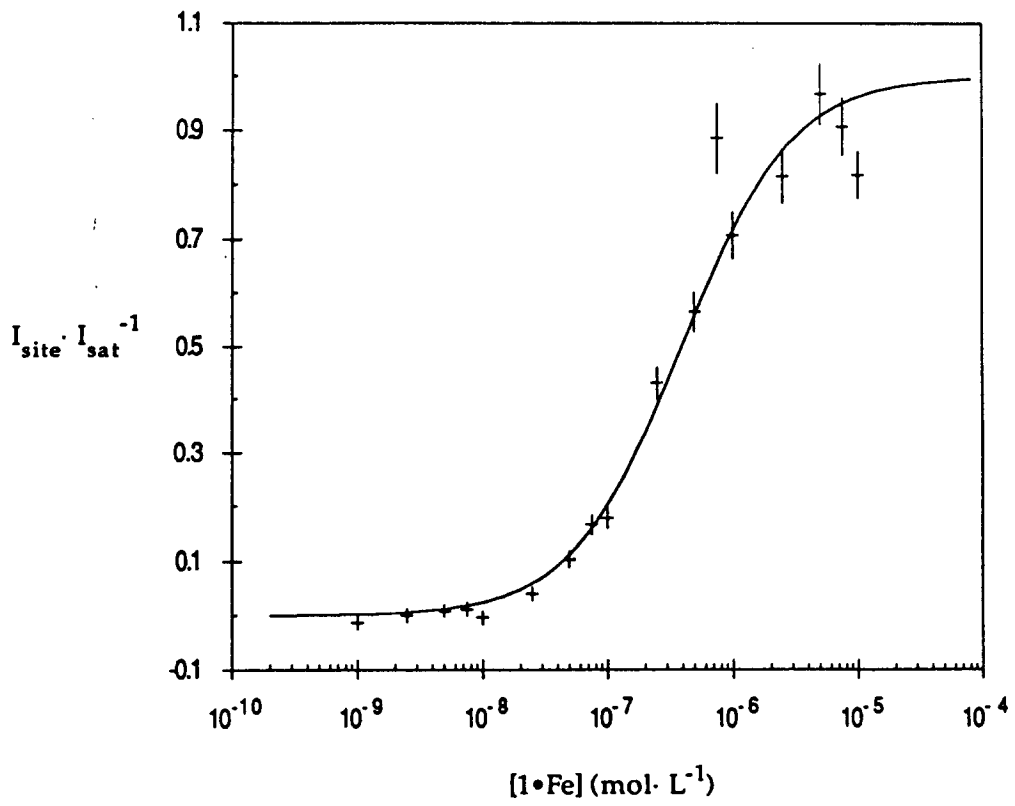


- 1 5'- *TTTTTCTCTCTCTCT
- 2 5'- *TTTTTCTCTCTCT
- 3 5'- *TTTTTCTCTCT
- 4 5'- *TTTTTCTCTCCCTCT
- 5 5'- *TTTTTCTCTTTCTCT
- 6 5'- TTTTTCTCTCTCTCT



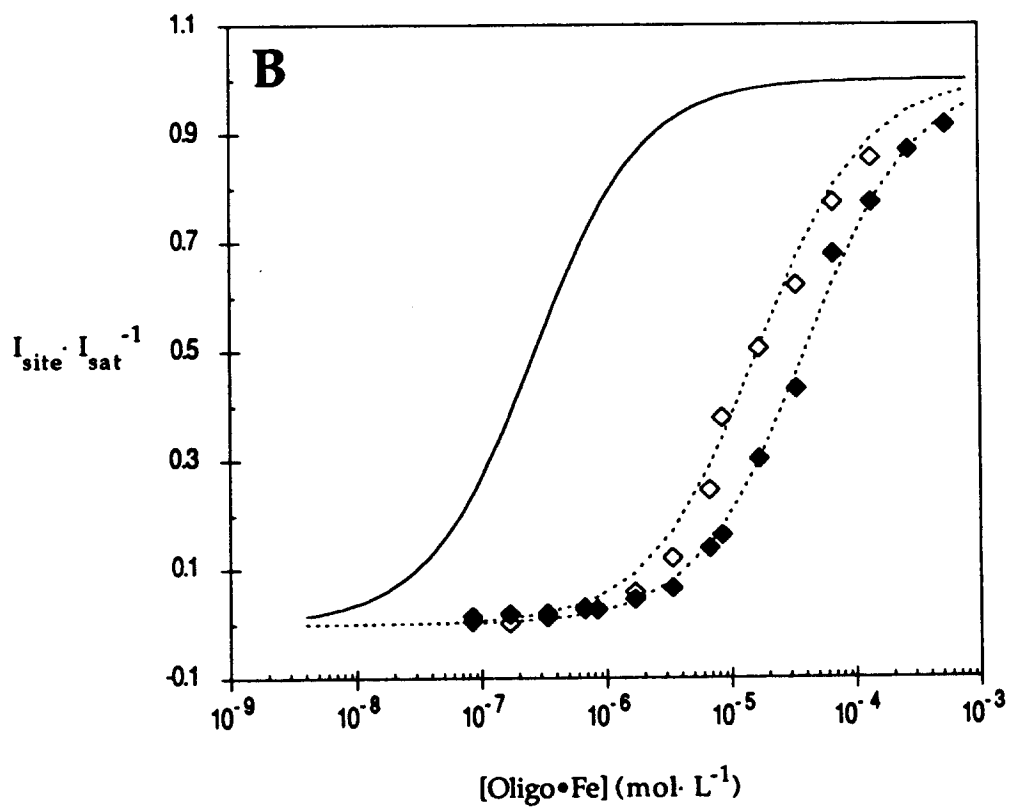
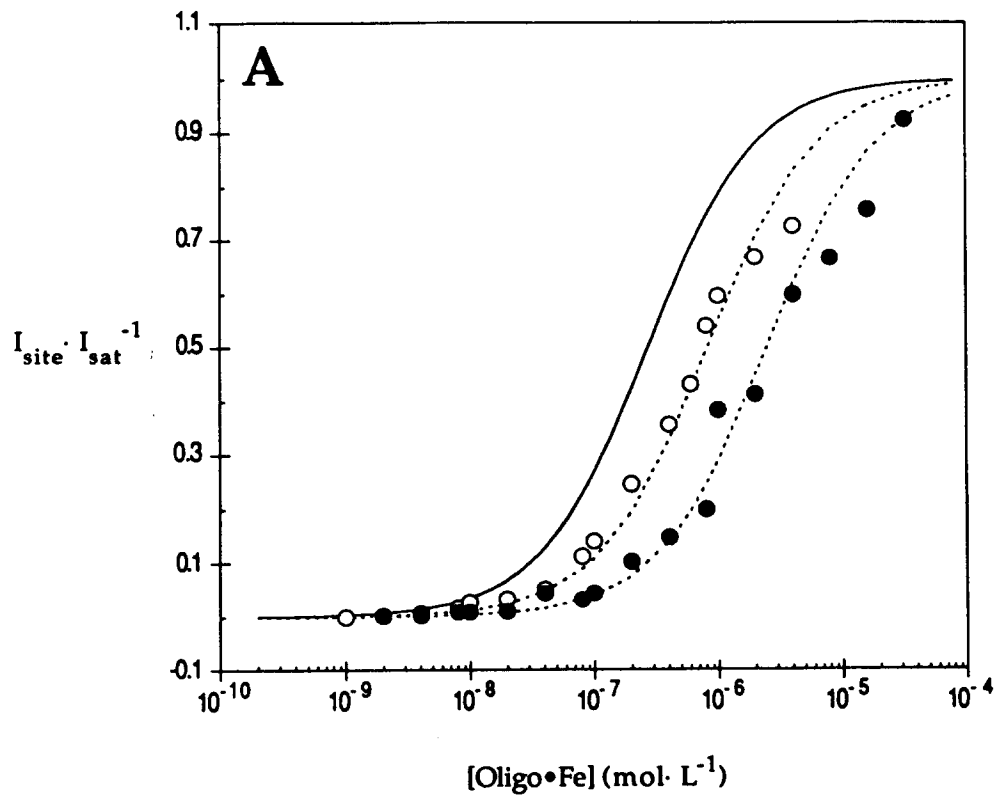
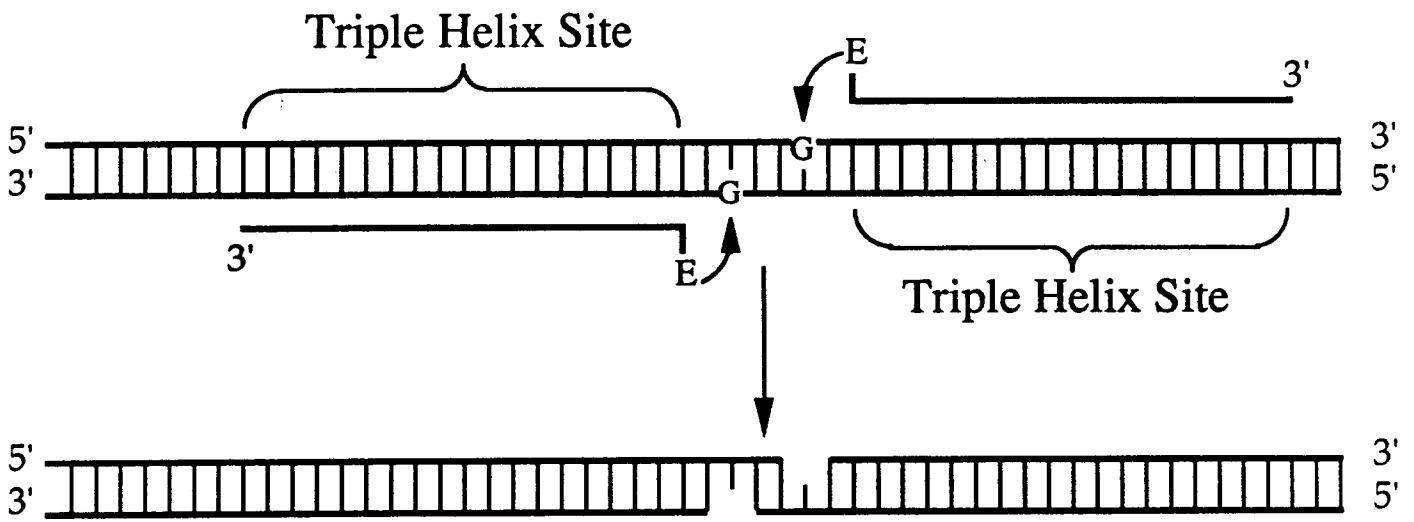
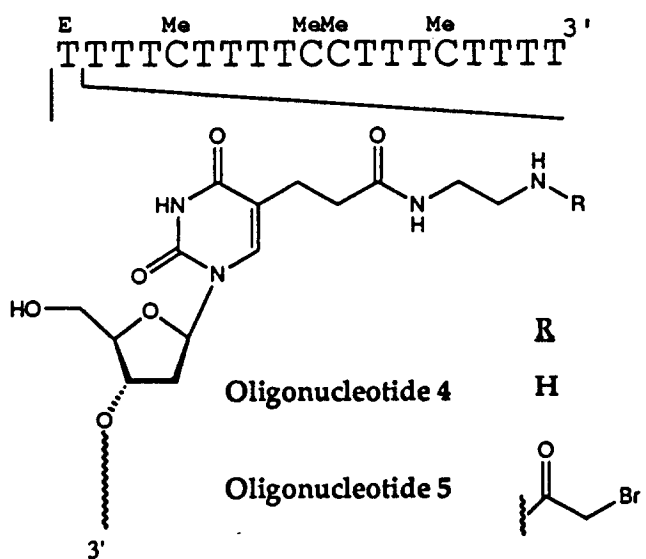
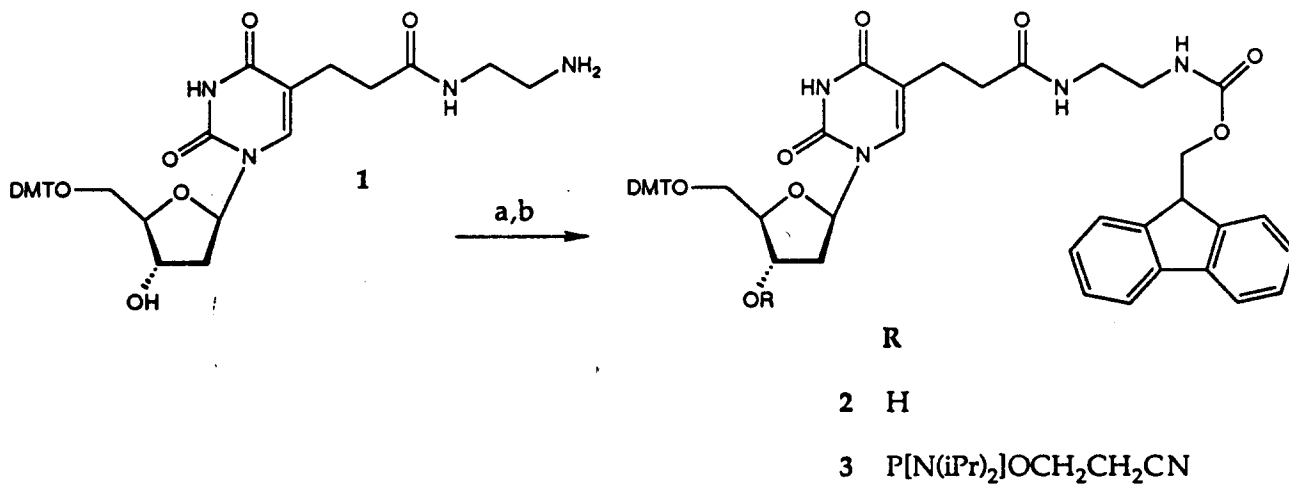


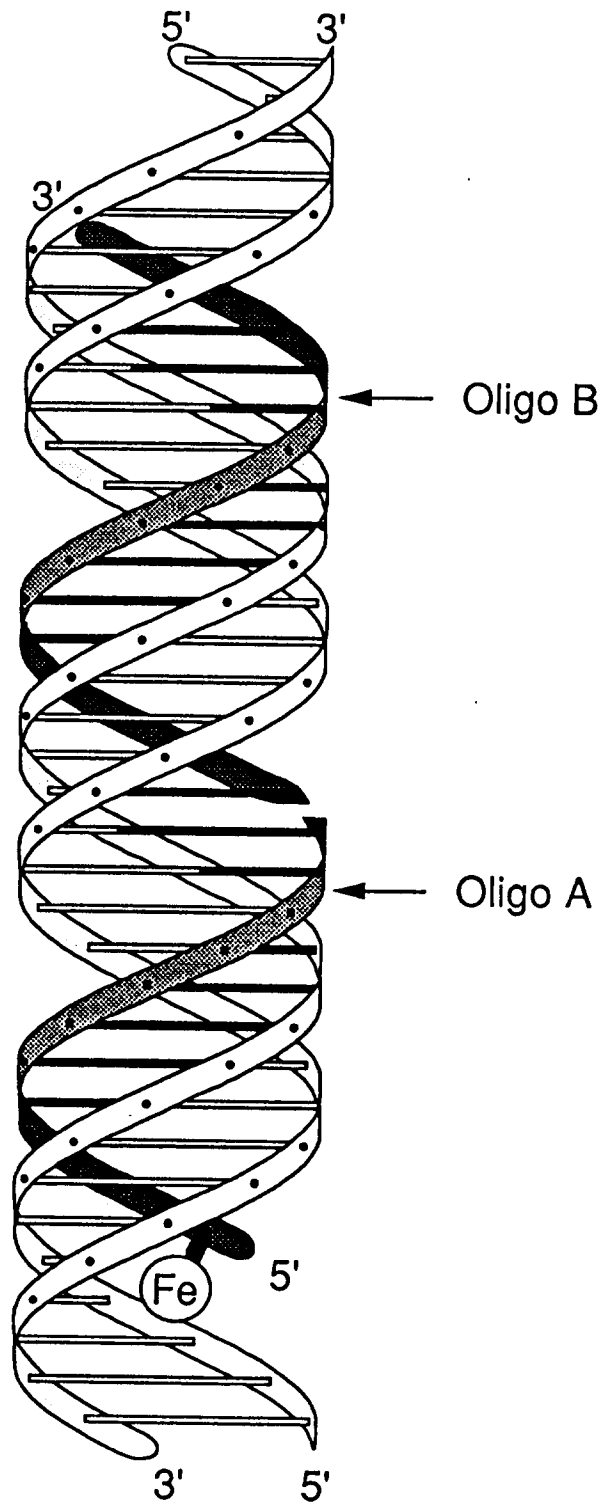
Table II. Length and Sequence Mismatch Effects on K_T at pH 7.0 and 24 °C.^a

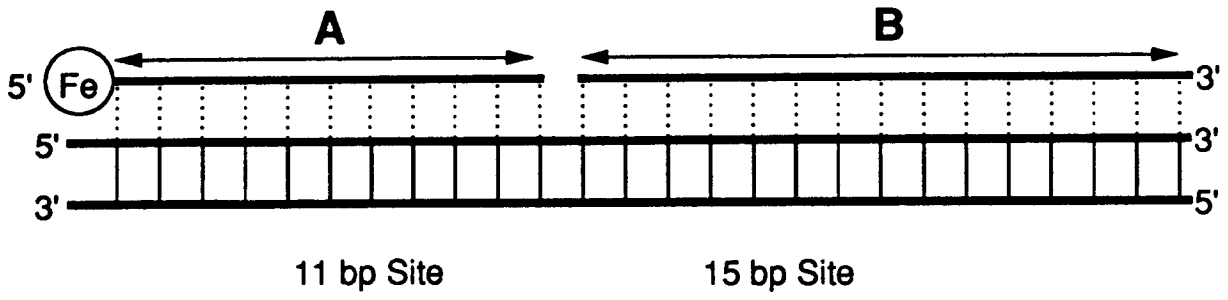
Oligo-nucleotide	Length (nts)	No. T's (nts)	No. C's (nts)	Base Mismatch	K_T (M^{-1})	ΔG_T (kcal·mol ⁻¹)
1	15	10	5	-	3.7 (± 1.1) $\times 10^6$	-9.0 (± 0.2)
2	13	9	4	-	1.8 (± 0.4) $\times 10^6$	-8.5 (± 0.1)
3	11	8	3	-	6.2 (± 3.8) $\times 10^5$	-7.9 (± 0.6)
4	15	9	6	C \circ AT	6.1 (± 2.3) $\times 10^4$	-6.5 (± 0.2)
5	15	11	4	T \circ GC	2.5 (± 0.1) $\times 10^4$	-6.0 (± 0.1)

^a Values reported in the table are mean values measured from affinity cleavage titration experiments performed in Association buffer (100 mM Na⁺, 1 mM spermine tetrahydrochloride, 50 mM Tris-acetate, pH 7.0).









Quantitative Affinity Cleavage Reaction Conditions

25 mM TRISOAc, pH 7.0; 10 mM NaCl; 1 mM Spermine;

100 μ M bp CT DNA; 4 mM DTT; ~10,000 cpm Target Site.

24 hr Room Temperature Preincubation;

6 hr Cleaving Reactions.

Oligonucleotide Concentrations used:

11mer: 100 pM \rightarrow 20 μ M;

15mer: 1 μ M.

Binding Constants Obtained from Quantitative Affinity Cleavage Experiments: 11mer + 15mer

11mer alone:

$$K_a = 1.16 (\pm 0.70) \cdot 10^6 \text{ M}^{-1}; K_d = 862 \text{ nM}$$

15mer alone:

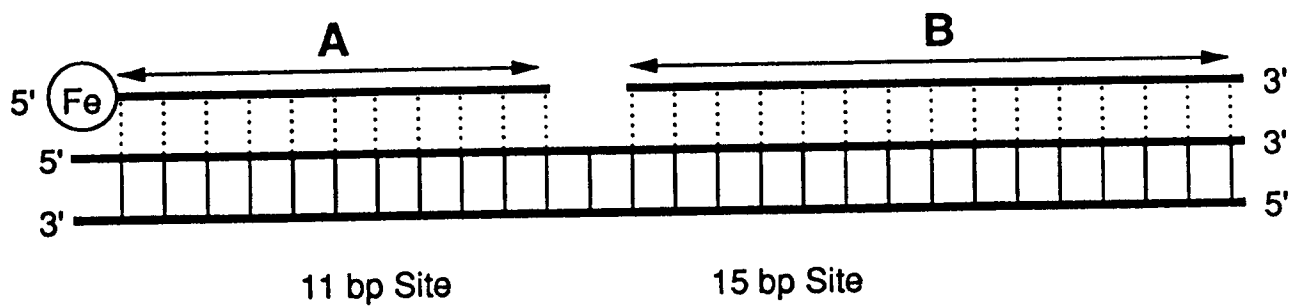
$$K_a = 1.26 (\pm 0.52) \cdot 10^7 \text{ M}^{-1}; K_d = 79 \text{ nM}$$

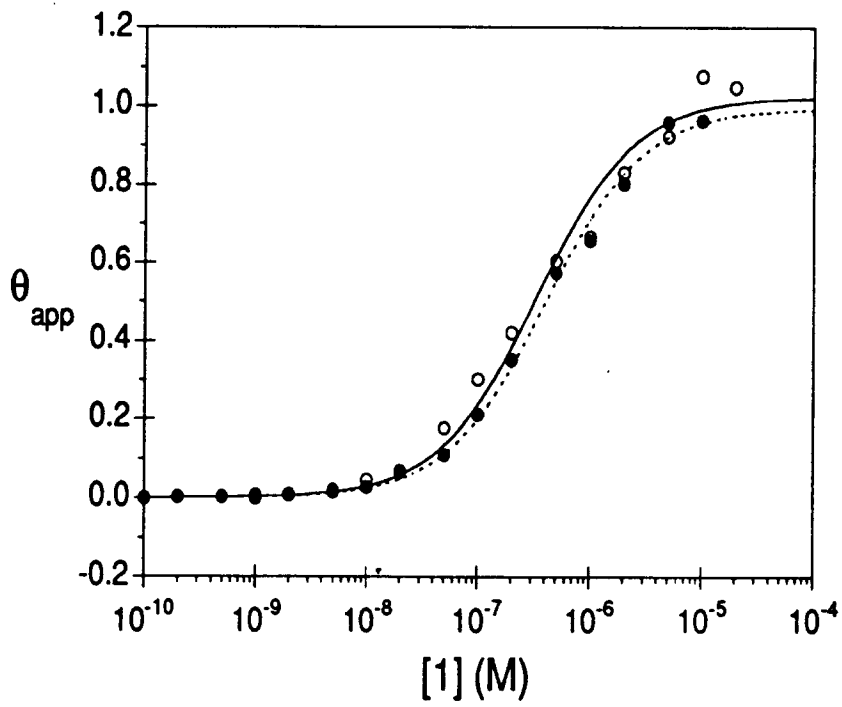
11mer + 15mer (1 μM):

$$K_d = 2.32 (\pm 1.14) \cdot 10^7 \text{ M}^{-1}; K_d = 43 \text{ nM}$$

$$\Delta\Delta G = 1.76 \text{ kcal/mol}$$

\therefore 20-fold Enhancement





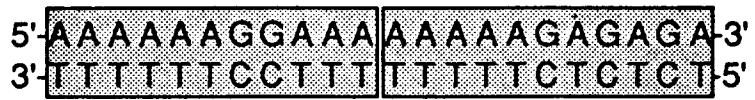
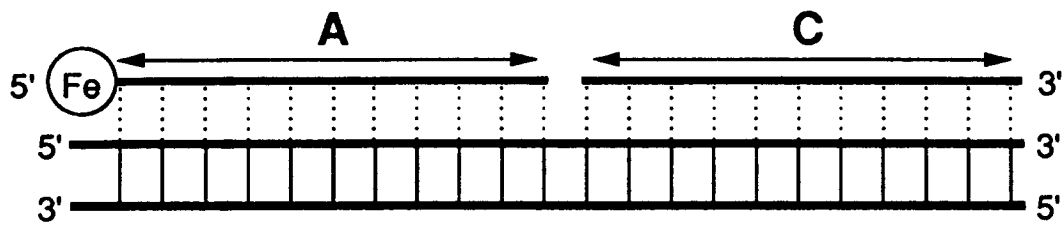
*Binding Constants Obtained from Quantitative Affinity
Cleavage Experiments: 11mer + 15mer in the Presence
of a One Base Pair Gap*

11mer alone:

$$K_a = 2.89 (\pm 0.35) \cdot 10^6 \text{ M}^{-1}; K_d = 346 \text{ nM}$$

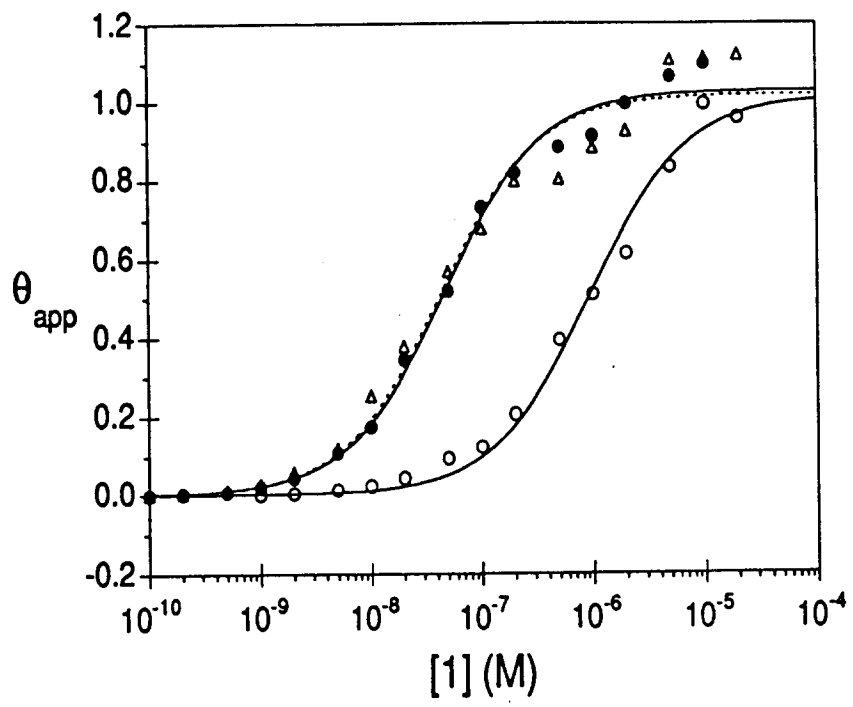
11mer + 15mer (1 μM):

$$K_d = 2.51 (\pm 0.51) \cdot 10^6 \text{ M}^{-1}; K_d = 398 \text{ nM}$$



1 5'-*^{MeMe}TTTTTCCTTT (A)

3 5'-*^{Me Me Me}TTTTTCTCTCT (C)



*Binding Constants obtained from Quantitative Affinity
Cleavage Reactions: 11mer + 11mer*

11mer alone:

$$K_a = 1.16 (\pm 0.70) \cdot 10^6 \text{ M}^{-1}; K_d = 862 \text{ nM}$$

11mer + 15mer:

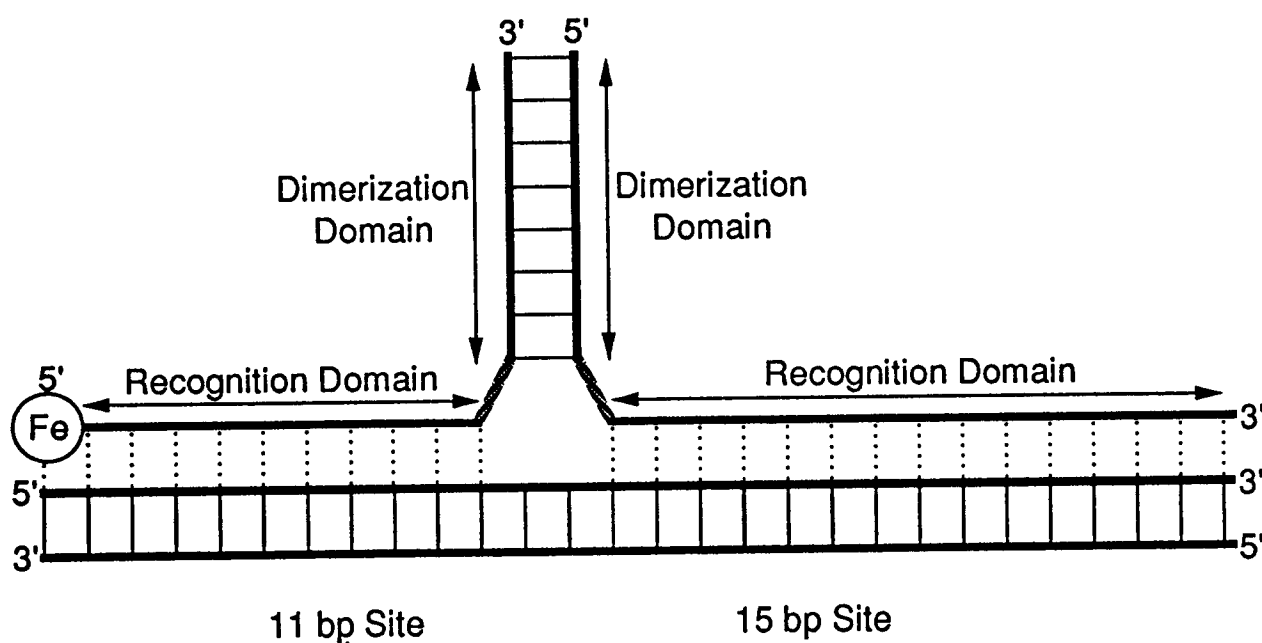
$$K_d = 2.32 (\pm 1.14) \cdot 10^7 \text{ M}^{-1}; K_d = 43 \text{ nM}$$

11mer + 11mer:

$$K_d = 2.37 (\pm 0.33) \cdot 10^7 \text{ M}^{-1}; K_d = 42 \text{ nM}$$

$$\Delta\Delta G = 1.76 \text{ kcal/mol}$$

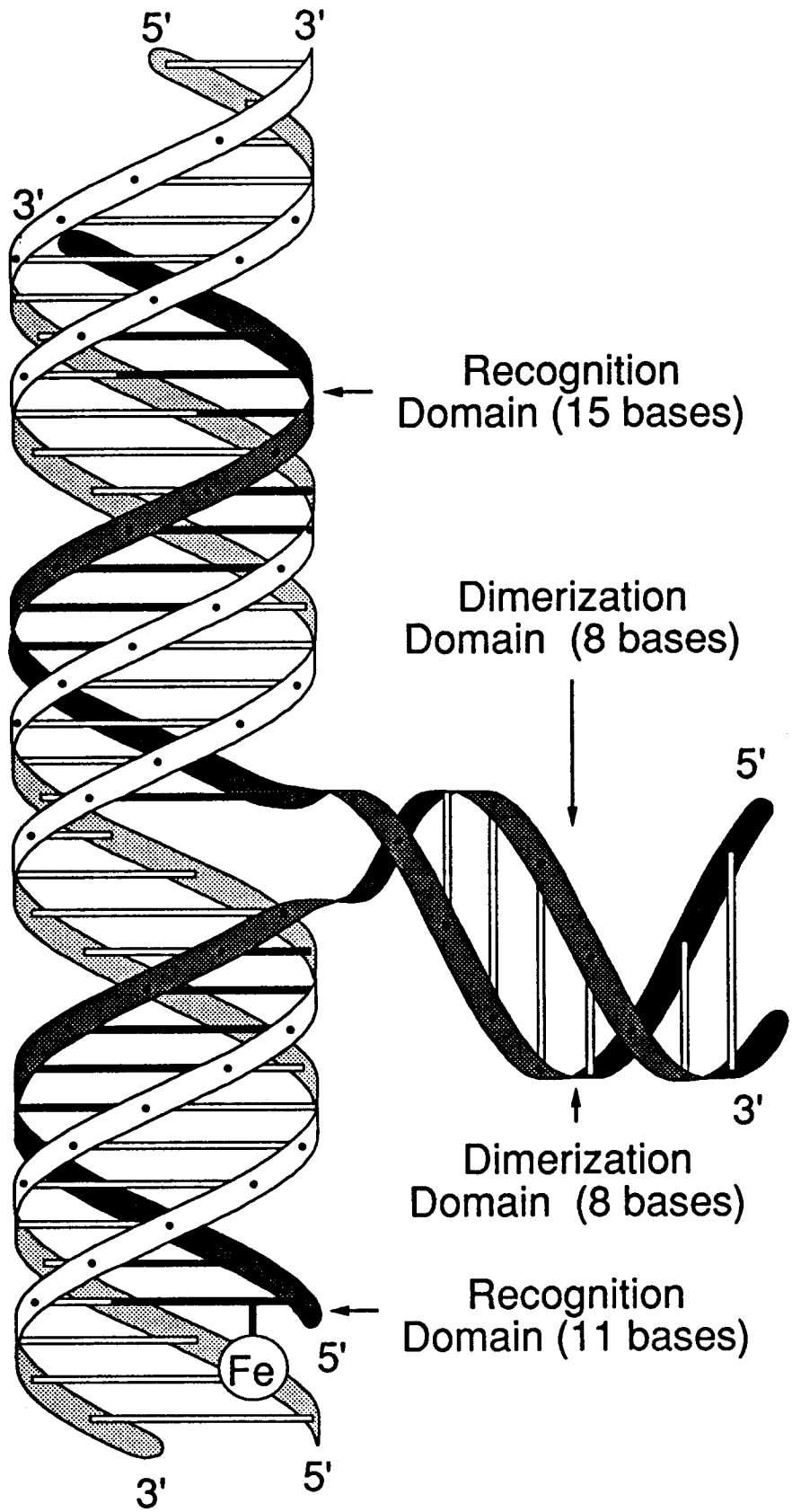
\therefore still 20-fold Enhancement

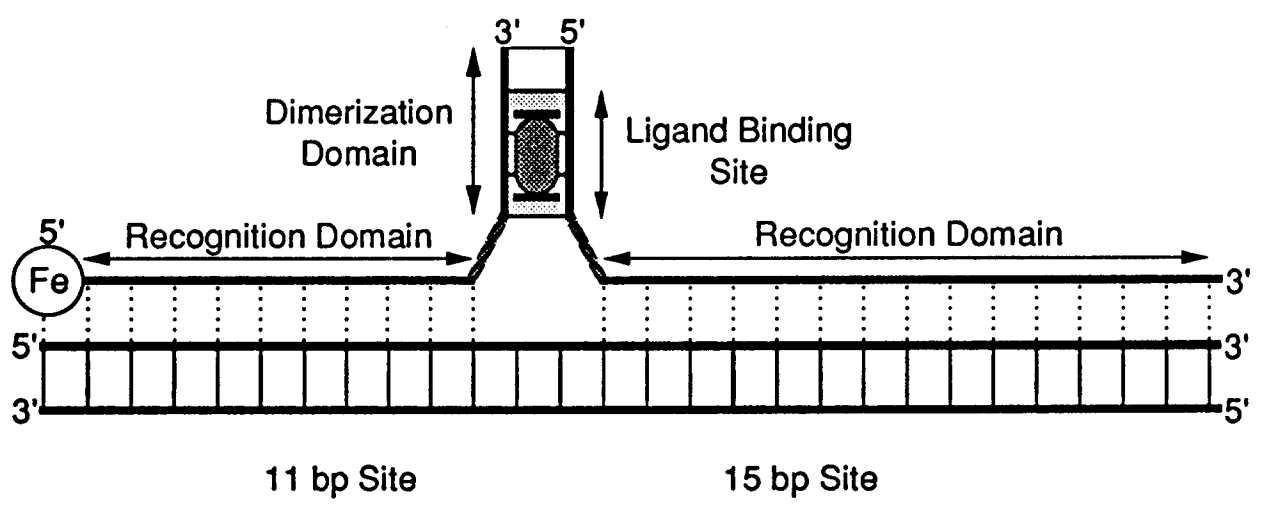
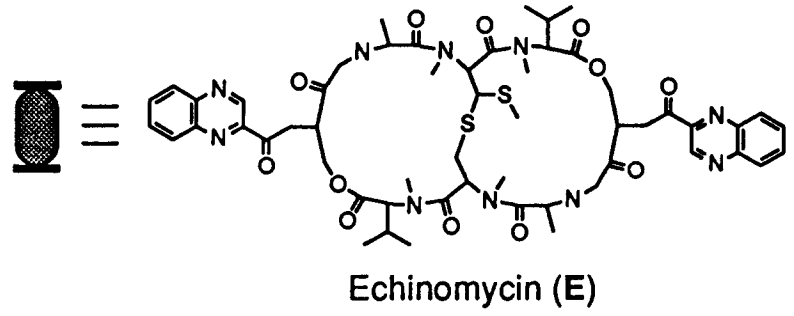
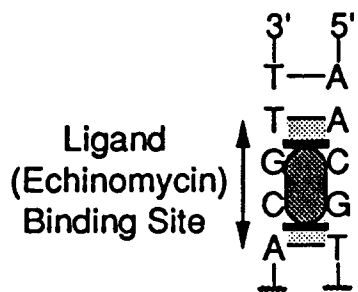


1 5'-^{Me Me}TTTTTTTTTCCTTTCGTC^{Me Me}CAATCG (11 bp Site, T*)

2 5'-CGATTGACCTTTTTCTCTCTCTCT (15 bp Site, No T*)

3 5'-CGATTGACCTTTTTCTCTCTCTCT (15 bp Site, T*)





5'-AAAAAAGGAAAAAAAAAAGAGAGAGAGA-3'
 3'-TTTTTTCCTTTTTTTTTTCTCTCTCTCT-5'

- 1 5'-*TTTTTTTT^{Me Me}CCTTTCACGIT (E Site)
- 2 5'-*TTTTTTTT^{Me Me}CCTTTCCAGIT (No E Site)
- 3 5'-AACGICTTTTT^{Me Me Me Me Me}TCTCTCTCT (E Site)
- 4 5'-AACTGCTTTTT^{Me Me Me Me Me}TCTCTCTCT (No E Site)