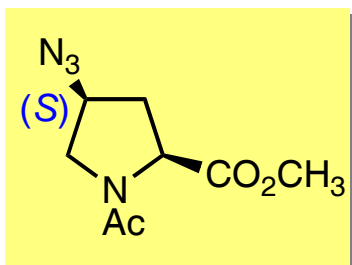
The absolute configuration at C(4) influences the cyclisation tendency!

Influence of the configuration at C(4) on the *s-cis* : *s-trans* ratio

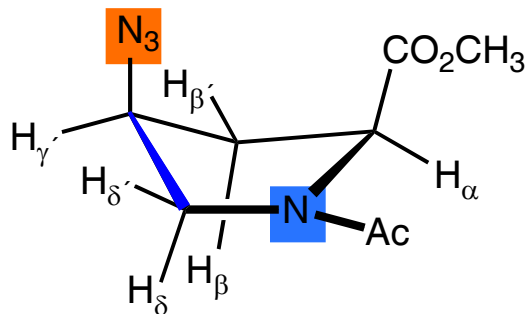


solvent	N-Ac-(4S)-N ₃ -Pro-OCH ₃ <i>s-cis</i> : <i>s-trans</i>	N-Ac-(4R)-N ₃ -Pro-OCH ₃ <i>s-cis</i> : <i>s-trans</i>
D ₂ O	1 : 2.6	1 : 6.1
d ₇ -DMF	1 : 1.6	1 : 3.8
d ₅ -pyridine	1 : 2.0	1 : 4.5
CD ₃ OD	1 : 1.7	1 : 4.0
d ₆ -acetone	1 : 1.6	1 : 3.7
CDCl ₃	1 : 1.9	1 : 3.9
d ₈ -dioxane	1 : 2.0	1 : 4.2

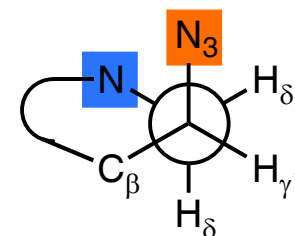
Conformational Analysis of the Pyrrolidine Rings



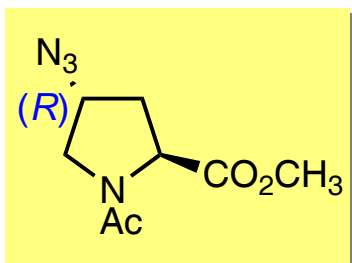
C(4)-endo-conformation



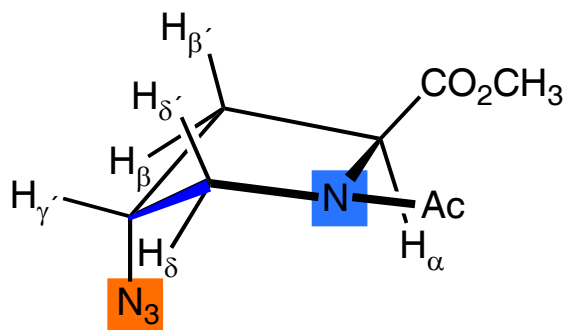
pseudo-axial position



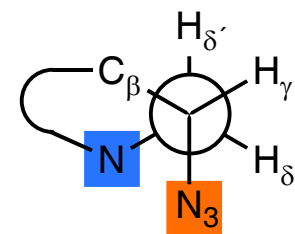
gauche conformation



C(4)-exo-conformation

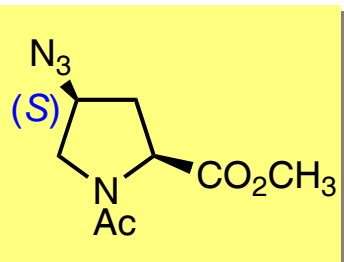


pseudo-axial position

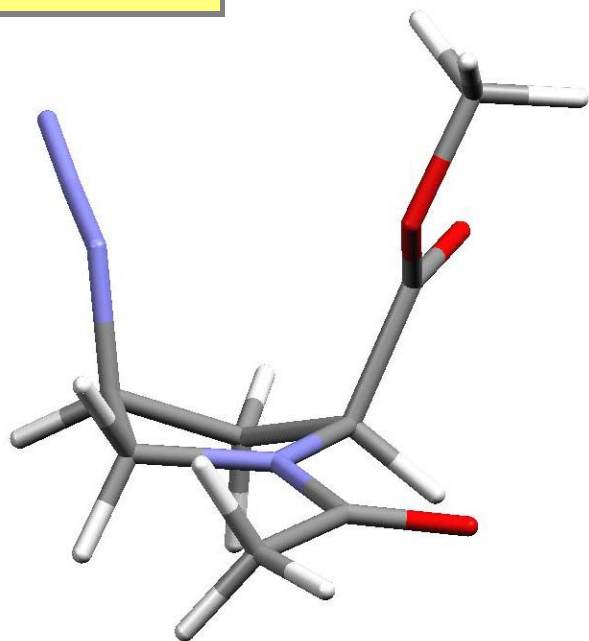


gauche conformation

An "azido gauche effect" determines the conformation of the pyrrolidine rings



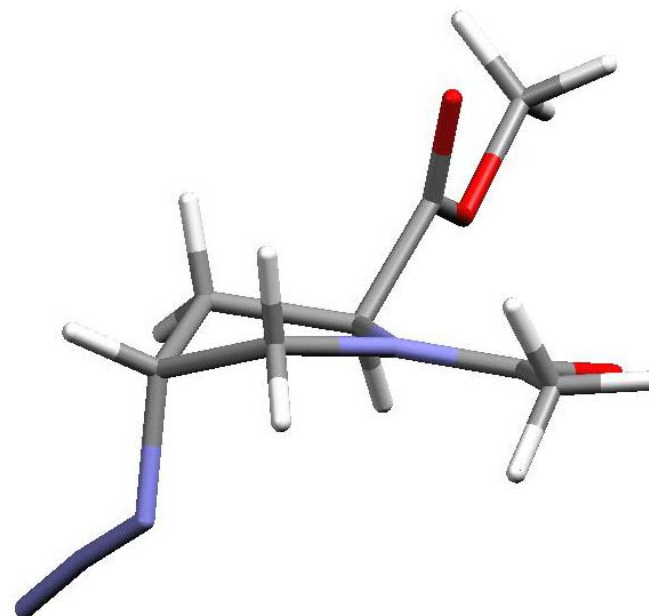
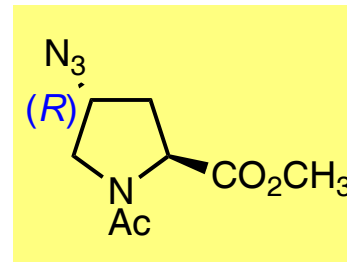
endo-conformation



ab-initio calculations

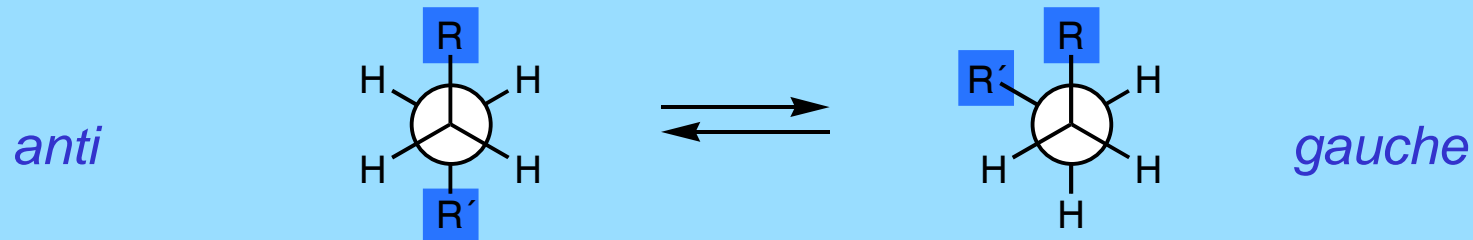
RI-MP2/TZVP level, B3LYP/6-31G** structures

exo-conformation



crystal structure analysis
(also ab-initio calculation)

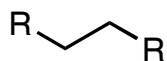
The “Azido Gauche Effect”



$$\Delta E = E_{anti} - E_{gauche} \text{ (kcal mol}^{-1}\text{)}$$

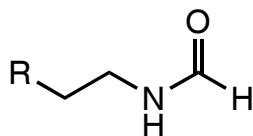
R = F

R = N₃



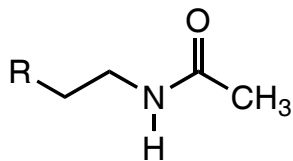
0.9

1.3



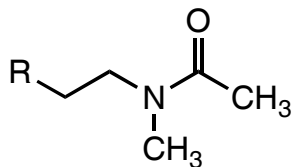
1.4

1.3



1.4

1.6



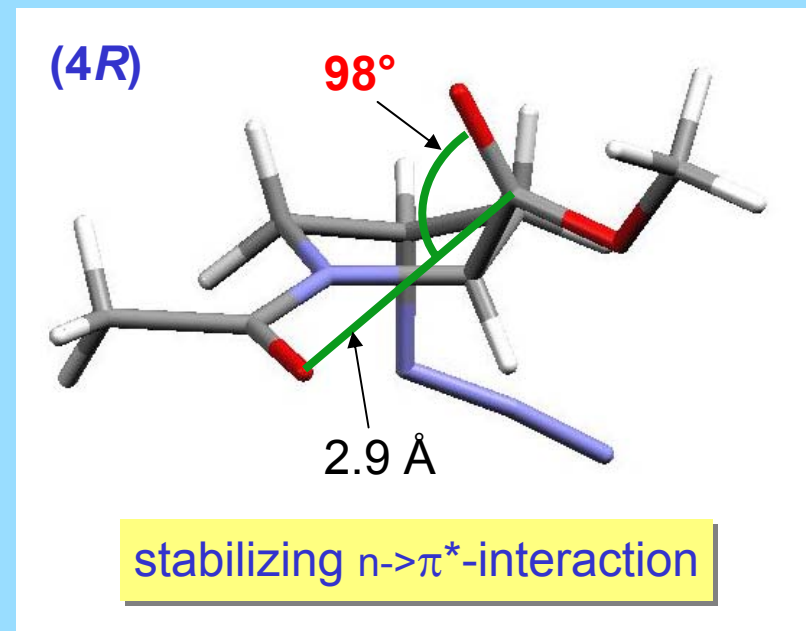
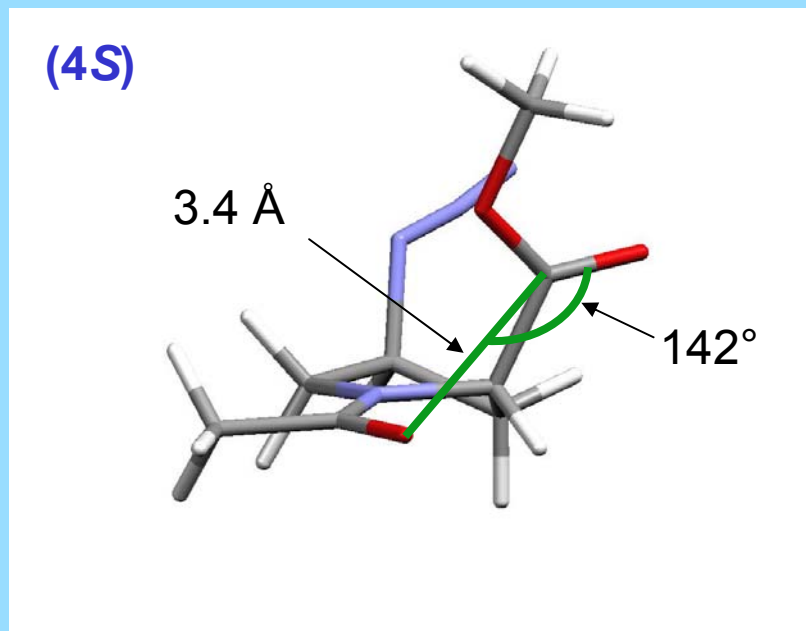
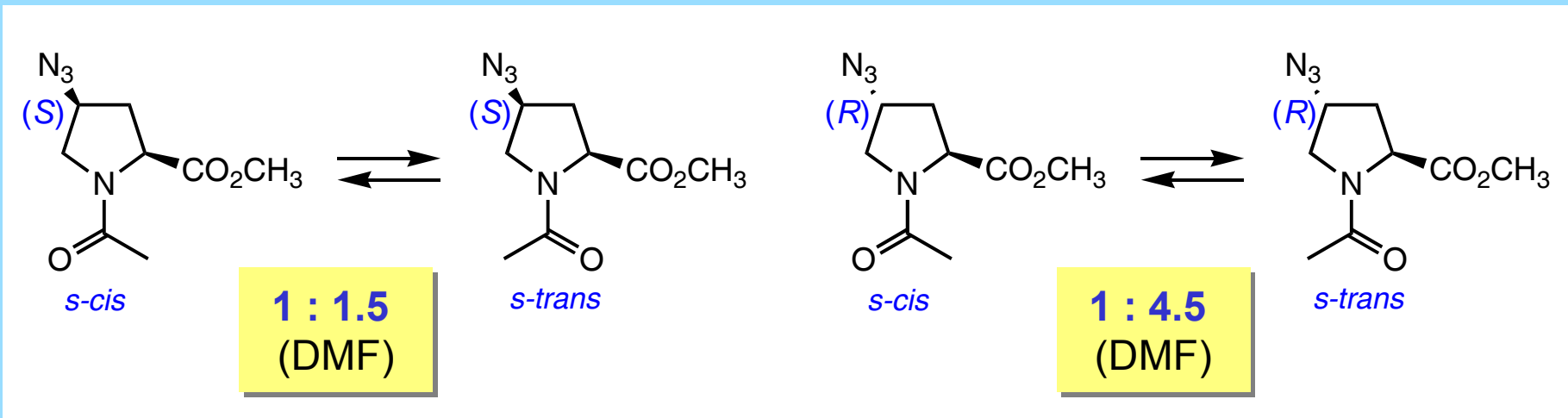
1.7

3.3

ab initio calculations using B3LYP/6-31G** for geometry optimization and RI_MP2/TZVP for energy optimisation

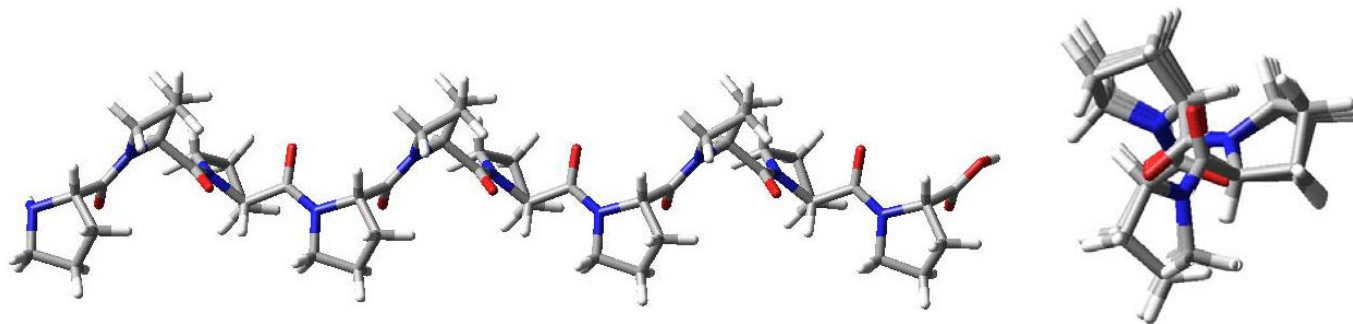
collaboration with Prof. Christian Ochsenfeld, University of Tübingen

What is the reason for the higher *s-trans* preference of (4*R*)-azidoproline?



Do azidoprolines effect the conformational stability of polyproline?

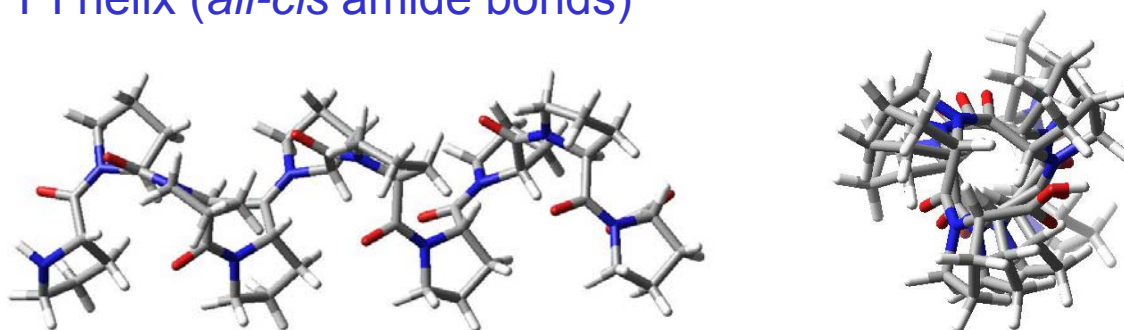
PPII helix (*all-trans* amide bonds)



in H₂O

should be stabilized by incorporation of (4*R*)-azidoproline

PPI helix (*all-cis* amide bonds)

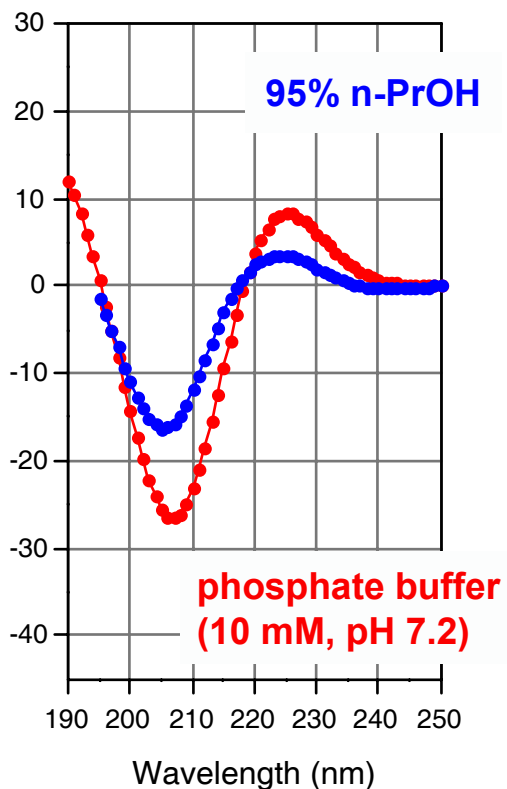


in n-PrOH

should be stabilized by incorporation of (4*S*)-azidoproline

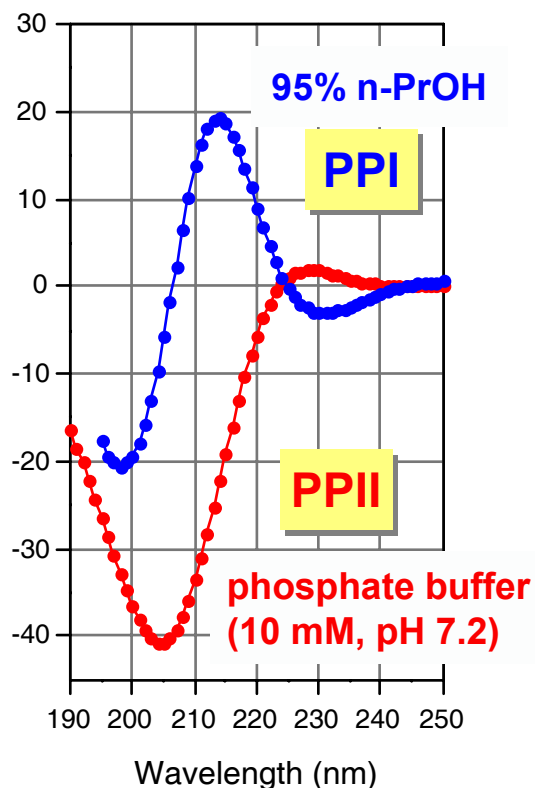
Polyprolines containing (4*S*)- and (4*R*)-azidoprolines

Ac-[(4*R*)Azp]₉-OH

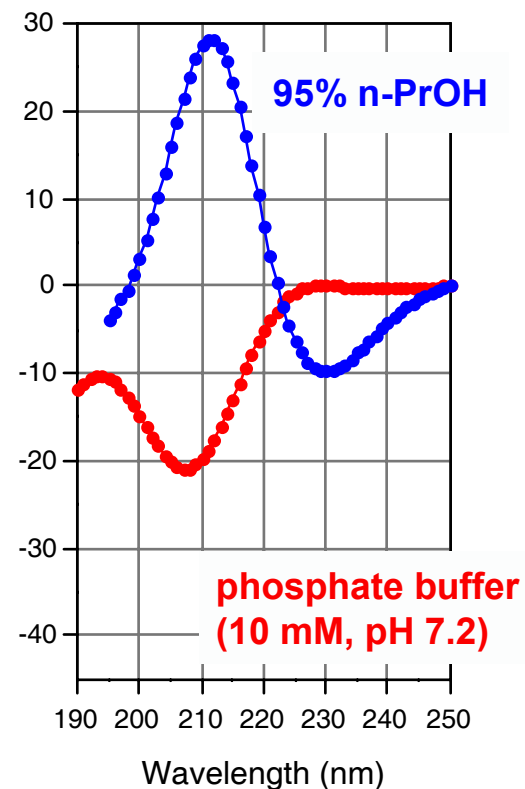


(4*R*)Azp stabilizes PPII

Ac-[Pro]₉-OH



Ac-[(4*S*)Azp]₉-OH

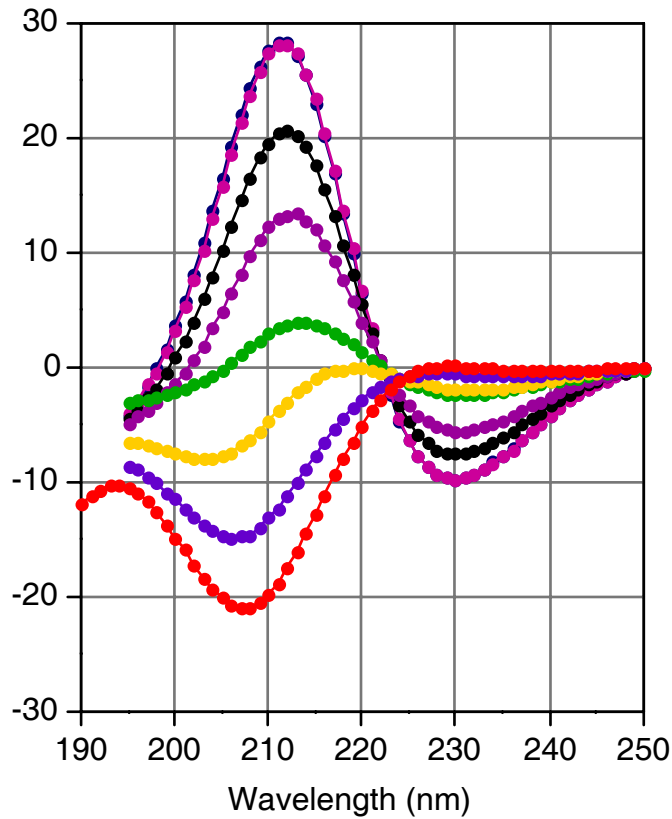


(4*S*)Azp stabilizes PPI

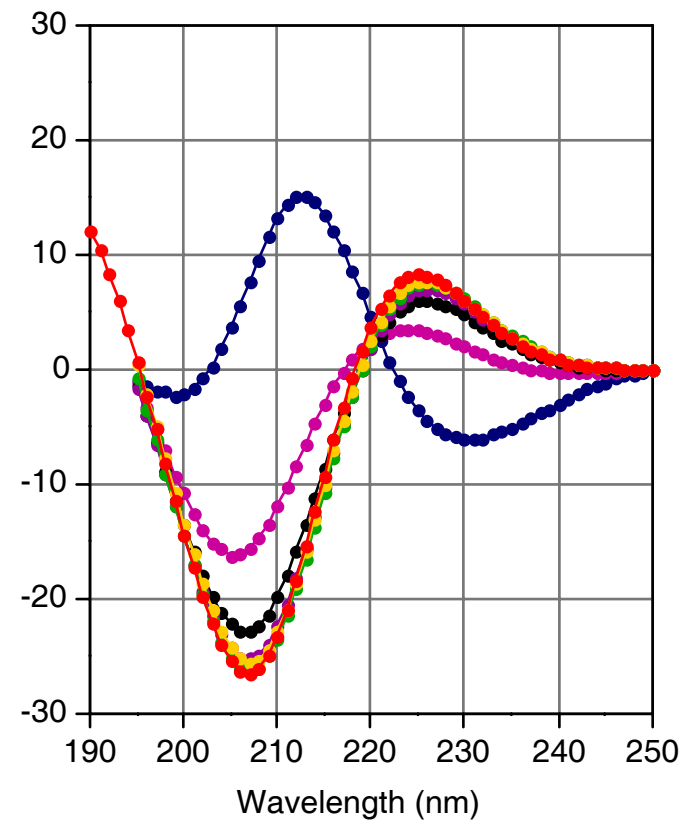
(4R)Azp stabilizes PPII, (4S)Azp stabilizes PPI - further support

CD-spectra in different mixtures of n-PrOH in phosphate buffer (10 mM, pH 7.2).

Ac-[(4S)Azp]₉-OH



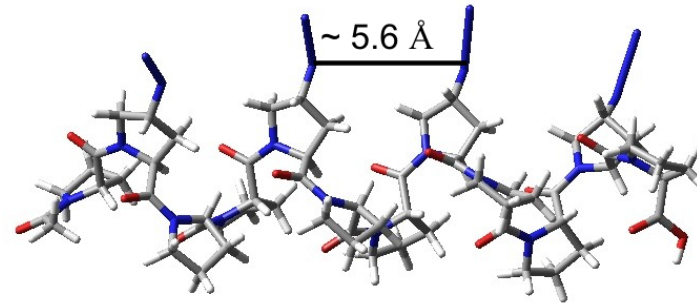
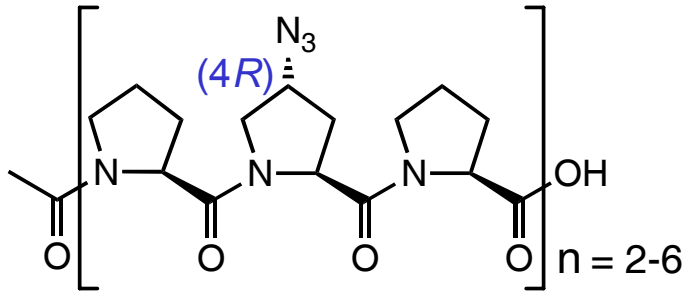
Ac-[(4R)Azp]₉-OH



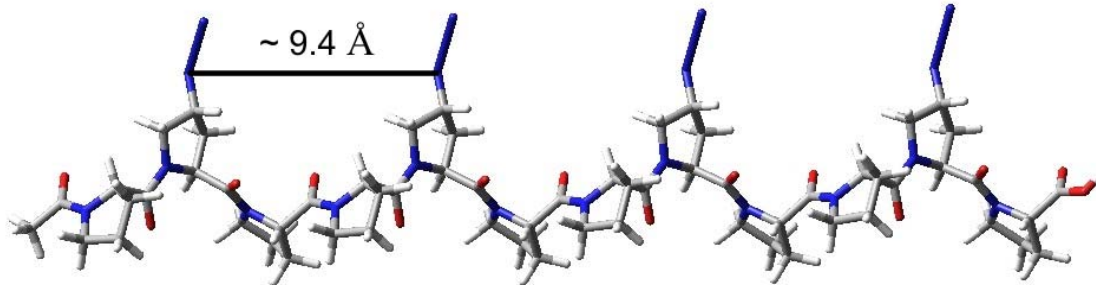
(4R)Azp and (4S)Azp are tools to tune the *s-cis*:*s-trans* ratio of Xxx-Pro bonds.

Azp containing Polyprolines as Functionalizable Scaffolds?

Models: Oligoprolines with (4*R*)Azp in every third position



PPI



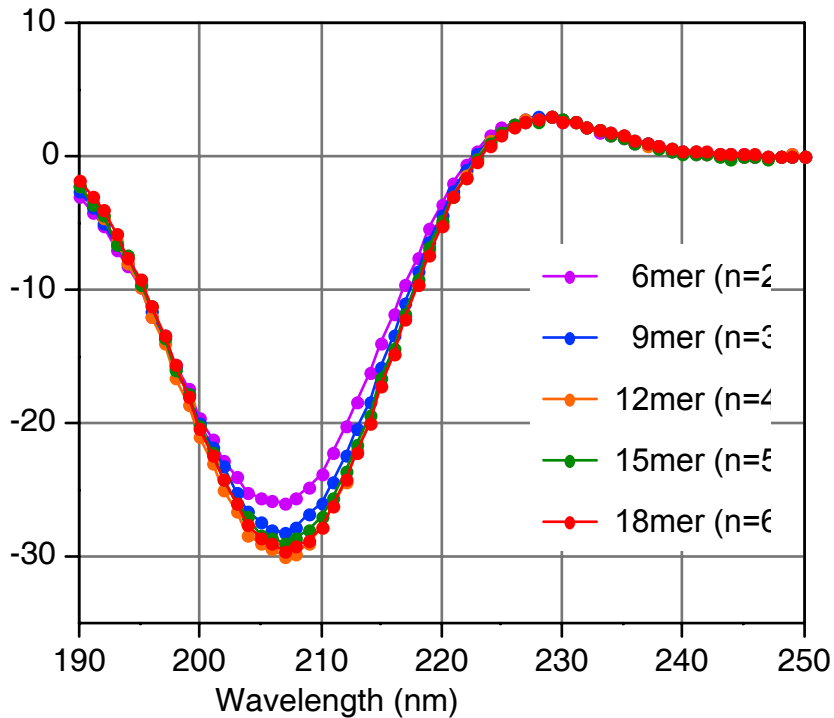
PPII

- What is the minimal chain length for conformationally well-defined PPI and PPII helices?
- Is functionalization possible?

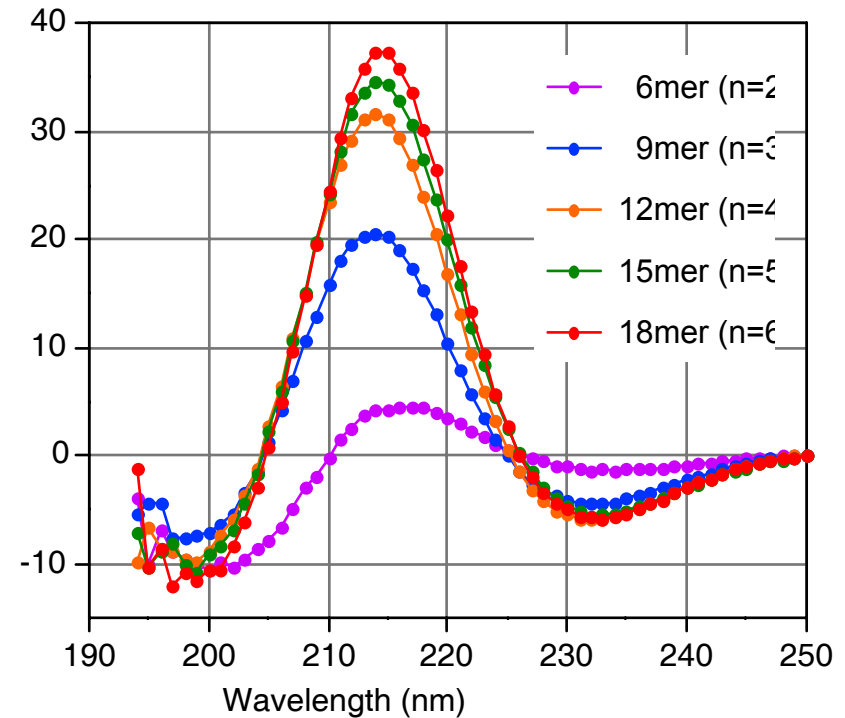
What is the minimal chain length for stable PPII and PPI helices?

Ac-[Pro-(4*R*)Azp-Pro]_n-OH (n=2-6), c = 630 mM/residue, T = 25°C

phosphate buffer (10 mM, pH 7.2)

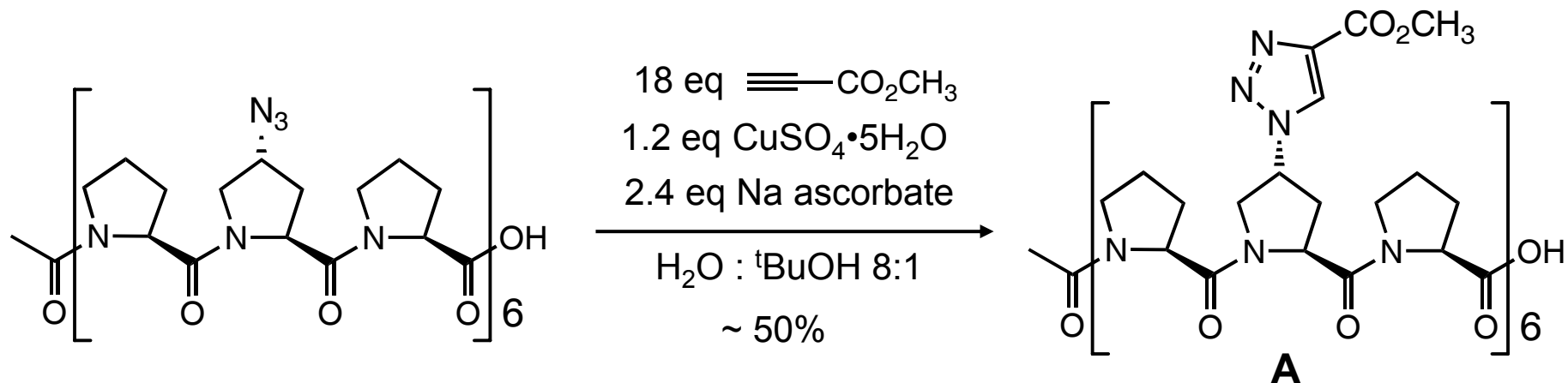


n-PrOH



Azp containing oligoprolines adopt already at short chain lengths conformationally well-defined PPI and PPII helices

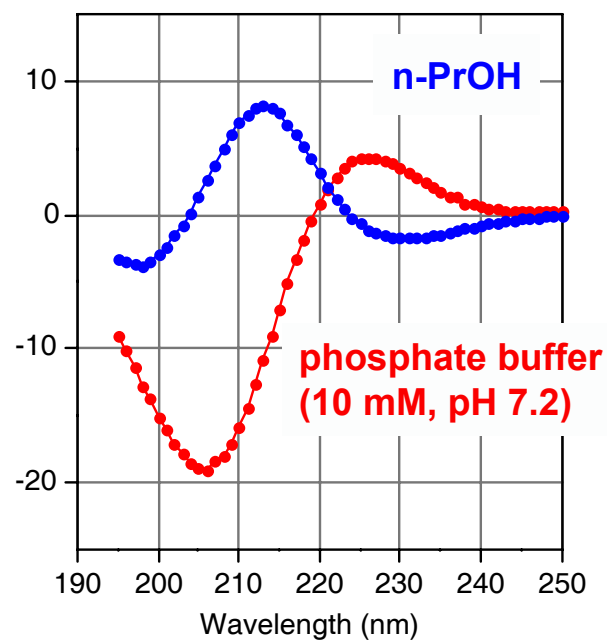
Functionalization via "Click Chemistry"



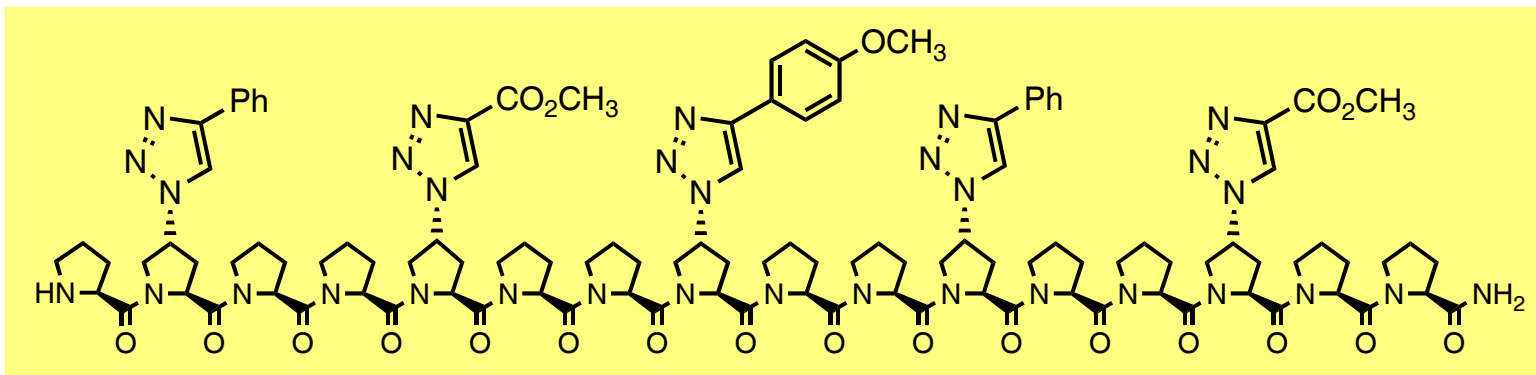
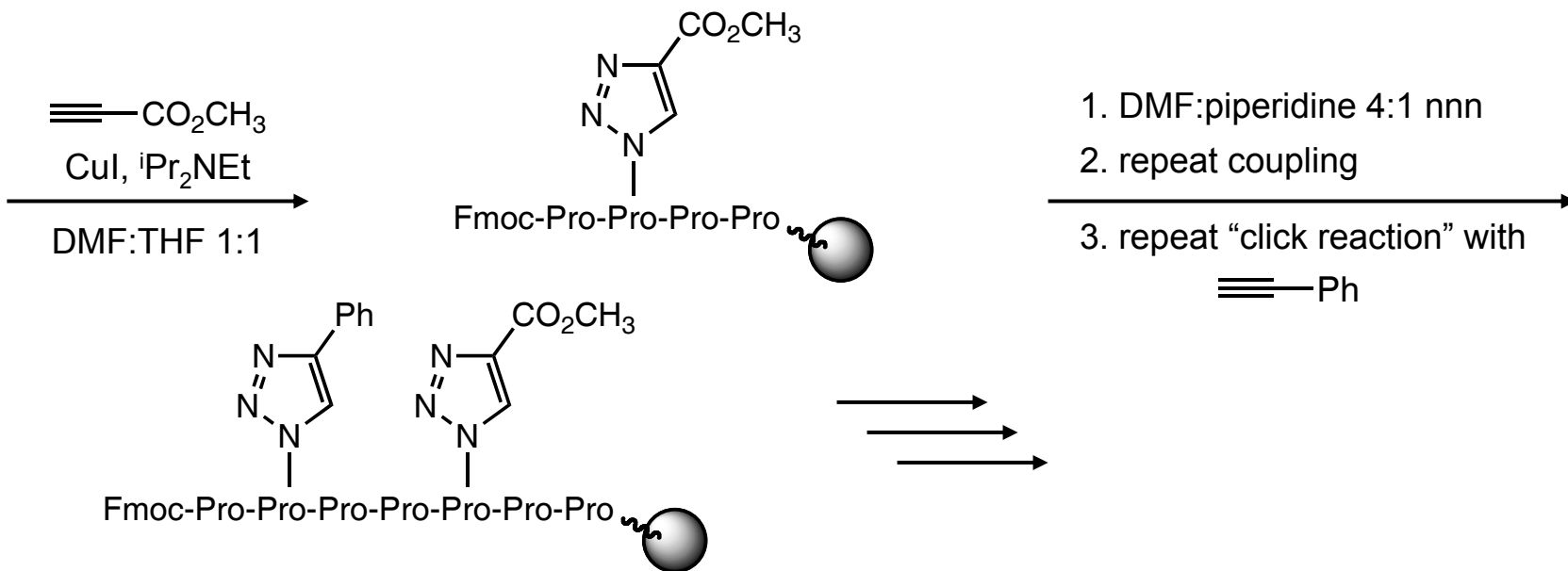
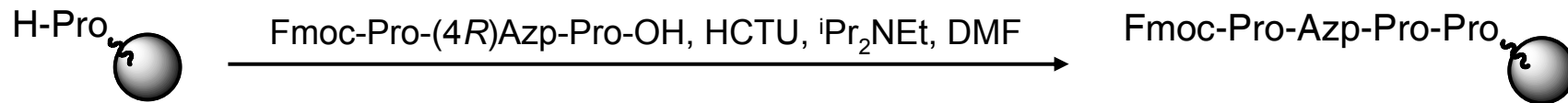
CD spectra of **A**

in n-PrOH: **PPI**

in phosphate buffer:
PPII



Differential Functionalization via "Click Chemistry"



Summary and Outlook

- (4*R*)Azp stabilizes, (4*S*)Azp destabilizes the PPII helix
 - an “azido gauche effect” and a $n \rightarrow \pi^*$ interaction determines the conformation of (4*R*)Azp
 - Azp containing oligoprolines are conformationally well-defined and switchable scaffolds that allow for further functionalization
-
- Use of Azp as tools to tune the conformation of the PPII helix
 - development of liquid crystalline materials, antibiotics,

Catalytic Activity

Selective Cleavage

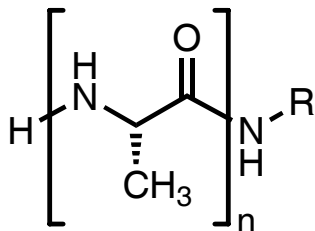
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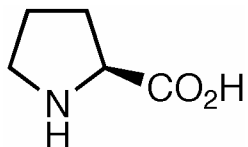
Polyproline

Peptidic Catalysts



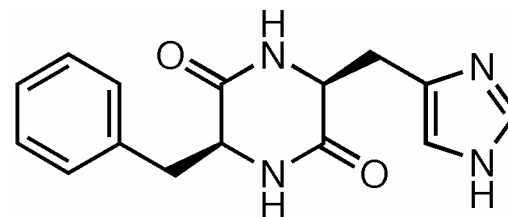
S. Juliá, 1983

A. Berkessel, 2001

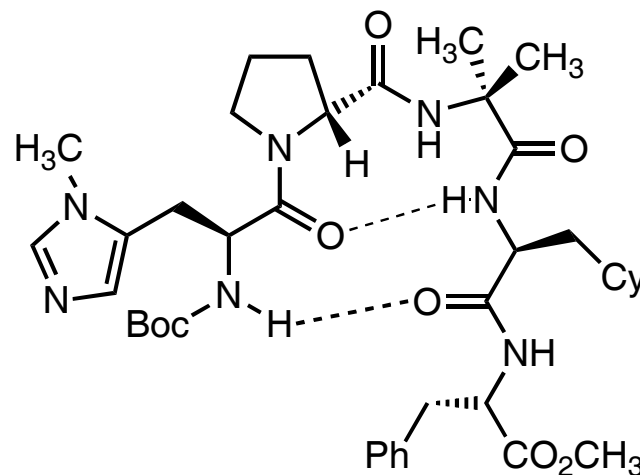


Hajos/Parrish/Wiechert, 1971

B. List, C.F. Barbas 2000,

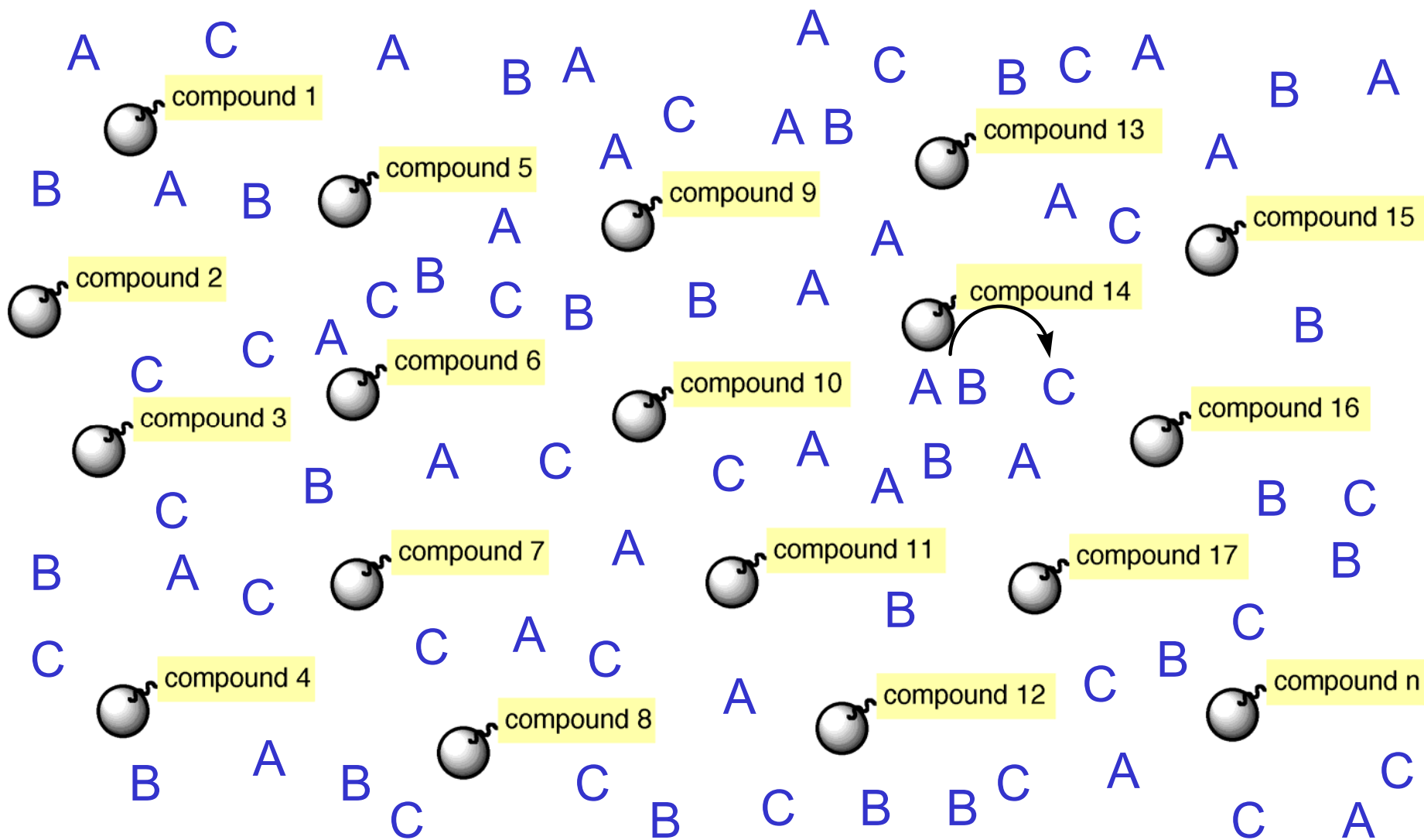


S. Inoue, 1992

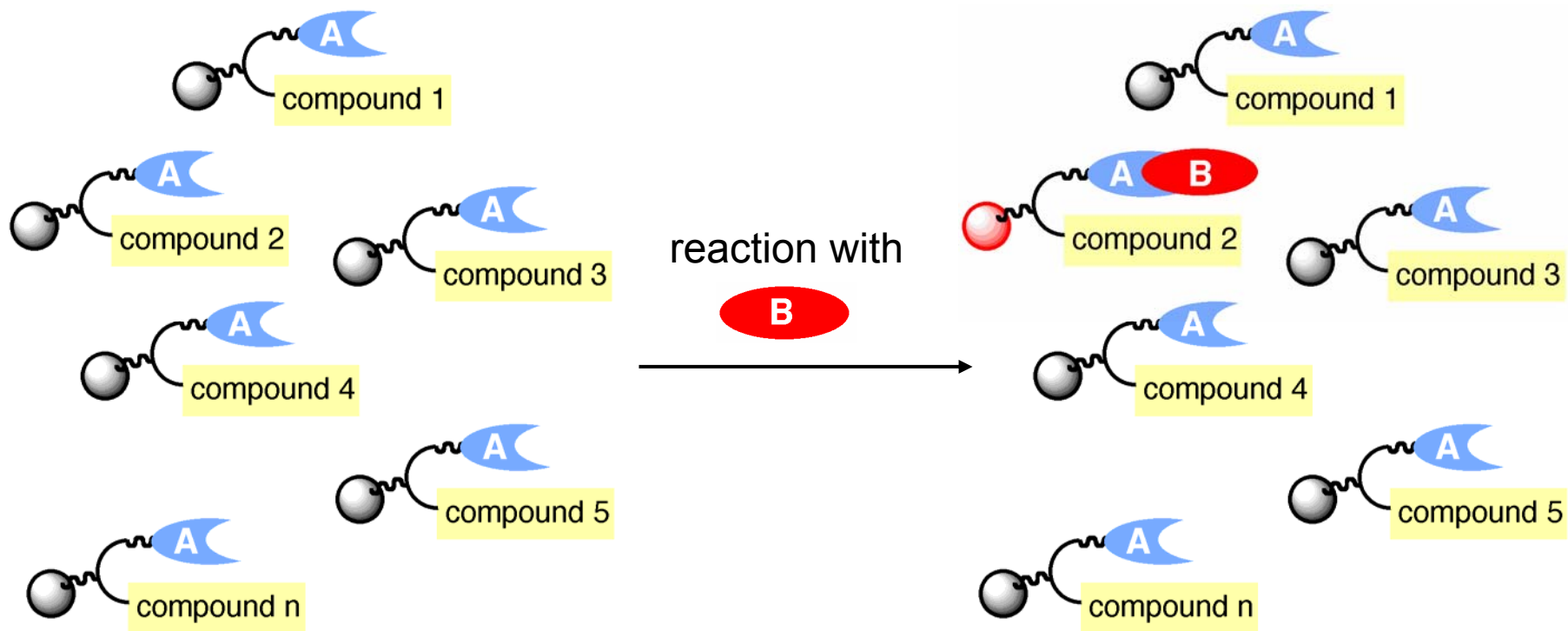


S.J. Miller, 1998

How to find the catalyst?

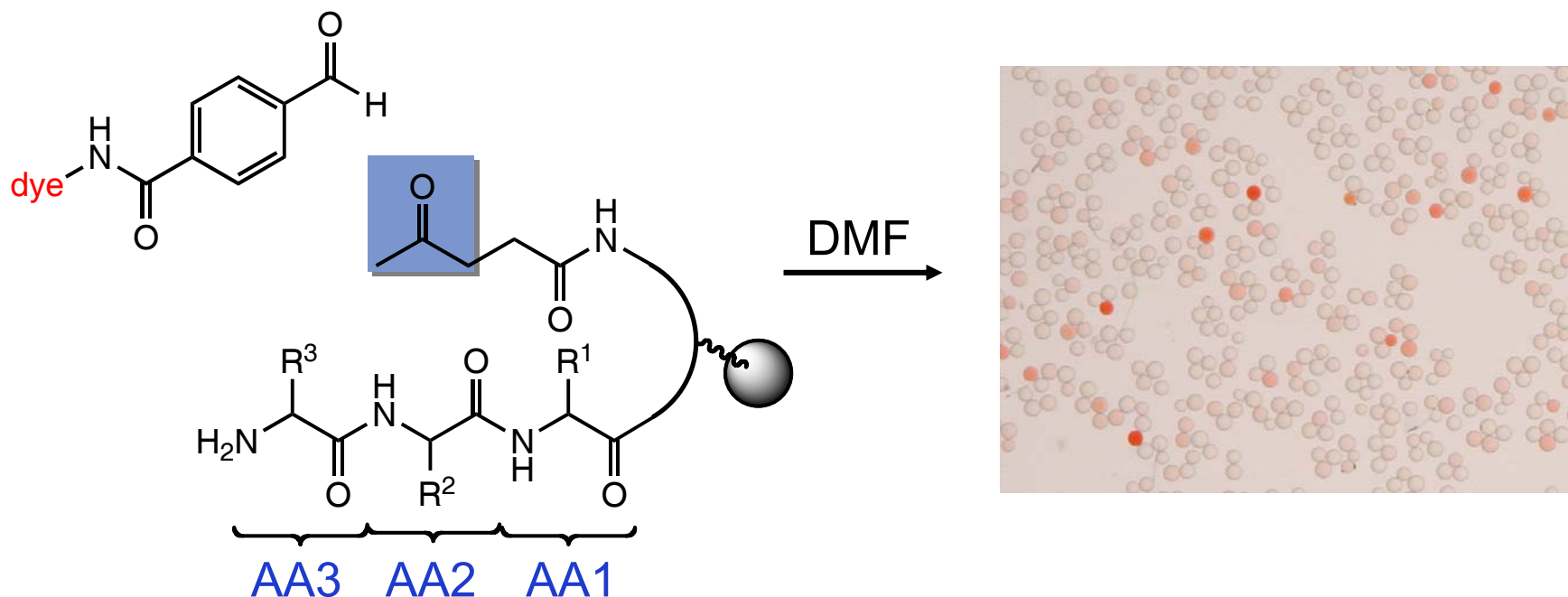


Catalyst-Substrate Co-immobilisation



Only *compound 2* is an active catalyst!

Screening Peptides for Catalysts of Aldol Reactions

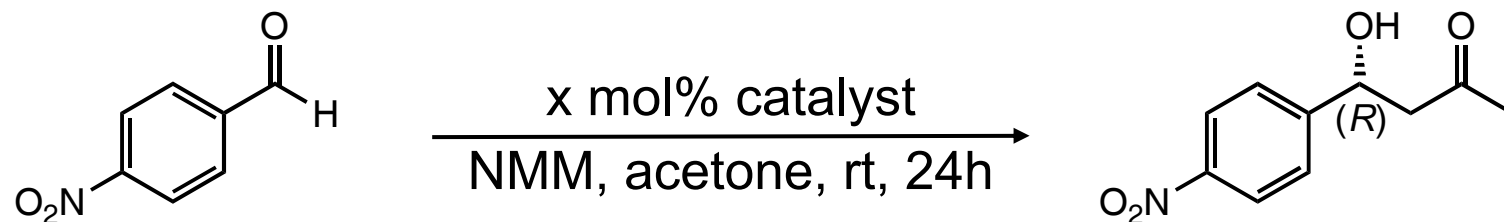


$AA_n = 15$ different D- and L-amino acids $\longrightarrow 15^3 = 3375$ tripeptides

consensus sequences: H-Pro-Pro-Asp-NHR

H-Pro-D-Ala-D-Asp-NHR

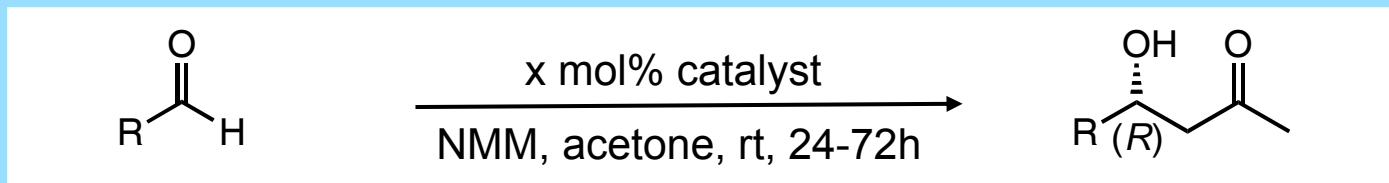
Solution Phase Experiments



catalyst	mol%	yield (%)	ee (%)	abs. conf.
H-Pro-D-Ala-D-Asp-NH ₂	10	73	70	R
H-Pro-D-Ala-D-Asp-NH ₂ (-20°C)	10	53	81	R
H-Pro-Pro-Asp-NH ₂	1	99	80	S
H-Pro-Pro-Asp-NH ₂ (-20°C)	5	98	90	S
H-Pro-OH ^a	30	68	76	R
H-Pro-OH (-20°C)	30	nd	71	R

^a B. List, *J. Am. Chem. Soc.* **2000**, *122*, 2395.

....other substrates

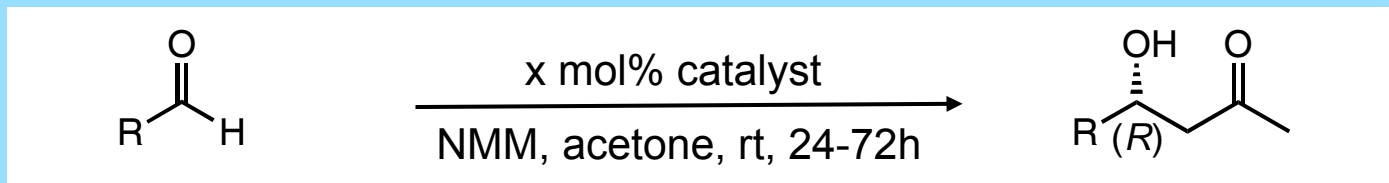


Product	H-Pro-OH ^a yield (%) / ee (%)	H-Pro-D-Ala-D-Asp-NH ₂ yield (%) / ee (%)	H-Pro-Pro-Asp-NH ₂ yield (%) / ee (%)
	68 /	73 /	99 /
	62 /	58 /	69 /
	63 /	56 /	66 /
	97 /	75 /	79 /
	22 /	24 /	28 /
	30 mol%	10 mol%	1 mol%

^a B. List, *J. Am. Chem. Soc.* **2000**, *122*, 2395.

P. Krattiger, R. Kovasy, J. D. Revell, S. Ivan, H. Wennemers, *Org. Lett.* **2005**, *7*, 1101.

....other substrates

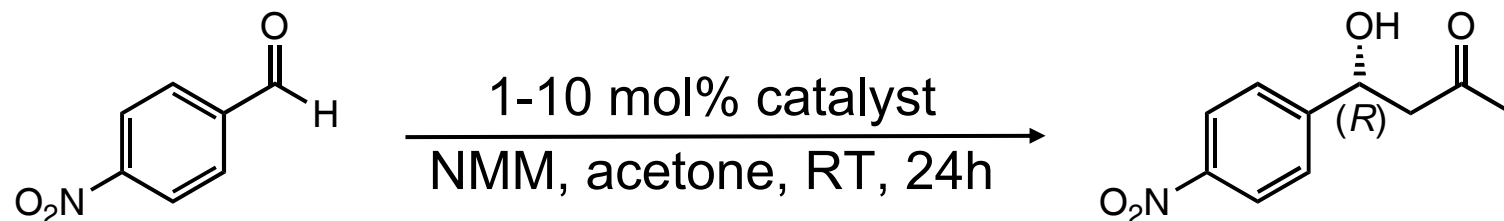


Product	H-Pro-OH ^a yield (%) / ee (%)	H-Pro-D-Ala-D-Asp-NH ₂ yield (%) / ee (%)	H-Pro-Pro-Asp-NH ₂ yield (%) / ee (%)
	76 (R)	70 (R)	80 (S)
	60 (R)	66 (R)	78 (S)
	84 (R)	83 (R)	82 (S)
	96 (R)	91 (R)	79 (S)
	36 (S)	70 (S)	73 (R)
	30 mol%	10 mol%	1 mol%

^a B. List, *J. Am. Chem. Soc.* **2000**, *122*, 2395.

P. Krattiger, R. Kovasy, J. D. Revell, S. Ivan, H. Wennemers, *Org. Lett.* **2005**, *7*, 1101.

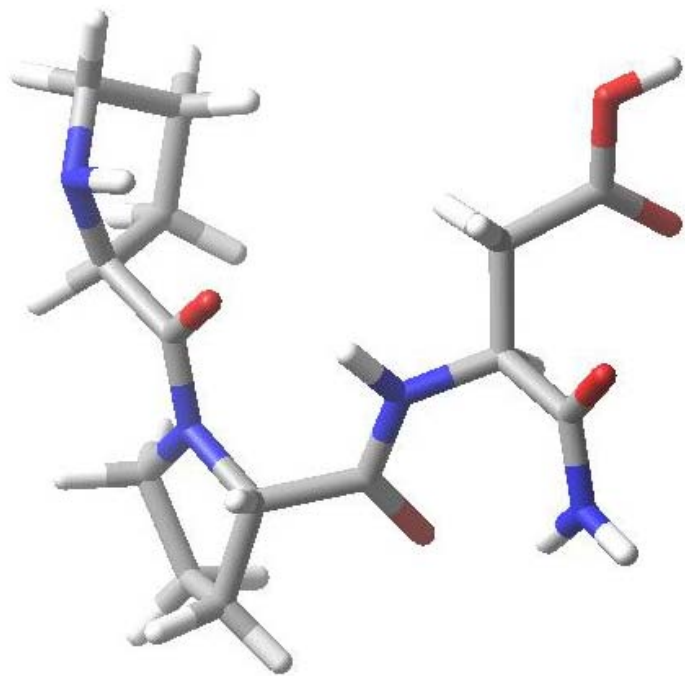
....other peptides



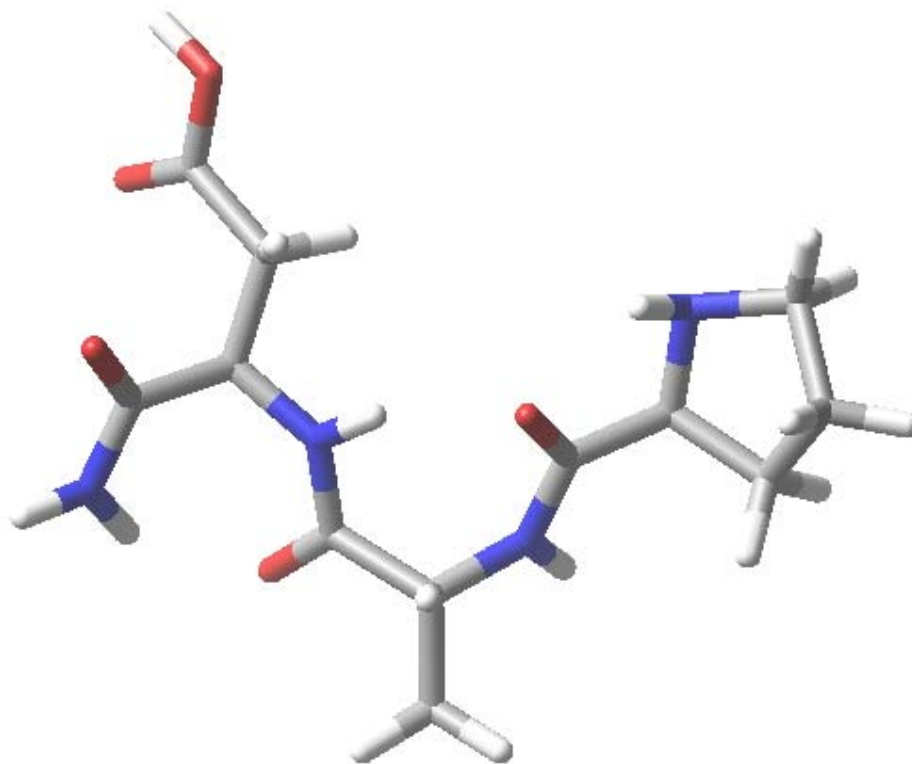
catalyst	yield (%)	ee (%)	abs. conf.
H-Pro-Pro-Asp-NH ₂	99	80	S
H-Pro-Pro-Asp-NH ₂ (-20°C)	98	90	S
H-Pro-Pro-Asn-NH ₂	23	50	S
H-Pro-Pro-Glu-NH ₂	60	62	S
H-D-Pro-D-Pro-Asp-NH ₂	27	11	R
H-Pro-Pro-OH	26	7	S
H-Pro-D-Ala-D-Asp-NH ₂	73	70	R
H-Pro-D-Ala-D-Asp-NH ₂ (-20°C)	53	81	R
H-Pro-Aib-D-Asp-NH ₂	73	62	R
H-Pro-D-Ala-OH	38	27	R

Conformational Analysis

H-Pro-Pro-Asp-NH₂



H-Pro-D-Ala-D-Asp-NH₂

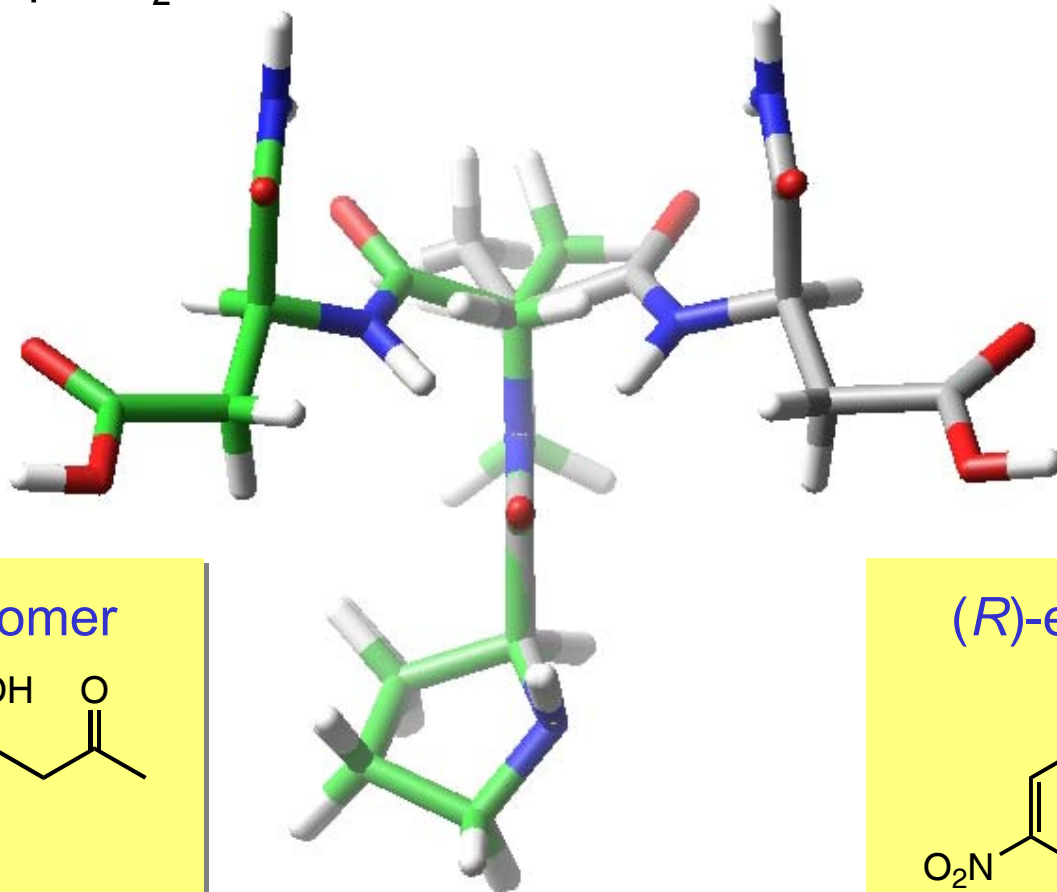


turn-conformations!

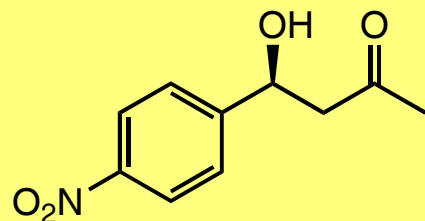
Conformational Analysis

H-Pro-Pro-Asp-NH₂

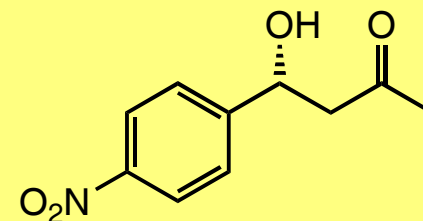
H-Pro-D-Ala-D-Asp-NH₂



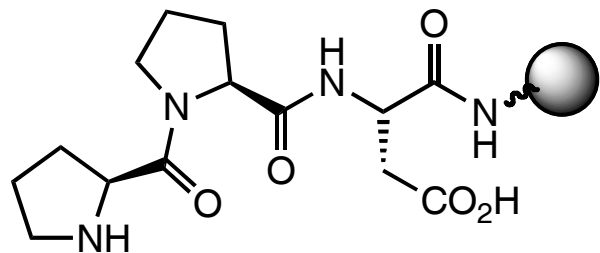
(*S*)-enantiomer



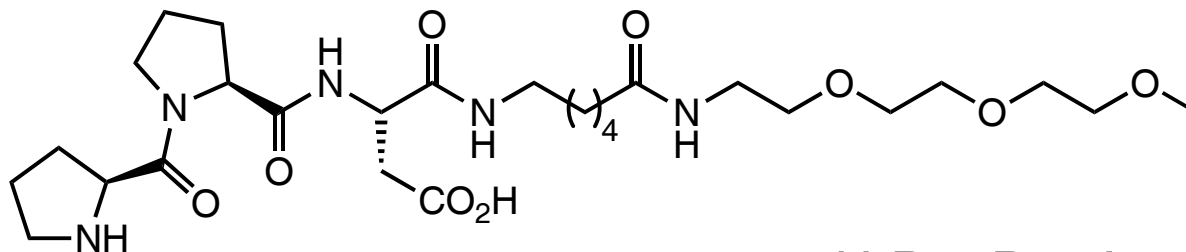
(*R*)-enantiomer



Solid-supported and Pegylated H-Pro-Pro-Asp-NHR

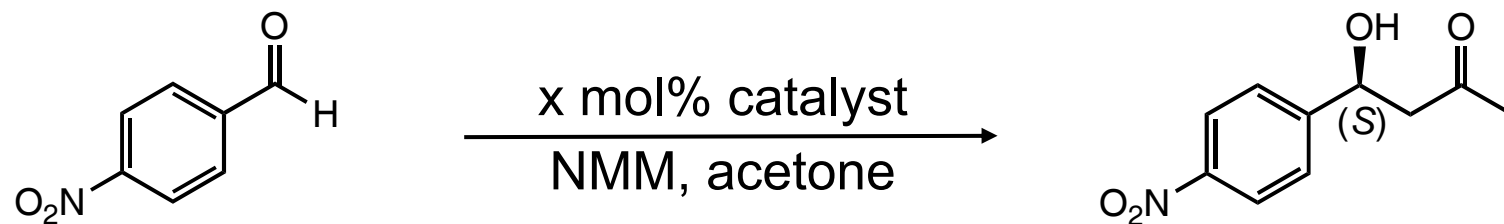


= H-Pro-Pro-Asp-resin (PPD-resin)



= H-Pro-Pro-Asp-Ahx-PEG

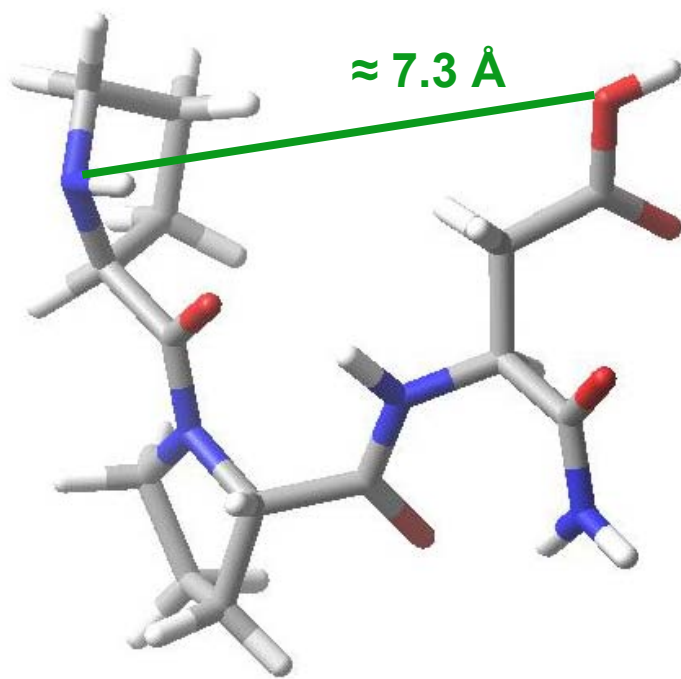
Pegylated H-Pro-Pro-Asp-NHR



catalyst		mol%	yield (%)	ee (%)	time
H-Pro-Pro-Asp-NH ₂		1	99	80	4
H-Pro-Pro-Asp-NH ₂	(-20°C)	5	98	90	24
H-Pro-Pro-Asp-Ahx-PEG		1	94	80	1
H-Pro-Pro-Asp-Ahx-PEG	(-20°C)	1	96	91	4
H-Pro-Pro-Asp-Ahx-PEG		0.5	93	80	4
H-Pro-Pro-Asp-Ahx-PEG	(-20°C)	0.5	95	91	8

Conformational Analysis

H-Pro-Pro-Asp-NH₂



Distance between the secondary amine and the carboxylate is ≥ 3 Å longer compared to proline.



H-Pro-Pro-Asp-NH₂ a catalyst for 1,4-additions?

Outlook

- **substrate/reaction scope, mechanism**
- **What is the optimal size of a catalytically active peptide?**
- **Peptides as catalysts for other reactions, e.g. Cycloadditions?
....non-covalent catalysts**

Acknowledgements

Current Coworkers

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Catalytic Activity

Selective Cleavage

Peptides

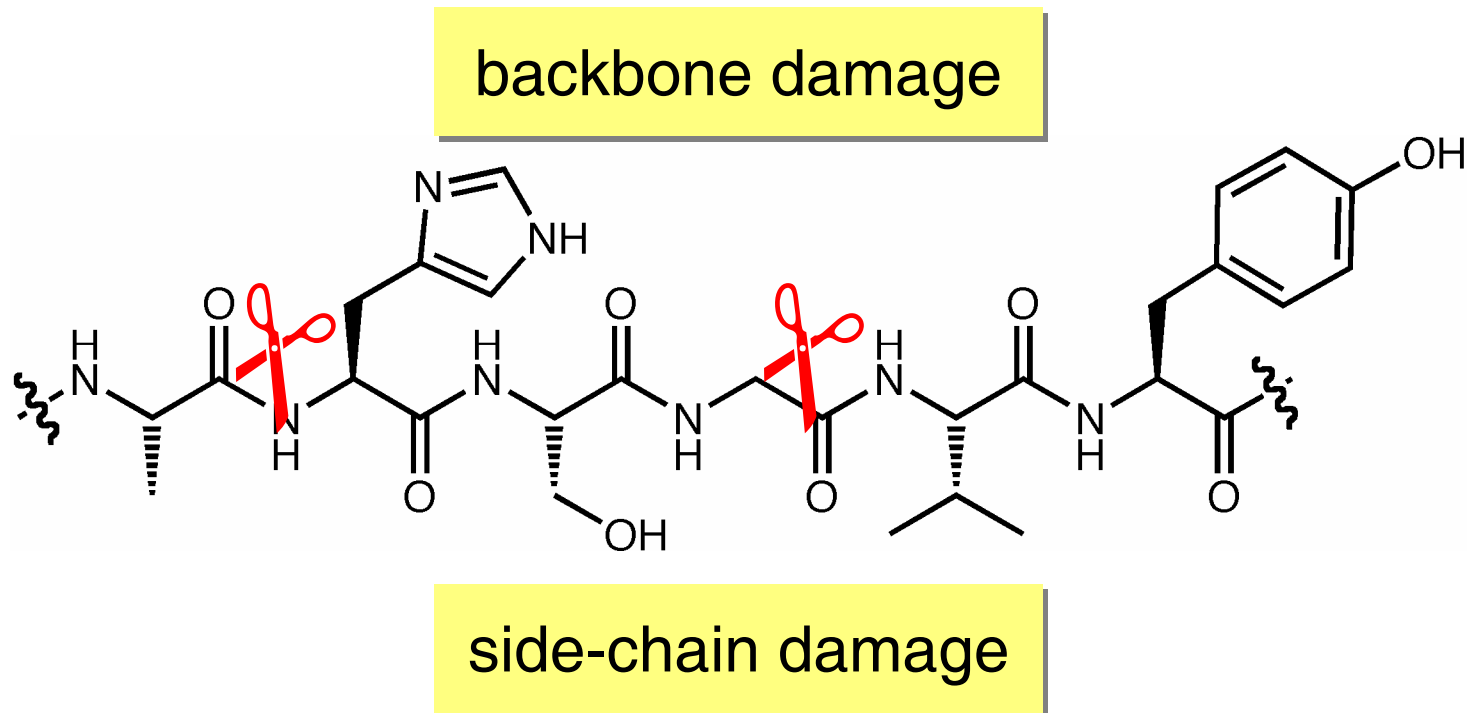
Molecular Recognition

Stereoelectronic Effects

Polyproline

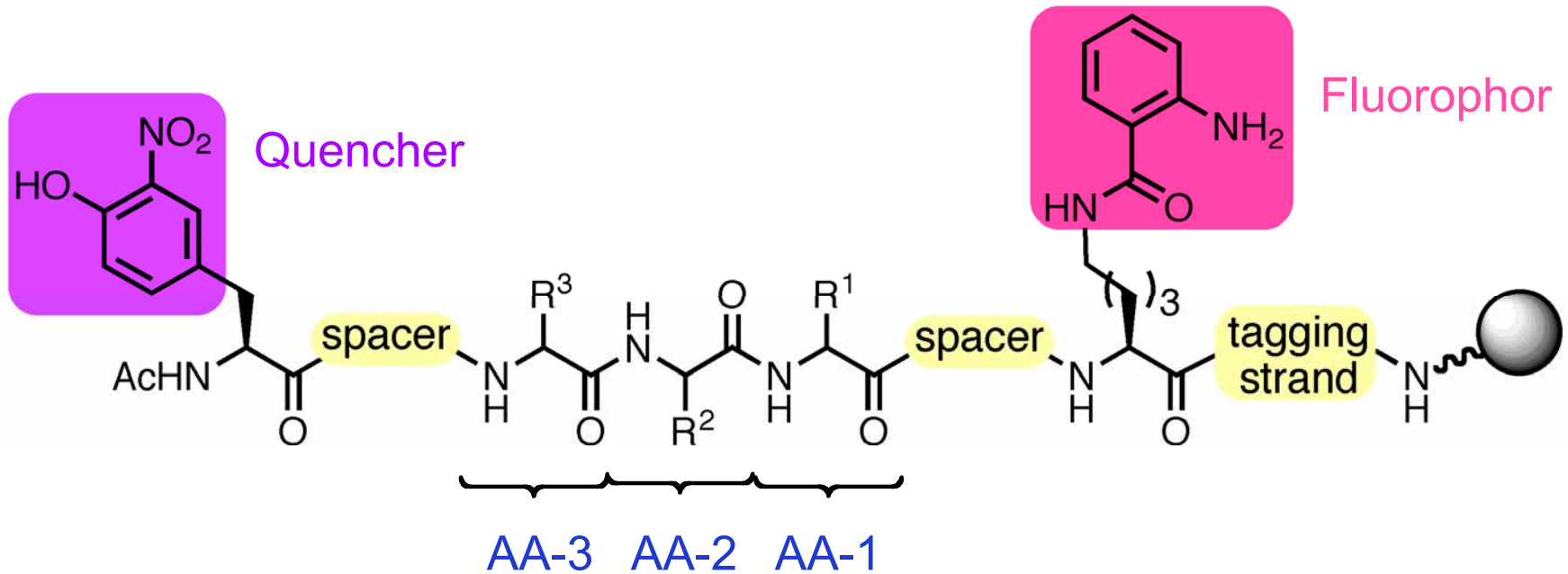
Protein Damage

for example, Fenton reaction: Fe^{3+} , reducing agent, H_2O_2



Is the cleavage sequence dependent?

Fluorophor-Quencher-Peptide Library

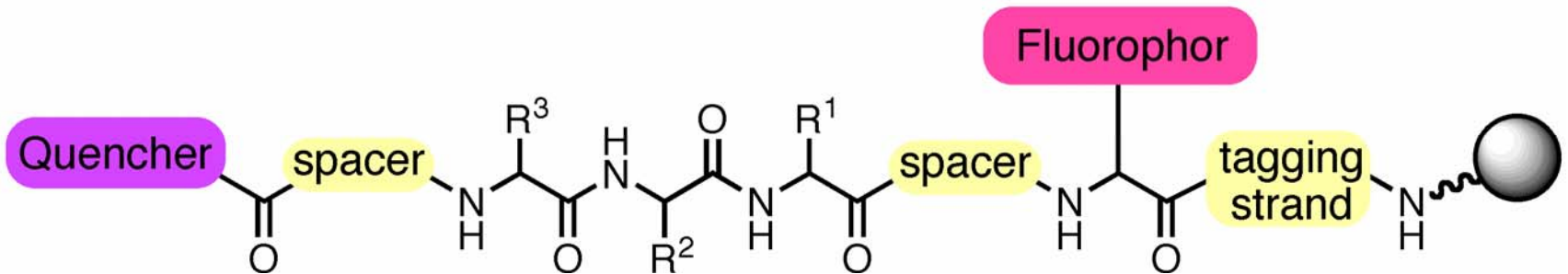


AA-n = 31 different D- and L-amino acids



$31^3 = 29791$ peptides

Cleavage under “Fenton Conditions”

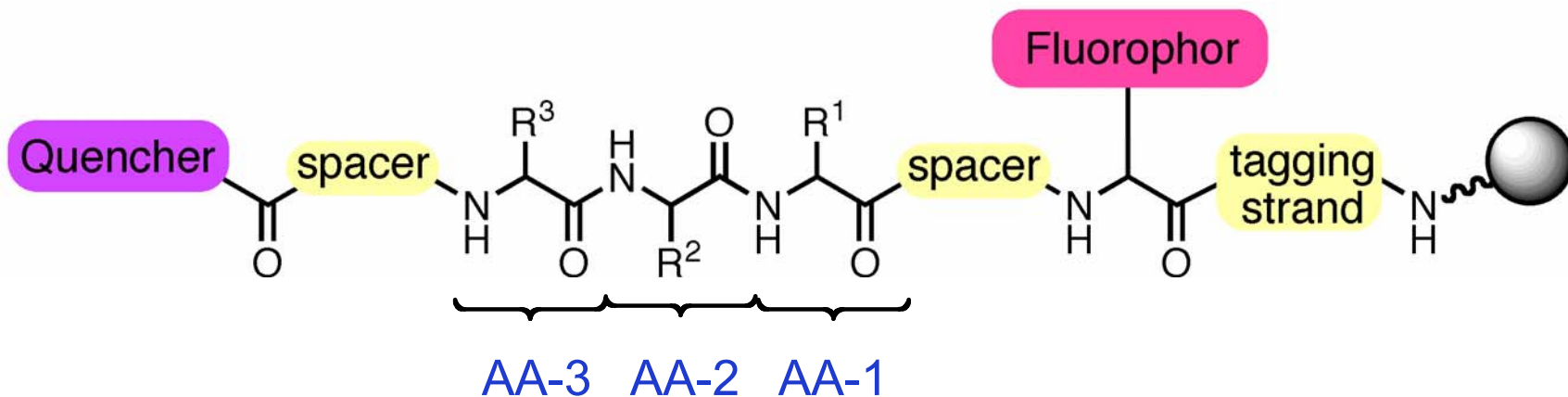


HEPES buffer pH 5

1. FeCl_3
2. H_2O_2 , NaAscorbate



Cleavage Selectivities



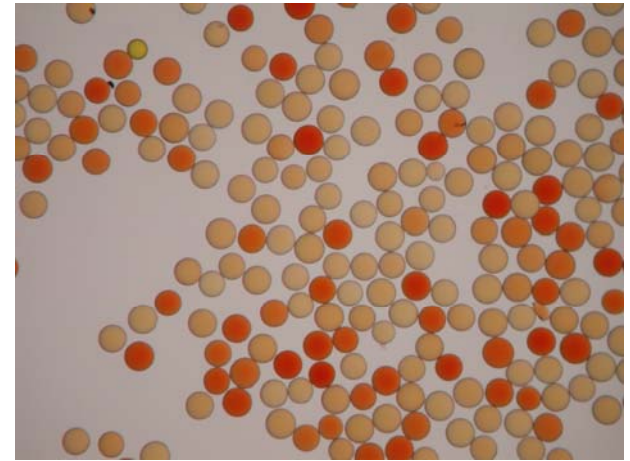
AA-3	AA-2	AA-1	frequency found
L/D-Asp/Glu	L/D-Asp/Glu	X	34%
X	L/D-Asp/Glu	L/D-Asp/Glu	46%
L/D-Asp/Glu	X	L/D-Asp/Glu	13%

Fe-Peptide Complexes

studies on solid support

tripeptide library complexed with FeCl_3
followed by staining with KSCN

red beads: carboxylic acid-rich peptides

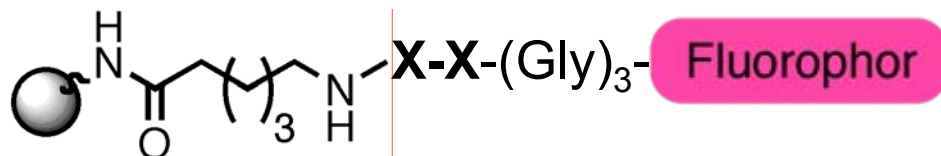


studies in solution

$\text{Ac-Glu-Glu-NHCH}_2\text{CH}_2\text{CH}_3$
binding to FeCl_2 : $K \approx 10^3\text{-}10^4 \text{ M}^{-1}$

$\text{Ac-Gly-Gly-NHCH}_2\text{CH}_2\text{CH}_3$
no binding to FeCl_2 observed

Cleavage Studies with Solid-supported Peptides



X-X = Glu-Glu

X-X = Gly-Gly

1. FeCl₃

2. H₂O₂, NaAsc

3. MnO₂ quench

filter and measure
fluorescence of solution

